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VANILLOID RECEPTOR-RELATED NUCLEIC ACIDS AND POLYPEPTIDES

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the benefit of U.S. Provisional Application No. 60/297,835 filed on June 13, 2001, U.S. Provisional Application No. 60/351,238, filed on January 22, 2002, U.S. Provisional Application No. 60/352,914, filed on January 29, 2002, U.S. Provisional Application No. 60/357,161, filed on February 12, 2002, U.S. Provisional Application No. 60/381,086, filed on May 15, 2002, and U.S. Provisional Application No. 60/381,739, filed on May 16, 2002. These applications are incorporated herein by reference for all purposes.

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BACKGROUND OF THE INVENTION

20 Field of the Invention

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[0003] This invention pertains to novel vanilloid receptor (VR) related nucleic acids and polypeptides. In particular, the invention relates to proteins that are homologous to known VRs, nucleic acids encoding such proteins, identification of trkA⁺ pain-specific genes, and the use of these genes and polypeptides in methods of diagnosing pain, methods of identifying compounds useful in treating pain and methods of treating pain.

Background

[0004] Pain has been defined as the sensory experience perceived by nerve tissue distinct from sensations of touch, pressure, heat and cold. Individuals suffering from pain

typically describe it by such terms as bright, dull, aching, pricking, cutting, burning, etc.

This range of sensations, as well as the variation in perception of pain by different individuals, makes a precise definition of pain difficult. Pain as suffering, however, is generally considered to include both the original sensation and the reaction to that sensation. Where pain results from the stimulation of nociceptive receptors and transmitted over intact neural pathways, this is termed nociceptive pain. Alternatively, pain may be caused by damage to neural structures, often manifesting itself as neural supersensitivity, and is referred to as neuropathic pain.

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[0005] Neuropathic pain is a particular type of pain that has a complex and variable etiology. It is generally a chronic condition attributable to complete or partial transection of a nerve or trauma to a nerve plexus or soft tissue. This condition is characterized by hyperesthesia (enhanced sensitivity to a natural stimulus), hyperalgesia (abnormal sensitivity to pain), allodynia (widespread tenderness, characterized by hypersensitivity to tactile stimuli) and/or spontaneous burning pain. In humans, neuropathic pain tends to be chronic and debilitating, and occurs during conditions such as trigeminal neuralgia, diabetic neuropathy, post-herpetic neuralgia, late-stage cancer, amputation or physical nerve damage.

[0006] Most drugs including conventional opioids and antidepressants are not practical against chronic pain such as neuropathic pain, either because they are not effective or have serious side effects. For these reasons, alternate therapies for the management of chronic or neuropathic pain are widely sought.

[0007] Stimuli such as heat, cold, stretch, and pressure are detected by specialized sensory neurons within the Dorsal Root Ganglia (DRG). These neurons fire action potentials in response to these mechanical and thermal stimuli, although the molecular mechanism for such detection is not known. Recently, two channels, vanilloid receptor 1 (VR1) and vanilloid receptor-like protein 1 (VRL1), have been isolated from DRG that respond to different thresholds of high heat, and hence act as pain receptors. These channels belong to a family of TRP channels that in *C. elegans* and *D. melanogaster* are involved in mechano- and osmoregulation.

[0008] The VR1 is a calcium channel with six transmembrane domains and a putative pore domain. The channel can be activated by many distinct reagents, including heat, low pH (high proton concentration is present during injury and inflammation), and

capsaicin (the active ingredient in hot chili peppers). The knockout of VR1 in mice has demonstrated that this channel plays a role in pain propagation; however, since the phenotype is rather subtle, it also implies that VR1 is not the sole receptor for high heat and pain. To date, one other homologue of VR1 is known in mammals - the VRL1. VRL1 is structurally very similar to VR1, but is expressed on DRG neurons that are not involved in pain reception (in contrast to VR1).

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[0009] The somatic sensory neurons detect external stimuli such as heat, cold and noxious stimuli through the activation of thermal and mechanical receptors/channels. The VR family represents the first example of molecules expressed within the DRG that have such activation capabilities. Since these molecules are relatively specific to sensory neurons (for example, VR1 knockout mice do not have phenotypes outside of pain perception), they represent highly promising targets for developing drugs against pain or other thermal noxious stimuli. VR1 knockout mice have demonstrated that other molecules have to be involved in pain perception. However, despite the large amount of interest generated in the scientific community concerning this class of receptors, so far, no other receptors of this class have been identified.

[0010] In view of the role of the VR members in pain perception, the identification of new members of VR would allow the development of therapeutic candidates specifically designed to block these new TRP channels, which would enable the treatment of various disorders associated with chronic pain. In addition, the identification of new VR members would permit the screening of various drugs to identify those compounds suitable for further, in-depth studies of therapeutic applications.

SUMMARY OF THE INVENTION

[0011] The present invention relates to members of the VR family, in particular TRPV3 (previously known as VRLS, VRLX, VR4 and TRPV7), TRPV4 (previously known as VRL3 and OTRPC4) and TRPM8 (previously known as TRPX) nucleic acids and polypeptides, recombinant materials and methods for their production. In another aspect, the present invention relates to the identification of trkA⁺ pain-specific genes expressed in the DRG. In yet another aspect, the present invention relates to methods for using the TRPV3, TRPV4, TRPM8 and trkA⁺ pain-specific nucleic acids and polypeptides, including methods for treating pain, inflammation, skin disorders and cancer, methods of diagnosing pain,

inflammation, skin disorders and cancer, methods of identifying agents useful in the treatment of pain, inflammation, skin disorders and cancer and in methods of monitoring the efficacy of a treatment for pain, inflammation, skin disorders and cancer.

TRPV3

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[0012] The invention provides isolated and/or purified TRPV3 nucleic acid molecules, such as: a) a polynucleotide that encodes a mouse TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO: 2; b) a polynucleotide that encodes a mouse TRPV3 protein comprising amino acid residues 2-791 of SEO ID NO: 2; c) a polynucleotide that encodes a functional domain of a mouse TRPV3 protein; d) a polynucleotide that encodes a human TRPV3 protein comprising amino acid residues 1-791 of SEO ID NO 5; e) a polynucleotide that encodes a human TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO 5; f) a polynucleotide that encodes a functional domain of a human TRPV3 protein; and g) a polynucleotide that is complementary to a polynucleotide of a) through f). In some embodiments, the nucleic acid molecule is a) or b) and comprises a first polynucleotide that is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 3 (mouse TRPV3), or is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 6 (human TRPV3). The nucleic acids can be 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 3 or SEQ ID NO: 6, or can be identical to the respective polynucleotide. Examples of TRPV3 nucleic acids of the invention include polynucleotides that are 80% or more, 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 65-2440 of SEQ ID NO: 1 (mouse TRPV3) or nucleotides 57-2432 of SEQ ID NO: 4 (human TRPV3).

[0013] The invention also provides isolated TRPV3 nucleic acid molecules that encode polypeptides that include one or more functional domains of a mammalian (e.g., human or mouse) TRPV3 polypeptide. The polypeptides encoded by these nucleic acid molecules can include, for example, one or more functional domains such as ankyrin domains, transmembrane regions, pore loop regions, and coiled-coil domains. As an example, the polypeptides can include a pore loop region flanked by two transmembrane regions, and/or four ankyrin domains.

[0014] Also provided by the invention are isolated and/or purified TRPV3 polypeptides. Such polypeptides include, for example, a) a mouse TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO: 2; b) a mouse TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO: 2; c) one or more functional domains of a mouse TRPV3 protein; d) a human TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO 5; e) a human TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO 5; and f) one or more functional domains of a human TRPV3 protein. For example, the TRPV3 polypeptides can include one or more functional domains selected from the group consisting of an ankyrin domain, a transmembrane region, a pore loop region, and a coiled-coil domain. In some embodiments, the polypeptides include a pore loop region flanked by two transmembrane regions, and/or four ankyrin domains.

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[0015] Methods for identifying an agent that modulates TRPV3-mediated cation passage through a membrane are also provided by the invention. These methods involve: a) providing a membrane that comprises a TRPV3 polypeptide; b) contacting the membrane with a candidate agent; and c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent. In some embodiments, the membrane is a cell membrane and cation passage through the membrane is detected by measuring cation influx or efflux across the membrane into or out of the cell. The assay is conducted at a temperature of at least 33°C, in some embodiments. Also provided are methods in which a candidate agent that reduces cation passage is further tested for ability to treat pain by administering the candidate agent to a test animal and determining whether the candidate agent decreases the test animal's response to a pain stimulus. A pain stimulus can include, for example exposure to a temperature above 33°C.

[0016] The invention also provides methods for reducing pain associated with TRPV3 activity. These methods involve administering to a subject suffering from pain an analgesically effective amount of a compound that reduces TRPV3-mediated cation passage through a membrane or reduces signal transduction from a TRPV3 polypeptide to a DRG neuron. The pain can be with, for example, one or more of heat exposure, inflammation, and tissue damage. Suitable compounds can include, for example, an antibody that specifically binds to a TRPV3 polypeptide; an antisense polynucleotide, ribozyme, or an interfering

RNA that reduces expression of a TRPV3 polypeptide; and/or a chemical compound that reduces cation passage through a membrane that comprises a TRPV3 polypeptide.

[0017] Methods for determining whether pain in a subject is mediated by TRPV3 are also provided by the invention. These methods can involve: obtaining a sample from a region of the subject at which the pain is felt; and testing the sample to determine whether a TRPV3 polypeptide or TRPV3 polynucleotide is present and/or active in the sample. In some embodiments, the presence of a TRPV3 polypeptide in the sample is detected by determining whether cation passage across membranes of cells in the sample is mediated by a TRPV3 polypeptide. For example, TRPV3 involvement in mediating cation passage across membranes of the cells can be determined by detecting an increase in cation passage across membranes of the cells when assayed above 33°C compared to cation passage when assayed below 33°C. To distinguish between TRPV3 involvement in mediating cation passage and involvement by other ion channels (e.g., TRPV1 or TRPV2), the assay can be conducted at a temperature above the activation threshold of TRPV3 but below the activation threshold of the other receptor (e.g., below about 43°C or below about 52°C, respectively, for TRPV1 and TRPV2). As an alternative to assaying for TRPV3-mediated ion channel activity, one can detect the presence of a TRPV3 polypeptide in the sample by contacting the sample with a reagent that specifically binds to a TRPV3 polypeptide, or detect the presence of a TRPV3 polynucleotide in the sample by contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPV3 polynucleotide.

TRPV4

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[0018] The invention also provides isolated TRPV4 nucleic acid molecules. These include, for example, a) a polynucleotide that encodes a mouse TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO: 14; b) a polynucleotide that encodes a mouse TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO: 14; c) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a mouse TRPV4 protein; d) a polynucleotide that encodes a human TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO 17; e) a polynucleotide that encodes a human TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO 17; f) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a human TRPV4 protein; and g) a polynucleotide that is complementary to a polynucleotide

of a) through f). In some embodiments, the nucleic acid molecule is a) or b) and comprises a first polynucleotide that is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 15 (mouse TRPV4), or is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 18 (human TRPV4). The nucleic acids can be 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 15 or SEQ ID NO: 18, or can be identical to the respective polynucleotide. Examples of TRPV4 nucleic acids of the invention include polynucleotides that are 80% or more, 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 156-2771 of SEQ ID NO: 13 (mouse TRPV4) or to a nucleotide sequence as set forth in SEQ ID NO: 16 (human TRPV4).

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[0019] The invention also provides isolated TRPV4 nucleic acid molecules that encode polypeptides that include one or more functional domains of a mammalian (e.g., human or mouse) TRPV4 polypeptide. The polypeptides encoded by these nucleic acid molecules can include, for example, one or more functional domains such as ankyrin domains, transmembrane regions, pore loop regions, and coiled-coil domains. As an example, the polypeptides can include a pore loop region flanked by two transmembrane regions, and/or three ankyrin domains.

[0020] Also provided by the invention are isolated and/or purified TRPV4 polypeptides. Such polypeptides include, for example, a) a mouse TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO: 14; b) a mouse TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO: 14; c) one or more functional domains of a mouse TRPV4 protein; d) a human TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO 17; e) a human TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO 17; and f) one or more functional domains of a human TRPV4 protein. For example, the TRPV4 polypeptides can include one or more functional domains selected from the group consisting of an ankyrin domain, a transmembrane region, a pore loop region, and a coiled-coil domain. In some embodiments, the polypeptides include a pore loop region flanked by two transmembrane regions, and/or three ankyrin domains.

[0021] Methods for identifying an agent that modulates TRPV4-mediated cation passage through a membrane are also provided by the invention. These methods involve: a)

providing a membrane that comprises a TRPV4 polypeptide; b) contacting the membrane with a candidate agent; and c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent. Cation influx and/or efflux can be measured as described above for TRPV3. In some embodiments, candidate agents that reduce cation passage are further tested for ability to treat pain by administering the candidate agent to a test animal and determining whether the candidate agent decreases the test animal's response to a pain stimulus.

[0022] Methods for reducing pain associated with TRPV4 activity are provided by the invention. These methods involve administering to a subject suffering from pain an analgesically effective amount of a compound that reduces TRPV4-mediated cation passage through a membrane or reduces signal transduction from a TRPV4 polypeptide to a DRG neuron. The compounds are suitable for treating, for example, neuropathic pain, and can include: a) an antibody that specifically binds to a TRPV4 polypeptide; b) an antisense polynucleotide, ribozyme, or an interfering RNA that reduces expression of a TRPV4 polypeptide; and c) a chemical compound that reduces cation passage through a membrane that comprises a TRPV4 polypeptide.

[0023] The invention also provides methods for determining whether pain in a subject is mediated by TRPV4. These methods involve obtaining a sample from a region of the subject at which the pain is felt, and testing the sample to determine whether a TRPV4 polypeptide or TRPV4 polynucleotide is present and/or active in the sample. The presence and/or activity of the TRPV4 polypeptide can be detected, for example, by determining whether cation passage across membranes of cells in the sample is mediated by a TRPV4 polypeptide, or by contacting the sample with a reagent that specifically binds to a TRPV4 polypeptide. One can detect the presence of a TRPV4 polynucleotide by, for example, contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPV4 polynucleotide.

TRPM8

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[0024] Isolated and/or purified TRPM8 nucleic acid molecules are also provided by the invention. These TRPM8 nucleic acid molecules include, for example, a) a polynucleotide that encodes a mouse TRPM8 protein comprising amino acid residues 1-1104

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of SEQ ID NO: 8; b) a polynucleotide that encodes a mouse TRPM8 protein comprising amino acid residues 2-1104 of SEQ ID NO: 8; c) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a mouse TRPM8 protein; d) a polynucleotide that encodes a human TRPM8 protein comprising amino acid residues 1-1268 of SEQ ID NO 11; e) a polynucleotide that encodes a human TRPM8 protein comprising amino acid residues 2-1268 of SEQ ID NO 11; f) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a human TRPM8 protein; and g) a polynucleotide that is complementary to a polynucleotide of a) through f). In some embodiments, the nucleic acid molecule is a) or b) and comprises a first polynucleotide that is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 9 (mouse TRPM8), or is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 12 (human TRPM8). The nucleic acids can be 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in SEO ID NO: 9 or SEQ ID NO: 12, or can be identical to the respective polynucleotide. Examples of TRPM8 nucleic acids of the invention include polynucleotides that are 80% or more, 90% or more, or 95% or more, identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7 (mouse TRPM8) or nucleotides 61-4821 of SEQ ID NO: 10 (human TRPM8).

[0025] The invention also provides isolated TRPM8 nucleic acid molecules that encode polypeptides that include one or more functional domains of a mammalian (e.g., human or mouse) TRPM8 polypeptide. The polypeptides encoded by these nucleic acid molecules can include, for example, one or more functional domains such as transmembrane regions, pore loop regions, and coiled-coil domains. As an example, the polypeptides can include a pore loop region flanked by two transmembrane regions.

[0026] The invention also provides isolated and/or purified TRPM8 polypeptides. The TRPM8 polypeptides include, for example, a) a mouse TRPM8 protein comprising amino acid residues 1-1104 of SEQ ID NO: 8; b) a mouse TRPM8 protein comprising amino acid residues 2-1104 of SEQ ID NO: 8; c) one or more functional domains of a mouse TRPM8 protein; d) a human TRPM8 protein comprising amino acid residues 1-1268 of SEQ ID NO 11; e) a human TRPM8 protein comprising amino acid residues 2-1268 of SEQ ID NO 11; and f) one or more functional domains of a human TRPM8 protein. For example, the

TRPM8 polypeptides can include one or more functional domains selected from the group consisting of a transmembrane region, a pore loop region, and a coiled-coil domain. In some embodiments, the TRPM8 polypeptides of the invention include a pore loop region flanked by two transmembrane regions.

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[0027] Methods for identifying an agent that modulates TRPM8-mediated cation passage through a membrane are also provided by the invention. These methods involve: a) providing a membrane that comprises a TRPM8 polypeptide; b) contacting the membrane with a candidate agent; and c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent. In some embodiments, the membrane is a cell membrane and cation passage through the membrane is detected by measuring cation influx or efflux across the membrane into or out of the cell. To identify antagonists that reduce TRPM8-mediated cation passage, the assay typically is conducted under conditions in which TRPM8 allows cation passage in the absence of the antagonist; e.g., at a temperature of about 20°C or less, or in the presence of menthol. Also provided are methods in which a candidate agent that reduces cation passage is further tested for ability to treat pain by administering the candidate agent to a test animal and determining whether the candidate agent decreases the test animal's response to a pain stimulus. A pain stimulus can include, for example exposure to a temperature below 20°C.

[0028] In other embodiments, the invention provides methods for identifying an agent that stimulates TRPM8-mediated cation passage through a membrane. These screens for identifying TRPM8 agonists generally are conducted under conditions in which the TRPM8 polypeptides do not mediate cation passage. Such conditions include, for example, temperatures above about 20°C. Agonists of TRPM8-mediated cation passage are useful as flavor enhancers, fragrances, and the like.

[0029] The invention also provides methods of reducing pain associated with TRPM8 activity. These methods involve administering to a subject suffering from pain an analgesically effective amount of a compound that reduces TRPM8-mediated cation passage through a membrane or reduces signal transduction from a TRPM8 polypeptide to a DRG neuron. These methods are useful for treating pain that results from, for example, cold exposure, inflammation, tissue damage, and the like. The compounds can be, for example, a) an antibody that specifically binds to a TRPM8 polypeptide; b) an antisense polynucleotide,

ribozyme, or an interfering RNA that reduces expression of a TRPM8 polypeptide; or c) a chemical compound that reduces cation passage through a membrane that comprises a TRPM8 polypeptide.

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[0030] Methods for determining whether pain in a subject is mediated by TRPM8 are also provided by the invention. These methods involve obtaining a sample from a region of the subject at which the pain is felt; and testing the sample to determine whether a TRPM8 polypeptide or TRPM8 polypucleotide is present and/or active in the sample. In some embodiments, the presence of a TRPM8 polypeptide in the sample is detected by determining whether cation passage across membranes of cells in the sample is mediated by a TRPM8 polypeptide. TRPM8 involvement in mediating cation passage across membranes of the cells can be determined, for example, by detecting an increase or decrease in cation passage across membranes of the cells when assayed below 20°C and/or in the presence of menthol, compared to cation passage when assayed above 20°C and/or in the absence of menthol. Alternatively, or additionally, the presence of a TRPM8 polypeptide in the sample is detected by contacting the sample with a reagent that specifically binds to a TRPM8 polypeptide. The presence of a TRPM8 polynucleotide in the sample can be detected by, for example, contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPM8 polynucleotide.

[0031] The invention also provides methods for identifying an agent useful in the modulation of a mammalian sensory response. These methods involve: a) contacting a candidate agent with a test system that comprises a receptor polypeptide selected from the group consisting of TRPM8, TRPV3 and TRPV4; and b) detecting a change in activity of the receptor polypeptide in the presence of the candidate agent as compared to the activity of the receptor polypeptide in the absence of the agent, thereby identifying an agent that modulates receptor activity.

[0032] Also provided by the invention are methods for monitoring the efficacy of a treatment of a subject suffering from pain. These methods involve: a) obtaining, at two or more time points in the course of treatment for pain, a sample from a region of the subject at which the pain is felt; and b) testing the samples to determine whether a reduction is observed in amount or activity of one or more members selected from the group consisting of: a TRPV3 polypeptide, a TRPV3 mRNA, a TRPV4 polypeptide, a TRPV4 mRNA, a TRPV4 polypeptide, and a TRPM8 mRNA. In some embodiments, one of the time points is

prior to or simultaneously with administration of the treatment, and the other time point is after treatment has begun.

[0033] The invention provides assays capable of detecting the expression of one or more of TRPV3, TRPV4 or TRPM8 in human tissue. The assays are selected from the group consisting of: a) an assay comprising contacting a human tissue sample with monoclonal antibodies binding to TRPV3, TRPV4 or TRPM8 and determining whether the monoclonal antibodies bind to polypeptides in the sample; and b) an assay comprising contacting a human tissue sample with an oligonucleotide that is capable of hybridizing to a nucleic acid that encodes TRPV3, TRPV4 or TRPM8.

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[0034] Methods of treating pain provided by the invention include methods in which a patient suffering from pain mediated by one or more polypeptides selected from the group consisting of TRPV3, TRPV4 and TRPM8 is identified by measuring expression of the polypeptide in tissue from such patient, and administering to such patient an analgesically effective amount of an agent which inhibits the polypeptide.

[0035] The invention also provides methods for identifying an agent useful in the treatment of pain. These methods involve: a) administering a candidate agent to a mammal suffering from pain; b) in a sample obtained from the mammal, detecting an activity or amount of one or more members selected from the group consisting of: a TRPV3 polypeptide, a TRPV3 mRNA, a TRPV4 polypeptide, a TRPV4 mRNA, a TRPM8 polypeptide, and a TRPM8 mRNA; and c) comparing the amount or activity of the member in the presence of the candidate agent with the amount or activity of the member in a sample obtained from the mammal in the absence of the candidate agent, wherein a decrease in amount or activity of the member in the sample in the presence of the candidate agent relative to the amount or activity in the absence of the candidate agent is indicative of an agent useful in the treatment of pain.

[0036] Also provided are methods for identifying an agent that binds to and/or modulates the activity of an mRNA or polypeptide encoded by a TRPV3, TRPV4, or TRPM8 nucleic acid. These methods involve: a) contacting an isolated cell which expresses a heterologous TRPV3, TRPV4, or TRPM8 nucleic acid encoding a polypeptide with the agent; and b) determining binding and/or modulation of the activity of the mRNA or polypeptide by the agent, to identify agents which bind with and/or modulate the activity of the polypeptide.

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BRIEF DESCRIPTION OF THE DRAWINGS

[0037] Figures 1A and 1B show differential expression of TRPV3 and TRPV4 genes in the Chung model. Figure 1A: mRNA levels of TRPV3 are increased in a rat model of chronic neuropathic pain. The human cDNA sequence of TRPV3 is used to search the Celera mouse genomic DNA database and two primers are derived from regions that are identical from human and mouse sequences. The primers are used to amplify the rat TRPV3 from total RNA samples from the Chung model (L4 and L5 DRG) and sham-operated animals in a standard reverse-transcriptase polymerase chain reaction (RT-PCR) protocol. The top panel shows the gel image from one RT-PCR experiment and the bottom shows the average fold of regulation of TRPV3 in L4 and L5 DRG neurons from Chung model from three independent experiments. Figure 1B: TRPV4 is up-regulated in a rat model of chronic neuropathic pain. For analysis TRPV4 expression in the Chung model (28- and 50-day). first-strand cDNA equivalent to 30 ng of total RNA is used per reaction and amplified between 32/35 cycles for higher expressing genes and 35/38 cycles for lower-expressing genes. Due to the constraints on the amount of total RNA available, half the volume of the PCR reaction is removed at the lower cycle and the remaining reaction is continued for a further 3 cycles. All the samples are resolved on 4-20% TBE gels and densitometry carried out on the clearest, non-saturated bands.

[0038] Figures 2A-2F show the TRPV3 sequence and genomic localization.

Figure 2A: Rooted tree showing protein sequence relationship of different members of the TRPV ion channel family. Figure 2B: Relative position of TRPV1 (VR1) and TRPV3 coding sequences on mouse (11B4) and human (17p13) chromosomes. Figure 2C: Comparison of mouse TRPV3 protein sequence to other TRPVs (excluding C-terminal half containing transmembrane domains). Identical sequences are highlighted in dark gray; conserved residues, in light gray. Predicted coiled-coil and ankyrin domains are marked and correspond to regions for TRPV3 only. The protein alignment is generated using Megalign and Boxshade at http://biowb.sdsc.edu/CGI/BW.cgi. The coiled-coil domains are predicted using the program Coils (http://searchlauncher.bcm.tmc.edu/seq-search/struc-predict.html). The ankyrin domains are predicted using the PFAM protein search

30 (http://pfam.wustl.edu/hmmsearch.shtml). Figure 2D: A schematic of TRPV3 and predicted membrane topology. Figure 2E: Kyte Doolittle hydrophobicity plot of TRPV3 sequences showing the 6 transmembrane domains (1-6) and the pore domain (P). Figure 2F: Coiled-

coil domain prediction of TRPV3 sequence by Coils shows two 14-mer peaks at the N-terminal, prior to ankyrin sequences.

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[0039] Figures 3A-3D demonstrate that TRPV3 is activated by heat. Currents evoked by heat in TRPV3 expressing Chinese Hamster Ovary (CHO) cells. Figure 3A: Inward current to temperature ramp, $V_h = -60$ mV, in calcium free external solutions. Figure 3B: Heat evoked currents of the same cell in Ca^{2+} -free and subsequently in Ca^{2+} containing solutions showing increased inward current in Ca^{2+} conditions. Figure 3C: Semi-logarithmic plot of current against temperature with double exponential fitted line for the same trace as Figure 3A. Note the discontinuity at ~32°C (arrow). Figure 3D: Current-voltage relationship in calcium containing external solution showing the pronounced outward rectification of TRPV3 at 48°C but not at room temperature. Note the small outward currents at room temperature.

[0040] Figures 4A-4D. TRPV3 becomes sensitized to repeated applications of the heat stimulus. Figure 4A: Repeated heat steps from 25-45°C evoke increased inward current responses. Figure 4B: The outward rectification becomes more pronounced with repeated voltage ramps in 48°C external solution. Both experiments are conducted in the presence of 2 mM CaCl₂ in the external solution. Figure 4C: Control protocol for antagonist experiments. Note that the responses continue to sensitize with repeated heat steps in the absence of putative antagonists. Figure 4D: 1 μM ruthenium red attenuates the sensitization and inhibits the heat response.

[0041] Figure 5. TRP Channels in thermosensation. Four TRP channels implicated in thermosensation cover most but not all physiologically relevant temperatures.

[0042] Figures 6A-6D show results of an analysis of the nucleotide and amino acid sequences of TRPM8. Figure 6A: Comparison of mouse TRPM8 protein sequence to some of its closest relatives, TRPM1 (human Melastatin, GI 6006023), TRPM2 (human, GI 4507688) and TRPM7 (mouse Chak, GI 14211382). The alignment is generated using Megalign and Boxshade. Identical or conserved residues are shown in white letters on a black background. Figure 6B: Phylogenetic tree showing protein sequence relationship of different members of the TRP ion channel super-family. TRPs are subdivided into three main subfamilies: TRPMs, TRPVs and TRPCs. The TRPMs do not contain any Ankyrin domains in their N-terminal domains. The transmembrane domains have the highest homology among different classes of TRP channels. Figure 6C: Kyte Doolittle

hydrophobicity plot of TRPM8 sequences showing the eight hydrophobic peaks demarking the potential transmembrane regions of the protein that spans from 695-1024 amino acids. Figure 6D: Coiled-coil domain prediction of TRPM8 sequence by the program coils shows multiple 14-mer peaks at the N- and C-terminus of the transmembrane spanning domains (http://searchlauncher.bcm.tmc.edu/seq-search/struc-predict.html).

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[0043] Figures 7A-7E: Increase in intracellular calcium concentration ([Ca²⁺];) in TRPM8-expressing CHO cells in response to cold and menthol. Figure 7A: mTRPM8 CHO cells show a rapid increase in $[Ca^{2+}]_i$ when the temperature reaches ~15°C. Non-transfected CHO cells do not show a response to cold. Removal of external Ca²⁺ completely abolishes the response to cooling. Figure 7B: The estimated average threshold temperature at which [Ca²⁺]_i begins to increase is approximately 23°C for mTRPM8. TRPM8-expressing CHO cells are cooled from 33-23°C, upon which an increase in Ca²⁺ is observed. Continuous cooling of the cells to 20°C shows a marked Ca²⁺ increase followed by a rapid return to nearbasal levels upon warming to 33°C. Figure 7C: TRPM8 responses, evoked by repeated applications of a 23°C temperature stimulus show little desensitization in calcium-containing standard bath solution. Figure 7D: TRPM8 responds to menthol at 25°C. Intensity of the TRPM8 response is dependent on menthol concentrations. A 10-fold increase in menthol concentration results in a larger influx of Ca²⁺. This response is suppressed in the absence of extracellular Ca²⁺. Non-transfected CHO cells exhibit no increase in [Ca²⁺]; upon application of menthol. Figure 7E: At 33°C, 10 µM menthol does not elicit an influx of Ca²⁺. When the temperature of the bath solution is lowered to 30°C, a marked increase in intracellular Ca²⁺ is observed. Additionally, repeated applications of menthol do not appear to desensitize TRPM8-expressing cells. These experiments suggest that menthol simulates the effect of cooling in TRPM8-expressing cells. This identification of a cold-sensing TRP channel involved in thermoreception reveals an expanded role for this family in somatic sensory detection.

[0044] Figures 8A-8B show an increase in intracellular calcium concentration [Ca²⁺]_i in TRPM8-expressing CHO cells in response to cold. Figure 8A: TRPM8-transfected CHO cells show a rapid increase in [Ca²⁺]_i when the temperature is lowered from 25°C to 15°C. The stimulus period is indicated below the traces. Non-transfected CHO cells do not show a response to cold. Removal of external Ca²⁺ completely suppresses the response to cooling. Experiments are performed in triplicate. The average response (± SEM) of 20-30

cells from a representative experiment is presented. Figure 8B: Increase in $[Ca^{2+}]_i$ due to decrease in temperature from 35°C to 13°C in TRPM8⁺ cells. The panel shows mean \pm SEM for 34 individual cells. Note the increase starts to occur between 22°C and 25°C.

[0045] Figures 9A-9B show that current is evoked by reduction in temperature in TRPM8-expressing CHO cells. Figure 9A: Outward currents evoked at +60 mV by reducing the temperature from 35°C to 10°C. In this cell the current activates at 19.3°C as indicated in the right hand panel. Figure 9B: Current-voltage relationship for currents activated at 20.5°C and 33.5°C. Increasing the temperature reduces the amplitude of outward currents.

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[0046] Figures 10A-10B show that current is evoked by menthol in TRPM8-expressing CHO cells. Figure 10A: Inward currents evoked by 1 mM menthol ($V_h = -60$ mV) are inactivated by increasing the temperature from 25°C to 45°C. Figure 10B: Current-voltage relationship for response to 1 mM menthol. Currents show pronounced outward-rectification in the presence of menthol not seen in the absence of this agonist.

[0047] Figures 11A-11B show a dose-response curve for menthol-stimulated current in TRPM8-expressing CHO cells. The voltage employed was +60 mV. Figure 11A: Single examples, from two different cells, of current evoked by applying 0.1, 0.5, 1 and 10 mM menthol at 22°C and 35°C. Figure 11B: Comparison of response (mean ± SEM, n=5 for all points) of current evoked by menthol either at 22°C or 35°C.

DESCRIPTION OF THE SEQUENCE LISTING

20 [0048] SEQ ID NO: 1 provides a nucleotide sequence that encodes a mouse TRPV3 polypeptide, and upstream and downstream regions. The open-reading frame extends from nucleotides 65-2440.

[0049] SEQ ID NO: 2 provides an amino acid sequence of a mouse TRPV3 polypeptide.

[0050] SEQ ID NO: 3 provides nucleotide sequences for all polynucleotides that code for the mouse TRPV3 amino acid sequence presented in SEQ ID NO: 2.

[0051] SEQ ID NO: 4 provides a nucleotide sequence that encodes a human TRPV3 polypeptide, and an upstream non-coding region. The open-reading frame extends from nucleotides 57-2432.

[0052] SEQ ID NO: 5 provides an amino acid sequence of a human TRPV3 polypeptide.

[0053] SEQ ID NO: 6 provides nucleotide sequences for all polynucleotides that code for the human TRPV3 amino acid sequence presented in SEQ ID NO: 5.

[0054] SEQ ID NO: 7 provides a nucleotide sequence that encodes a mouse TRPM8 polypeptide, and upstream and downstream non-coding regions. The coding region extends from nucleotides 448-3762.

[0055] SEQ ID NO: 8 provides an amino acid sequence of a mouse TRPM8 polypeptide.

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[0056] SEQ ID NO: 9 provides nucleotide sequences for all polynucleotides that code for the mouse TRPM8 amino acid sequence presented in SEQ ID NO: 8.

[0057] SEQ ID NO: 10 provides a nucleotide sequence that encodes a human TRPM8 polypeptide, and upstream and downstream non-coding regions. The coding region extends from nucleotides 61-4821.

[0058] SEQ ID NO: 11 provides an amino acid sequence of a human TRPM8 polypeptide.

[0059] SEQ ID NO: 12 provides nucleotide sequences for all polynucleotides that code for the human TRPM8 amino acid sequence presented in SEQ ID NO: 11.

[0060] SEQ ID NO: 13 provides a nucleotide sequence that encodes a mouse TRPV4 polypeptide, and upstream and downstream regions. The open-reading frame extends from nucleotides 156-2771.

[0061] SEQ ID NO: 14 provides an amino acid sequence of a mouse TRPV4 polypeptide.

[0062] SEQ ID NO: 15 provides nucleotide sequences for all polynucleotides that code for the mouse TRPV4 amino acid sequence presented in SEQ ID NO: 14.

[0063] SEQ ID NO: 16 provides a nucleotide sequence that encodes a human TRPV4 polypeptide.

[0064] SEQ ID NO: 17 provides an amino acid sequence of a human TRPV4 polypeptide.

[0065] SEQ ID NO: 18 provides nucleotide sequences for all polynucleotides that code for the human TRPV4 amino acid sequence presented in SEQ ID NO: 17.

DETAILED DESCRIPTION

Definitions

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[0066] A "host cell," as used herein, refers to a prokaryotic or eukaryotic cell that contains heterologous DNA that has been introduced into the cell by any means, e.g., electroporation, calcium phosphate precipitation, microinjection, transformation, viral infection and the like.

[0067] "Heterologous" as used herein means "of different natural origin" or represent a non-natural state. For example, if a host cell is transformed with a DNA or gene derived from another organism, particularly from another species, that gene is heterologous with respect to that host cell and also with respect to descendants of the host cell which carry that gene. Similarly, heterologous refers to a nucleotide sequence derived from and inserted into the same natural, original cell type, but which is present in a non-natural state, e.g., a different copy number, or under the control of different regulatory elements.

[0068] A "vector" molecule is a nucleic acid molecule into which heterologous nucleic acid may be inserted which can then be introduced into an appropriate host cell. Vectors preferably have one or more origins of replication, and one or more sites into which the recombinant DNA can be inserted. Vectors often have convenient means by which cells with vectors can be selected from those without, e.g., they encode drug resistance genes. Common vectors include plasmids, viral genomes, and (primarily in yeast and bacteria) "artificial chromosomes".

[0069] "Plasmids" generally are designated herein by a lower case p preceded and/or followed by capital letters and/or numbers, in accordance with standard naming conventions that are familiar to those of skill in the art. Starting plasmids disclosed herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids by routine application of well-known, published procedures. Many plasmids and other cloning and expression vectors that can be used in accordance with the present invention are well-known and readily available to those of skill in the art. Moreover, those of skill readily may construct any number of other plasmids suitable for use in the invention. The properties, construction and use of such plasmids, as well as other vectors, in the present invention will be readily apparent to those of skill from the present disclosure.

[0070] The terms "nucleic acid", "DNA sequence" or "polynucleotide" refer to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues of natural nucleotides that hybridize to nucleic acids in manner similar to naturally-occurring nucleotides. Although polynucleotide sequences presented herein recite "T" (for thymidine), which is found only in DNA, the sequences also encompass the corresponding RNA molecules in which each "T" in the DNA sequence is replaced by "U" for uridine.

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[0071] The term "isolated" refers to material that is substantially or essentially free from components which normally accompany the material as found in its native state. Thus, the polypeptides and nucleic acids of the invention do not include materials normally associated with their in situ environment. An isolated nucleic acid, for example, is not associated with all or part of the chromosomal DNA that would otherwise flank the nucleic acid. Typically, isolated proteins of the invention are at least about 80% pure, usually at least about 90%, and preferably at least about 95% pure as measured by band intensity on a silver stained gel or other method for determining purity. Protein purity or homogeneity can be indicated by a number of means well-known in the art, such as polyacrylamide gel electrophoresis of a protein sample, followed by visualization upon staining. For certain purposes high resolution will be needed and HPLC or a similar means for purification utilized.

[0072] The terms "identical" or percent "identity", in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same, when compared and aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection.

[0073] The phrase "substantially identical", in the context of two nucleic acids or polypeptides, refers to two or more sequences or subsequences that have at least 70%, preferably 80%, most preferably 90-95% nucleotide or amino acid residue identity, when compared and aligned for maximum correspondence, as measured using one of the following sequence comparison algorithms or by visual inspection. Preferably, the substantial identity exists over a region of the sequences that is at least about 50 residues in length, more preferably over a region of at least about 100 residues, and most preferably the sequences are

substantially identical over at least about 150 residues. In a most preferred embodiment, the sequences are substantially identical over the entire length of the coding regions.

[0074] For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

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[0075] Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, Adv. Appl. Math., 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, J. Mol. Biol., 48:443 (1970), by the search for similarity method of Pearson & Lipman, Proc. Natl. Acad. Sci. USA, 85:2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, WI), or by visual inspection (see generally, Current Protocols in Molecular Biology, F.M. Ausubel et al., eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (1995 Supplement) (Ausubel)).

[0076] Examples of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al., *J. Mol. Biol.*, 215:403-410 (1990) and Altschul et al., *Nucleic Acids Res.*, 25:3389-3402 (1977), respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (http://www.ncbi.nlm.nih.gov/). This algorithm involves first identifying high-scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for

mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters wordlength (W), T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a W of 11, an expectation (E) of 10, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a W of 3, an E of 10 and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, *Proc. Natl. Acad. Sci. USA*, 89:10915 (1989)). Percent identities, where specified herein, are typically calculated using the Blast 2.0 implementation using the default parameters.

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[0077] In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul, *Proc. Natl. Acad. Sci. USA*, 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

[0078] Another indication that two polynucleotides are substantially identical is that the polynucleotides hybridize to each other under specified hybridization conditions. Examples of stringent hybridization conditions include: incubation temperatures of about 25°C to about 37°C; hybridization buffer concentrations of about 6 x SSC to about 10 x SSC; formamide concentrations of about 0% to about 25%; and wash solutions of about 6 x SSC. Examples of moderate hybridization conditions include: incubation temperatures of about 40°C to about 50°C; buffer concentrations of about 9 x SSC to about 2 x SSC; formamide concentrations of about 30% to about 50%; and wash solutions of about 5 x SSC to about 2 x SSC. Examples of high stringency conditions include: incubation temperatures of about 55°C to about 68°C; buffer concentrations of about 1 x SSC to about 0.1 x SSC; formamide concentrations of about 55% to about 75%; and wash solutions of about 1 x SSC,

0.1 x SSC or deionized water. In general, hybridization incubation times are from 5 minutes to 24 hours, with 1, 2 or more washing steps, and wash incubation times are about 1, 2 or 15 minutes. SSC is 0.15 M NaC1 and 15 mM citrate buffer. It is understood that equivalents of SSC using other buffer systems can be employed.

[0079] A further indication that two nucleic acid sequences or polypeptides are substantially identical is that the polypeptide encoded by the first nucleic acid is immunologically cross-reactive with the polypeptide encoded by the second nucleic acid, as described below. Thus, a polypeptide is typically substantially identical to a second polypeptide, for example, where the two peptides differ only by conservative substitutions. Another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions, as described below.

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[0080] "Conservatively modified variations" of a particular polynucleotide sequence refers to those polynucleotides that encode identical or essentially identical amino acid sequences, or where the polynucleotide does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given polypeptide. For instance, the codons CGU, CGC, CGA, CGG, AGA and AGG all encode the amino acid arginine. Thus, at every position where an arginine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations," which are one species of "conservatively modified variations". Every polynucleotide sequence described herein which encodes a polypeptide also describes every possible silent variation, except where otherwise noted. One of skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine) can be modified to yield a functionally identical molecule by standard techniques. Accordingly, each "silent variation" of a nucleic acid which encodes a polypeptide is implicit in each described sequence.

[0081] Furthermore, one of skill will recognize that individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are "conservatively modified variations" where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art (see, e.g., Creighton, *Proteins*,

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W.H. Freeman and Company (1984)). Individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids in an encoded sequence are also "conservatively modified variations".

[0082] The term "recombinant" when used with reference to a cell, or nucleic acid, or vector, indicates that the cell, or nucleic acid, or vector, has been modified by the introduction of a heterologous nucleic acid or the alteration of a native nucleic acid, or that the cell is derived from a cell so modified. Thus, for example, recombinant cells can contain genes that are not found within the native (non-recombinant) form of the cell or can express native genes that are otherwise abnormally expressed, under expressed or not expressed at all. Recombinant cells can also contain genes found in the native form of the cell wherein the genes are modified and re-introduced into the cell by artificial means. The term also encompasses cells that contain a nucleic acid endogenous to the cell that has been modified without removing the nucleic acid from the cell; such modifications include those obtained by gene replacement, site-specific mutation and related techniques.

[0083] The term "modulate" refers to a change in the activity and/or amount of TRPV3, TRPV4 or TRPM8 proteins. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional or immunological properties of such proteins. The term "modulation" also refers to a change in the increase or decrease in the level of expression of mRNA or protein encoded by the TRPV3, TRPV4, and TRPM8 genes.

[0084] The term "operably-linked", as used herein, refer to functionally-related nucleic acid sequences. A promoter is operably associated or operably-linked with a coding sequence if the promoter controls the translation of the encoded polypeptide. While operably-linked nucleic acid sequences can be contiguous and in the same reading frame, certain genetic elements, e.g., repressor genes, are not contiguously linked to the sequence encoding the polypeptide but still bind to operator sequences that control expression of the polypeptide.

[0085] The term "agonist", as used herein, refers to a molecule which, when bound to the TRPV3, TRPV4 and TRPM8 proteins, increases or prolongs the duration of the effect of the biological or immunological activity of such proteins. Agonists may include proteins, nucleic acids, carbohydrates or any other molecules which bind to and modulate the effect of these proteins.

[0086] The term "antagonist", as used herein, refers to a molecule which, when bound to TRPV3, TRPV4 and TRPM8 proteins, decreases the amount or the duration of the effect of the biological or immunological activity of these proteins. Antagonists may include proteins, nucleic acids, carbohydrates, antibodies or any other molecules which decrease the effect of these proteins. The term "antagonist" can also refer to a molecule which decreases the level of expression of mRNA and/or translation of protein encoded by TRPV3, TRPV4, and TRPM8 genes. Examples of such antagonists include antisense polynucleotides, ribozymes and double-stranded RNAs.

[0087] In practicing the present invention, many conventional techniques in 10 molecular biology, microbiology and recombinant DNA are used. These techniques are well-known and are explained in, e.g., Current Protocols in Molecular Biology, Vols. I, II and III, F.M. Ausubel, ed. (1997); Sambrook et al., Molecular Cloning: A Laboratory Manual, 3rd Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (2001); DNA Cloning: A Practical Approach, Vols. I and II, D.N. Glover, ed. (1985); 15 Oligonucleotide Synthesis, M.L. Gait, ed. (1984); Nucleic Acid Hybridization, Hames and Higgins (1985); Transcription and Translation, Hames and Higgins, eds. (1984); Animal Cell Culture, R.I. Freshney, ed. (1986); Immobilized Cells and Enzymes, IRL Press (1986); Perbal, A Practical Guide to Molecular Cloning; the series, Methods in Enzymology, Academic Press, Inc. (1984); Gene Transfer Vectors for Mammalian Cells, J.H. Miller and 20 M.P. Calos, eds., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1987); and Methods in Enzymology, Vols. 154 and 155, Wu and Grossman, and Wu, eds., respectively.

Description of the Preferred Embodiments

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[0088] The present invention relates to novel nucleic acids known as TRPV3 (previously known as VRLX, VRL-S, VR4 and TRPV7), TRPV4 (previously known as VRL3 and OTRPC4), and TRPM8 (previously known as TRPX) that are homologous to the VR1, polypeptides encoded by these nucleic acids, recombinant materials and methods for their production. The specific names given to the three genes follow the nomenclature suggested in Montell et al., *Molecular Cell*, 9:229-231 (2002). The genes have been found to be expressed either in keratinocytes or the DRG, and both TRPV3 and TRPM8 proteins function in temperature sensation. In addition, expression of the TRPV3 and TRPV4 genes

is up-regulated in a rat injury model (see Examples 4 and 6). The present invention also relates to the identification of trkA⁺ pain-specific genes that are expressed in the DRG. Since the aforementioned genes are expressed in keratinocytes and the DRG, function in temperature sensation, and are up-regulated in response to injury, these genes and their related polypeptides can serve as specific therapeutic targets for the design of drugs to treat chronic and nociceptive pain, inflammation and skin disorders. Accordingly, the invention also relates to methods for identifying agents useful in treating pain, inflammation and skin disorders and methods of monitoring the efficacy of a treatment, utilizing these genes and polypeptides. These genes and related polypeptides can also be utilized in diagnostic methods for the detection of pain, inflammation and skin disorders.

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[0089] TRPV3, TRPV4 and TRPM8 belong to the VR family. A Hidden Markov Model (HMM) of the VR1 and VRL1 proteins from different mammalian species including human and an HMM model against Transmembrane 6 (TM6) domain of all known TRP/VRs has been constructed. The six-frame translation of the Human Celera database has been searched against the VR model. Multiple new putative exons with high homology (70% identical and 82% similar in conserved regions among the different VR/TRPs) to Transmembrane 4 (TM4) and TM6 domains to the known TRPs have been identified. These exons map to bacterial artificial chromosomes containing specific human sequences from the High Throughput Genome Sequence (HTGS) database. All the newly-identified exons belong to three new genes of the VR family. Subsequently, RT-PCR has confirmed that these genes are expressed in the DRG or keratinocytes. The structural homology to known TRP channels, the genes' expression in DRG or keratinocytes, their function as temperature-sensitive channels, and the up-regulation of TRPV3 and TRPV4 gene expression observed in a rat injury model in the DRG, indicate that the new genes act as important sensory receptors.

TRPV3: An Ion Channel Responsive to Warm and Hot Temperatures

[0090] TRPV3 is the first molecule described to be activated at warm and hot temperatures, and to be expressed in skin cells (see Examples 2 and 3). TRPV3 signaling mediates a cell-autonomous response in keratinocytes upon exposure to heat. The heat-induced TRPV3 signal is transferred to nearby free nerve endings, thereby contributing to

conscious sensations of warm and hot. This is supported by indirect evidence that skin cells can act as thermal receptors. For instance, while dissociated DRG neurons can be directly activated by heat and cold, warm receptors have only been demonstrated in experiments where skin-nerve connectivity is intact (see Hensel et al., *Pfugers Arch.*, 329:1-8 (1971), Hensel et al., J. Physio., 204:99-112 (1969)). TRPV3 has an activation threshold around 5 33-35°C. The presence of such a warm receptor in skin (with a resting temperature of 34°C) and not DRG neurons (with a resting temperature of 37°C at the cell body) prevents a warmchannel like TRPV3 from being constitutively active at core 37°C temperatures. The residual heat sensitivity in TRPV1 knockout mice also involves skin cells: while dissociated DRG neurons from TRPV1-null animals do not respond to moderate noxious stimulus at all, 10 skin-nerve preparations from such animals do respond (see Caterina et al., Science, 288:306-13 (2000); Davis et al., *Nature*, 405:183-187 (2000); Roza et al., Paper presented at the 31st Annual meeting for the Society of Neuroscience, San Diego, CA (2001)). Collectively these data indicate that a warm/heat receptor is present in the skin, in addition to the heat receptors in DRGs. While synapses have not been found between keratinocytes and sensory termini; 15 ultrastructural studies have shown that keratinocytes contact, and often surround, DRG nerve fibers through membrane-membrane apposition (see Hilliges et al., J. Invest. Dermatol., 104:134-137 (1995) and Cauna., J. Anat., 115:277-288 (1973)). Therefore, heat-activated TRPV3 signal from keratinocytes can be transduced to DRG neurons through direct chemical signaling. One potential signaling mechanism can involve ATP. P2X3, an 20 ATP-gated channel, is present in sensory endings, and analysis of P2X3 knockout mice show a strong deficit in coding of warm temperatures (see Souslova et al., Nature, 407:1015-1017 (2000); Cockayne et al., Nature, 407:1011-1015 (2000)). Furthermore, release of ATP from damaged keratinocytes has been shown to cause action potentials in nociceptors via the P2X 25 receptors (see Cook et al., Pain, 95:41-47 (2002)). Since TRPV3 is activated at innocuous warm and noxious hot temperatures and is expressed in skin, this gene can serve as a therapeutic target for the design of drugs useful in treating pain, inflammation and skin disorders, e.g., those associated with sunburn and other sensitized states.

[0091] In one aspect, the invention provides isolated nucleic acids encoding a mammalian TRPV3 protein. These include an isolated and/or recombinant nucleic acid molecule that encodes a mouse TRPV3 protein having an amino acid sequence as shown in SEQ ID NO: 2. For example, the TRPV3-encoding nucleic acids of the invention include

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those that have a nucleotide sequence as set forth in SEQ ID NO: 1, from nucleotides 65-2440. The nucleic acids of the invention can include not only the coding region, but also the non-coding regions that are upstream and downstream of the coding region and also are provided in SEO ID NO: 1. The invention also provides an isolated mouse TRPV3 polypeptide having an amino acid sequence as shown in SEQ ID NO: 2. Also provided are numerous other nucleic acids that encode this mouse TRPV3 polypeptide; the nucleotide sequences of these nucleic acids are shown in SEQ ID NO: 3.

[0092] Human TRPV3 polypeptides and polynucleotides are also provided by the invention. For example, the invention provides an isolated and/or recombinant human TRPV3-encoding polynucleotide encoding a human TRPV3 polypeptide having an amino acid sequence as set forth in SEQ ID NO: 5. These nucleic acid molecules include those that have a nucleotide sequence as set forth in nucleotides 57-2432 of SEO ID NO: 4. Upstream and downstream non-coding regions are also provided in SEQ ID NO: 4. Also provided by the invention are isolated human TRPV3 polypeptides having an amino acid sequence as set forth in SEQ ID NO: 5. The invention also provides numerous other nucleic acids that encode this human TRPV3 polypeptide; the nucleotide sequences of these nucleic acids are shown in SEQ ID NO: 6.

TRPV4: An Ion Channel that is Activated by Pain

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[0093] TRPV4 is a TRP channel protein that is expressed in adult mouse kidney, 20 newborn dorsal root ganglion and adult trigeminal tissue (see Example 5). TRPV4 is a nonselective cation channel that is activated by decreases in, and is inhibited by increases in, extracellular osmolarity indicating that this channel functions as an osmosensor channel (see, e.g., Strotmann et al., Nat. Cell Biol., 2:695-702 (2000)). In addition, expression of the TRPV4 gene is up-regulated in a rat injury model (see Example 6). Accordingly, the TRPV4 gene can serve as a therapeutic target for the design of drugs to treat pain, kidney disorders and migraine.

[0094] The invention provides isolated nucleic acids that encode a mammalian TRPV4 protein. These include the isolated and/or recombinant nucleic acid molecule that encodes mouse TRPV4 protein having an amino acid sequence as set forth in SEQ ID NO: 14. Included among these nucleic acid molecules are those that have a nucleotide sequence as set forth in nucleotides 156-2771 of SEQ ID NO: 13. Upstream and downstream non-

coding sequences are also provided. Also provided by the invention are isolated mouse TRPV4 polypeptides having an amino acid sequence as set forth in SEQ ID NO: 14. Numerous other nucleic acids that encode this mouse TRPV4 polypeptide are also provided by the invention. The nucleotide sequences of such nucleic acids are shown in SEQ ID NO: 15.

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[0095] The mammalian TRPV4-encoding nucleic acids also include the isolated and/or recombinant nucleic acid molecules that encode human TRPV4 protein that has an amino acid sequence as set forth in SEQ ID NO: 17. Such nucleic acid molecules include those having a nucleotide sequence as set forth in SEQ ID NO: 16. Also provided are isolated human TRPV4 polypeptides having an amino acid sequence as set forth in SEQ ID NO: 17. The invention also provides numerous other nucleic acids that encode this human TRPV4 polypeptide; the nucleotide sequences of these nucleic acids are shown in SEQ ID NO: 18.

TRPM8: An Ion Channel Responsive to Cold Temperatures and to Menthol

[0096] TRPM8 is activated by cold stimuli and a cooling agent (menthol) and is expressed in a select group of DRG neurons that share characteristics of thermoreceptive neurons (see Examples 8 and 9).

[0097] Cells over-expressing TRPM8 show increased intracellular calcium levels when subjected to cold temperatures ranging from 23°C to 10°C (the lower limit of our temperature-controlled perfusion system). The calcium influx and electrophysiological studies described below demonstrate that TRMP8 is a non-selective, plasma membrane cation channel activated by cold temperatures. The ionic permeability of TRPM8 is similar to that of other TRP channels, which are permeable to both monovalent and divalent cations, although calcium permeability estimates (P_{Ca}/P_{Na}) vary from 0.3 to 14 (see, e.g., Harteneck et al., *Trends Neurosci.*, 23:159-166 (2000)). Menthol is a cooling compound that likely acts on endogenous cold-sensitive channel(s) (see Schafer et al., *J. Gen. Physiol.*, 88:757-776 (1986)). That TRPM8-expressing cells are activated and modulated by menthol reinforces the idea that TRPM8 indeed functions as a cold-sensitive channel *in vivo*. The finding that the sensitivity to menthol is dependent on temperature is consistent with the behavior of a subset of isolated DRG neurons that show a raised 'cold' threshold in the presence of menthol (see Reid and Flonta, *Nature*, 413:480 (2001)). With respect to the mechanism of

TRPM8 activation, TRPM8 could be directly gated by cold stimulus through a conformational change, or cold temperatures could act through a second messenger system that in turn activates TRPM8. The rapid activation by menthol suggests a direct gating mechanism, at least for this mode of activation.

[0098] The expression pattern observed for TRPM8 is consistent with a role in

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cold thermoception. First, TRPM8 mRNA is highly-specific to DRG neurons. Within the DRG, TRPM8 is expressed in the small-diameter non-myelinated neurons, which correspond to the c-fiber thermoreceptor and nociceptors (see Scott, Sensory Neurons: Diversity, Development and Plasticity, Oxford University Press, NY (1992)). The lack of TRPM8

10 expression in trkA knockout mice, whose DRGs lack all thermoreceptor and nociceptive neurons, corroborates this finding. Furthermore, the lack of co-expression with VR1, CGRP or IB4 in the adult suggests that TRPM8 is expressed in a unique population of DRG neurons distinct from well-characterized heat nociceptors. Both soma size of neurons that express VRL1 (medium-large neurons) and their co-expression with NF200 (80% co-expression (see Caterina et al., Nature, 398:436-441(1999)) strongly argues that cells expressing TRPM8 and VRL1 are also distinct. Therefore, by using various markers it is

studies of cold-sensitive DRG neurons (see Hensel, *Thermoreception and Temperature Regulation*, Academic Press, London (1981)). A human gene with a high degree of similarity to mouse TRPM8 but no known function was recently shown to be expressed in prostate tissue (see Tsavaler et al., *Cancer Res.*, 61:3760-3769 (2001)).

shown below that TRPM8 is expressed in a sub-class of nociceptors/thermoreceptors that is

distinct from noxious heat sensing neurons, and this correlates well with physiological

[0099] As the first molecule to respond to cold temperatures and menthol, TRPM8 offers interesting insight into the fundamental biology of cold perception. Modulation of TRPM8 activity is also relevant for therapeutic applications: cold treatment is often used as a method of pain relief, and in some instances, hypersensitivity to cold can lead to cold allodynia in patients suffering from neuropathic pain. Modulation of TRPM8 activity is also relevant for treating acute pain, e.g., toothache and other trigeminal focused pain; and for treating cancer, particularly prostate cancer and other prostate disorders.

[0100] The invention provides isolated nucleic acids encoding a TRPM8 mammalian protein. These include the isolated and/or recombinant nucleic acid molecules that encode mouse TRPM8 protein that have an amino acid sequence as set forth in SEQ ID

NO: 8. For example, the invention provides recombinant and/or isolated nucleic acid molecules that have a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7. Upstream and downstream non-coding regions are also provided. The invention also provides isolated mouse TRPM8 polypeptides that include an amino acid sequence as set forth in SEQ ID NO: 8. Also provided are numerous other nucleic acids that encode this mouse TRPM8 polypeptide. Nucleotide sequences of these nucleic acids are provided in SEQ ID NO: 9.

[0101] The nucleic acids encoding a mammalian TRPM8 protein also include isolated and/or recombinant nucleic acid molecules that encode a human TRPM8 protein comprising an amino acid sequence as set forth in SEQ ID NO: 11. For example, the invention provides an isolated and/or recombinant nucleic acid molecule that includes a nucleotide sequence as set forth from nucleotides 61-4821 of SEQ ID NO: 10. Upstream and downstream non-coding regions are also provided by the invention. The invention also provides isolated human TRPM8 polypeptides having an amino acid sequence as set forth in SEQ ID NO:11. The TRPM8 protein is responsive to cold and menthol.

Nucleic Acid Molecules

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[0102] Nucleic acid molecules of the present invention also include isolated nucleic acid molecules that have at least 80% sequence identity, preferably at least 90% identity, preferably at least 95% identity, more preferably at least 98% identity, and most preferably at least 99% identity to a nucleic acid encoding a polypeptide comprising the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 5, SEQ ID NO: 8, SEQ ID NO: 11, SEQ ID NO: 14 or SEQ ID NO: 17, respectively, over the entire coding region or over a subsequence thereof. Such nucleic acid molecules include a nucleic acid having a nucleotide sequence as set forth in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 16 or SEQ ID NO: 18, as set forth above.

[0103] Nucleic acids of the present invention include isolated nucleic acid molecules encoding polypeptide variants which comprise the amino acid sequences of SEQ ID NO: 2, SEQ ID NO: 5, SEQ ID NO: 8, SEQ ID NO: 11, SEQ ID NO: 14 or SEQ ID NO: 17, respectively. Nucleic acids that are amplified using a primer pair disclosed herein are also encompassed by the present invention.

[0104] Further nucleic acids of the present invention also include fragments of the aforementioned nucleic acid molecules. These oligonucleotide probes are preferably of sufficient length to specifically hybridize only to complementary transcripts of the above identified gene(s) of interest under the desired hybridization conditions (e.g., stringent conditions). As used herein, the term "oligonucleotide" refers to a single-stranded nucleic acid. Generally the oligonucleotides probes will be at least 16-20 nucleotides in length, although in some cases longer probes of at least 20-25 nucleotides will be desirable.

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[0105] The oligonucleotide probes can be labeled with one or more labeling moieties to permit detection of the hybridized probe/target polynucleotide complexes. Labeling moieties can include compositions that can be detected by spectroscopic, biochemical, photochemical, bioelectronic, immunochemical, electrical optical or chemical means. Examples of labeling moieties include, but are not limited to, radioisotopes, e.g., ³²P, ³³P, ³⁵S, chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, such as fluorescent markers and dyes, linked enzymes, mass spectrometry tags and magnetic labels.

[0106] Oligonucleotide probe arrays for expression monitoring can be prepared and used according to techniques which are well known to those skilled in the art as described, e.g., in Lockhart et al., *Nature Biotech.*, 14:1675-1680 (1996); McGall et al., *Proc. Natl. Acad. Sci. USA*, 93:13555-13460 (1996); and U.S. Patent No. 6,040,138.

[0107] The invention also provides isolated nucleic acid molecules that are complementary to all the above described isolated nucleic acid molecules.

[0108] An isolated nucleic acid encoding one of the above polypeptides including homologs from species other than mouse or human, may be obtained by a method which comprises the steps of screening an appropriate library under stringent conditions with a labeled probe having the sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 16 or SEQ ID NO: 18, or a fragment thereof; and isolating full-length cDNA and genomic clones containing the nucleotide sequences. Such hybridization techniques are well-known to a skilled artisan.

[0109] Nucleic acid molecules of the present invention may be obtained, using standard cloning and screening techniques, from a cDNA library derived from mRNA in cells of the DRG using the expressed sequence tag (EST) analysis (see Adams et al.,

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Science, 252:1651-1656 (1991); Adams et al., Nature, 355:632-634 (1992); Adams et al., Nature, 377; Suppl. 3:174 (1995)). Polynucleotides of the invention can also be obtained from natural sources such as genomic DNA libraries or can be synthesized using well-known and commercially available techniques.

[0110] It is also appreciated by one skilled in the art, that an isolated cDNA sequence can be incomplete, in that the region coding for the polypeptide is short at the 5' end of the DNA. This can occur due to the failure of the reverse transcriptase to complete a DNA copy of the mRNA transcript during the synthesis of the first strand of cDNA. Methods for obtaining full-length cDNAs, or to extend short cDNAs, are well-known in the art, e.g., those based on the method of RACE as described in Frohman et al., Proc. Natl. Acad. Sci. USA, 85:8998-9002 (1988). The RACE technique has been modified as exemplified by MarathonTM technology (Clontech Laboratories, Inc.), wherein cDNAs have been prepared from mRNA extracted from a selected tissues and an adaptor sequence is ligated to each end. Subsequently, nucleic acid amplification (PCR) is carried out to amplify the missing 5-end of the cDNA using a combination of gene specific and adaptor specific oligonucleotide primers. The PCR reaction is repeated using primers known as nested primers that are designed to anneal with the amplified product, which is generally an adaptor specific primer that anneals further 3' in the adaptor sequence and a gene specific primer that anneals further 5' in the known gene sequence. The reaction products are then analyzed by DNA sequencing and a full-length cDNA is prepared either by directly joining the product to the existing cDNA to provide a complete sequence, or by carrying out a separate full-length PCR using the new sequence information for the design of the 5'primer.

[0111] When nucleic acid molecules of the present invention are utilized for the recombinant production of polypeptides of the present invention, the polynucleotide can include the coding sequence for the mature polypeptide, by itself; or the coding sequence for the mature polypeptide in reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, pro- or prepro-protein sequence, or other fusion peptide portions. For example, a marker sequence which facilitates purification of the fused polypeptide can be encoded, e.g., a hexa-histidine peptide, as provided in the pQE vector (Qiagen, Inc.) and described in Gentz et al., *Proc. Natl. Acad. Sci. USA*, 86:821-824 (1989), or is an HA tag. The nucleic acid molecule can also contain non-coding 5' and 3'

sequences, such as transcribed, non-translated sequences, splicing and polyadenylation signals, ribosome binding sites and sequences that stabilize mRNA.

Polypeptides and Antibodies

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[0112] In another aspect, the present invention relates to mammalian TRPV3,

TRPV4 and TRPM8 polypeptides. These include the mouse TRPV3 polypeptide comprising an amino acid sequence as set forth in SEQ ID NO: 2, the human TRPV3 polypeptide comprising an amino acid sequence as set forth in SEQ ID: 5, the mouse TRPV4 polypeptide comprising an amino acid sequence as set forth in SEQ ID NO: 14, the human TRPV4 polypeptide comprising an amino acid sequence as set forth in SEQ ID NO: 17, the mouse

TRPM8 polypeptide comprising an amino acid sequence as set forth in SEQ ID NO: 8, and the human TRPM8 polypeptide having an amino acid sequence as set forth in SEQ ID NO: 11.

[0113] Further polypeptides of the present invention include isolated polypeptides, i.e., variants, in which the amino acid sequence has at least 90% identity, preferably at least 95% identity, more preferably at least 98% identity and most preferably at least 99% identity, to the amino acid sequences as set forth in SEQ ID NO: 2, SEQ ID NO: 5, SEQ ID NO: 8, SEQ ID NO: 11, SEQ ID NO: 14 or SEQ ID NO: 17 over the entire length of these sequences, or a subsequence thereof. Such sequences include the sequences of SEQ ID NO: 2, SEQ ID NO: 5, SEQ ID NO: 8, SEQ ID NO: 11, SEQ ID NO: 14 and SEQ ID NO: 17.

[0114] The polypeptides of the present invention also include fragments of the aforementioned sequences. For example, the polypeptides of the invention can include amino acids that comprise one or more functional domains of a TRPV3, TRPV4, or TRPM8 polypeptide of the invention. Examples of such domains are described below; other functional domains can be determined using methods known to those of skill in the art.

[0115] The aforementioned TRPV3, TRPV4 and TRPM8 polypeptides can be obtained by a variety of means. Smaller peptides (generally less than 50 amino acids long) may be conveniently synthesized by standard chemical techniques. These polypeptides may also be purified from biological sources by methods well known in the art (see *Protein Purification, Principles and Practice*, 2nd Edition, Scopes, Springer Verlag, NY (1987)). They may also be produced in their naturally occurring, truncated or fusion protein forms by

recombinant DNA technology using techniques well-known in the art. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic techniques and *in vivo* genetic recombination (see, e.g., the techniques described in Sambrook et al., *Molecular Cloning, A Laboratory Manual*, 3rd Edition, Cold Spring Harbor Press, NY (2001); and Ausubel et al., eds., *Short Protocols in Molecular Biology*, 4th Edition, John Wiley & Sons, Inc., NY (1999)). Alternatively, RNA encoding the proteins may be chemically synthesized (see, e.g., the techniques described in *Oligonucleotide Synthesis*, Gait, Ed., IRL Press, Oxford (1984)). Obtaining large quantities of these polypeptides is preferably by recombinant techniques as further described herein.

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[0116] Accordingly, another aspect of the present invention relates to a method for producing a TRPV3, TRPV4 or TRPM8 polypeptide. These methods generally involve:

- a) obtaining a DNA sequence encoding the TRPV3, TRPV4 or TRPM8 polypeptide as defined above; and
- b) inserting the DNA into a host cell and expressing the TRPV3, TRPV4 or TRPM8 polypeptide. In some embodiments, the methods further include:
 - c) isolating the TRPV3, TRPV4 or TRPM8 polypeptide.

[0117] The nucleic acid molecules described herein can be expressed in a suitable host cell to produce active TRPV3, TRPV4 or TRPM8 protein. Expression occurs by placing a nucleotide sequence encoding these proteins into an appropriate expression vector and introducing the expression vector into a suitable host cell, growing the transformed host cell, inducing the expression of one of these proteins, and purifying the recombinant proteins from the host cell to obtain purified, and preferably active, TRPV3, TRPV4 or TRPM8 protein. Appropriate expression vectors are known in the art. For example, pET-14b. pCDNA1Amp and pVL1392 are available from Novagen and Invitrogen and are suitable vectors for expression in E. Coli, COS cells and baculovirus infected insect cells, respectively. These vectors are illustrative of those that are known in the art. Suitable host cells can be any cell capable of growth in a suitable media and allowing purification of the expressed TRPV3, TRPV4 or TRPM8 protein. Examples of suitable host cells include bacterial cells, such as E. Coli, Streptococci, Staphylococci, Streptomyces and Bacillus subtilis cells; fungal cells, such as yeast cells, e.g., Pichia and Aspergillus cells; insect cells, such as Drosophila S2 and Spodoptera Sf9 cells; mammalian cells, such as CHO, COS, HeLa; and plant cells.

[0118] Growth of the transformed host cells can occur under conditions that are known in the art. The conditions will generally depend upon the host cell and the type of vector used. Suitable induction conditions may be used such as temperature and chemicals and will depend on the type of promoter utilized.

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[0119] Purification of the TRPV3, TRPV4 or TRPM8 protein can be accomplished using known techniques without performing undue experimentation. Generally, the transformed cells expressing one of these proteins are broken, crude purification occurs to remove debris and some contaminating proteins, followed by chromatography to further purify the protein to the desired level of purity. Cells can be broken by known techniques such as homogenization, sonication, detergent lysis and freeze-thaw techniques. Crude purification can occur using ammonium sulfate precipitation, centrifugation or other known techniques. Suitable chromatography includes anion exchange, cation exchange, high performance liquid chromatography (HPLC), gel filtration, affinity chromatography, hydrophobic interaction chromatography, etc. Well-known techniques for refolding proteins may be used to obtain the active conformation of the protein when the protein is denatured during intracellular synthesis, isolation or purification.

[0120] In another aspect, the present invention relates to antibodies that recognize epitopes within the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 5, SEQ ID NO: 8, SEQ ID NO: 11, SEQ ID NO: 14 or SEQ ID NO: 17. As used herein, the term "antibody" includes, but is not limited to, polyclonal antibodies, monoclonal antibodies, humanized or chimeric antibodies and biologically-functional antibody fragments which are those fragments sufficient for binding of the antibody fragment to the protein. Antibodies specific for proteins encoded by the aforementioned sequences have utilities in several types of applications. These may include, e.g., the production of diagnostic kits for use in detecting and diagnosing pain, particularly in differentiating among different types of pain. Another use would be to link such antibodies to therapeutic agents, such as chemotherapeutic agents, followed by administration to subjects suffering from pain. These and other uses are described in more detail below.

[0121] For the production of antibodies to a protein encoded by one of the disclosed genes, various host animals may be immunized by injection with the polypeptide, or a portion thereof. Such host animals may include but are not limited to rabbits, mice and rats, to name but a few. Various adjuvants may be used to increase the immunological

response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances, such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants, such as BCG (Bacille Calmette-Guerin) and Corynebacterium parvum.

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[0122] Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of animals immunized with an antigen, such as target gene product, or an antigenic functional derivative thereof. For the production of polyclonal antibodies, host animals, such as those described above, may be immunized by injection with the encoded protein, or a portion thereof, supplemented with adjuvants as also described above.

[0123] Monoclonal antibodies (mAbs), which are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to the hybridoma technique of Kohler and Milstein, *Nature*, 256:495-497 (1975); and U.S. Patent No. 4,376,110, the human B-cell hybridoma technique (see Kosbor et al., *Immunology Today*, 4:72 (1983); Cole et al., *Proc. Natl. Acad. Sci. USA*, 80:2026-2030 (1983), and the EBV-hybridoma technique (see Cole et al., *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96 (1985)). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

[0124] In addition, techniques developed for the production of "chimeric antibodies" (see Morrison et al., *Proc. Natl. Acad. Sci. USA*, 81:6851-6855 (1984); Neuberger et al., *Nature*, 312:604-608 (1984); Takeda et al., *Nature*, 314:452-454 (1985)) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable or hypervariable region derived from a murine mAb and a human immunoglobulin constant region.

[0125] Alternatively, techniques described for the production of single chain antibodies (see U.S. Patent No. 4,946,778; Bird, *Science*, 242:423-426 (1988); Huston et al.,

Proc. Natl. Acad. Sci. USA, 85:5879-5883 (1988); and Ward et al., Nature, 334:544-546 (1989)) can be adapted to produce differentially expressed gene single-chain antibodies. Single-chain antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single-chain polypeptide.

[0126] Most preferably, techniques useful for the production of "humanized antibodies" can be adapted to produce antibodies to the proteins, fragments or derivatives thereof. Such techniques are disclosed in U.S. Patent Nos. 5,932,448; 5,693,762; 5,693,761; 5,585,089; 5,530,101; 5,569,825; 5,625,126; 5,633,425; 5,789,650; 5,661,016; and 5,770,429.

[0127] Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (see Huse et al., *Science*, 246:1275-1281 (1989)) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

Assays for Expression of TRPV3, TRPV4 and TRPM8

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[0128] In another aspect, diagnostic assays are provided which are capable of detecting the expression of one or more of TRPV3, TRPV4 or TRPM8 in human tissue. Such assays are particularly useful in identifying subjects suffering from pain and differentiating among different types of pain. As stated above, expression of the TRPV3 and TRPV4 genes are up-regulated in a rat injury model. Accordingly, up-regulation of the TRPV3 and TRPV4 genes in a sample obtained from a subject suffering from pain compared with a normal value of expression of these genes, e.g., a sample obtained from a subject not suffering from pain, or a pre-established control for which expression of the gene was determined at an earlier time, is indicative of a subject suffering from pain. Expression of one or more of these genes can be detected by measuring either protein encoded by the gene or mRNA corresponding to the gene in a tissue sample, particularly from a human tissue sample obtained from a site of pain.

[0129] Expression of the TRPV3, TRPV4 and TRPM8 proteins can be detected by a probe which is detectably-labeled, or which can be subsequently-labeled. Generally, the

probe is an antibody which recognizes the expressed protein as described above, especially a monoclonal antibody. Accordingly, in one embodiment, an assay capable of detecting the expression of one or more of TRPV3, TRPV4 or TRPM8 genes comprises contacting a human tissue sample with antibodies preferably monoclonal antibodies, that bind to TRPV3, TRPV4 or TRPM8 polypeptides and determining whether the monoclonal antibodies bind to the polypeptides in the sample.

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[0130] Immunoassay methods which utilize the antibodies include, but are not limited to, dot blotting, western blotting, competitive and non-competitive protein binding assays, enzyme-linked immunosorbant assays (ELISA), immunohistochemistry, fluorescence-activated cell sorting (FACS) and others commonly used and widely-described in scientific and patent literature, and many employed commercially.

[0131] Particularly preferred, for ease of detection, is the sandwich ELISA, of which a number of variations exist, all of which are intended to be encompassed by the present invention. For example, in a typical forward assay, unlabeled antibody is immobilized on a solid substrate and the sample to be tested is brought into contact with the bound molecule, followed by incubation for a period of time sufficient to allow formation of an antibody-antigen binary complex. At this point, a second antibody, labeled with a reporter molecule capable of inducing a detectable signal, is then added and incubated, allowing time sufficient for the formation of a ternary complex of antibody-antigen-labeled antibody. Any unreacted material is washed away, and the presence of the antigen is determined by observation of a signal, or may be quantitated by comparing with a control sample containing known amounts of antigen. Variations on the forward assay include the simultaneous assay, in which both sample and antibody are added simultaneously to the bound antibody, or a reverse assay in which the labeled antibody and sample to be tested are first combined, incubated and added to the unlabeled surface bound antibody. These techniques are well-known to those skilled in the art, and the possibility of minor variations will be readily apparent. As used herein, "sandwich assay" is intended to encompass all variations on the basic two-site technique. For the immunoassays of the present invention, the only limiting factor is that the labeled antibody be an antibody which is specific for the protein expressed by the gene of interest, e.g., TRPV3 or a fragment thereof.

[0132] The most commonly used reporter molecules in this type of assay are either enzymes, fluorophore- or radionuclide-containing molecules. In the case of an

enzyme immunoassay an enzyme is conjugated to the second antibody, usually by means of glutaraldehyde or periodate. As will be readily recognized, however, a wide variety of different ligation techniques exist, which are well-known to the skilled artisan. Commonly used enzymes include horseradish peroxidase, glucose oxidase, beta-galactosidase and alkaline phosphatase, among others. The substrates to be used with the specific enzymes are generally chosen for the production, upon hydrolysis by the corresponding enzyme, of a detectable color change. For example, p-nitrophenyl phosphate is suitable for use with alkaline phosphatase conjugates; for peroxidase conjugates, 1,2-phenylenediamine or toluidine are commonly used. It is also possible to employ fluorogenic substrates, which yield a fluorescent product rather than the chromogenic substrates noted above. A solution containing the appropriate substrate is then added to the tertiary complex. The substrate reacts with the enzyme linked to the second antibody, giving a qualitative visual signal, which may be further quantitated, usually spectrophotometrically, to give an evaluation of the amount of TRPV3, TRPV4 or TRPM8 protein which is present in the tissue sample.

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[0133] Alternately, fluorescent compounds, such as fluorescein and rhodamine, may be chemically coupled to antibodies without altering their binding capacity. When activated by illumination with light of a particular wavelength, the fluorochrome-labeled antibody absorbs the light energy, inducing a state of excitability in the molecule, followed by emission of the light at a characteristic longer wavelength. The emission appears as a characteristic color visually detectable with a light microscope. Immunofluorescence and EIA techniques are both very well-established in the art and are particularly preferred for the present method. However, other reporter molecules, such as radioisotopes, chemiluminescent or bioluminescent molecules may also be employed. It will be readily apparent to the skilled artisan how to vary the procedure to suit the required use.

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[0134] The level of expression of mRNA corresponding to the TRPV3, TRPV4 and TRPM8 genes can be detected utilizing methods well-known to those skilled in the art, e.g., northern blotting, RT-PCR, real time quantitative PCR, high density arrays and other hybridization methods. Accordingly, in another embodiment, an assay capable of detecting the expression of one or more of TRPV3, TRPV4 or TRPM8 genes in a sample of tissue, preferably human tissue, is provided which comprises contacting a human tissue sample with an oligonucleotide, i.e., a primer, that is capable of hybridizing to a nucleic acid, particularly

a mRNA, that encodes TRPV3, TRPV4 or TRPM8. The oligonucleotide primer is generally from 10-20 nucleotides in length, but longer sequences can also be employed.

[0135] RNA can be isolated from the tissue sample by methods well-known to those skilled in the art as described, e.g., in Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., 1:4.1.1-4.2.9 and 4.5.1-4.5.3 (1996).

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[0136] One preferred method for detecting the level of mRNA transcribed from the TRPV3, TRPV3, and TRPM8 genes is RT-PCR. In this method, an mRNA species is first transcribed to complementary DNA (cDNA) with use of the enzyme reverse transcriptase. Methods of reverse transcribing RNA into cDNA are well-known and described in Sambrook et al., *supra*. The cDNA is then amplified as in a standard PCR reaction (referred to as PCR) which is described in detail in U.S. Patent Nos. 4,683,195; 4,683,202; and 4,800,159.

[0137] Briefly, in PCR, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target nucleic acid sequence. An excess of deoxynucleoside triphosphates are added to a reaction mixture along with a DNA polymerase, e.g., Taq polymerase. The primers will bind to the target nucleic acid and the polymerase will cause the primers to be extended along the target nucleic acid sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target nucleic acid to form reaction products, excess primers will bind to the target nucleic acid and to the reaction products and the process is repeated.

[0138] Another preferred method for detecting the level of mRNA transcripts obtained from more than one of the disclosed genes involves hybridization of labeled mRNA to an ordered array of oligonucleotides. Such a method allows the level of transcription of a plurality of these genes to be determined simultaneously to generate gene expression profiles or patterns. In particularly useful embodiments, a gene expression profile derived from a tissue sample obtained from a subject suffering from pain can be compared with a gene expression profile derived from a sample obtained from a normal subject, i.e., a subject not suffering from pain, to determine whether one or more of the TRPV3, TRPV4 and TRPM8 genes are over-expressed in the sample obtained from the subject suffering from pain relative to the genes in the sample obtained from the normal subject, and thereby determine

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which gene is responsible for the pain. Ligase chain reaction is another assay that is suitable for detecting the presence of a TRPV3, TRPV4, or TRPM8 polynucleotide.

[0139] The oligonucleotides utilized in this hybridization method typically are bound to a solid support. Examples of solid supports include, but are not limited to, membranes, filters, slides, paper, nylon, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, polymers, polyvinyl chloride dishes, etc. Any solid surface to which the oligonucleotides can be bound, either directly or indirectly, either covalently or noncovalently, can be used. A particularly preferred solid substrate is a high density array or DNA chip. These high density arrays contain a particular oligonucleotide probe in a preselected location on the array. Each pre-selected location can contain more than one molecule of the particular probe. Because the oligonucleotides are at specified locations on the substrate, the hybridization patterns and intensities (which together result in a unique expression profile or pattern) can be interpreted in terms of expression levels of particular genes.

[0140] The oligonucleotide probes are preferably of sufficient length to specifically hybridize only to complementary transcripts of the above identified gene(s) of interest. As used herein, the term "oligonucleotide" refers to a single-stranded nucleic acid. Generally the oligonucleotides probes will be at least 16-20 nucleotides in length, although in some cases longer probes of at least 20-25 nucleotides will be desirable.

[0141] The oligonucleotide probes can be labeled with one or more labeling moieties to permit detection of the hybridized probe/target polynucleotide complexes. Labeling moieties can include compositions that can be detected by spectroscopic, biochemical, photochemical, bioelectronic, immunochemical, electrical optical or chemical means. Examples of labeling moieties include, but are not limited to, radioisotopes, e.g., ³²P, ³³P, ³⁵S, chemiluminescent compounds, labeled binding proteins, heavy metal atoms, spectroscopic markers, such as fluorescent markers and dyes, linked enzymes, mass spectrometry tags and magnetic labels.

[0142] Oligonucleotide probe arrays for expression monitoring can be prepared and used according to techniques which are well-known to those skilled in the art as described, e.g., in Lockhart et al., *supra*); McGall et al., *supra*; and U.S. Patent No. 6,040,138.

[0143] In another aspect, kits are provided for detecting the level of expression of one or more of the TRPV3, TRPV4 and TRPM8 genes in a sample of tissue, e.g., a sample of tissue from a site of pain. For example, the kit can comprise a labeled compound or agent capable of detecting a protein encoded by, or mRNA corresponding to, at least one of the genes TRPV3, TRPV4 and TRPM8; or fragment of the protein, means for determining the amount of protein encoded by or mRNA corresponding to the gene or fragment of the protein; and means for comparing the amount of protein encoded by or mRNA corresponding to the gene or fragment of the protein, obtained from the subject sample with a standard level of expression of the gene, e.g., from a sample obtained from a subject not suffering pain. With respect to detection of TRPV3, TRPV4 and TRPM8 proteins, the agent can be an antibody specific for these proteins. With respect to detection of mRNA, the agent can be pre-selected primer pairs that selectively hybridize to mRNA corresponding to SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 16 and SEQ ID NO: 18. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect protein encoded by or mRNA corresponding to the gene.

[0144] In another aspect, the present invention is based on the identification of novel genes that are specific for trkA⁺ pain-specific DRG neurons. DRG neurons can be classified into several distinct subpopulations with different functional, biochemical and morphological characteristics. The only known early markers differentially expressed by the DRG subtypes are the trk family of neurotrophin receptors. Gene-targeted deletion of the mouse neurotrophins and trks (receptor tyrosine kinases) have demonstrated that neurotrophin signaling is required for the survival of the different subpopulations of DRG neurons that trks specifically mark. For example, trkA knockout mice lack the nociceptive and thermoceptive neurons that sense pain and temperature.

Identification of Agonists and Antagonists

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[0145] In another aspect, the present invention relates to the use of the TRPV3, TRPV4 and TRPM8 genes in methods for identifying agents useful in treating pain, or modulating responses to heat and cold, as flavor enhancers (e.g., menthol mimetics that one can identify using TRPM8 in a screening assay) and as cosmetic additives that provide a

cool or warm sensation to the skin (e.g., menthol mimetics, capsaicin mimetics or other compounds identified using TRPM8 or TRPV3 in screening assays). These methods comprise assaying for the ability of various agents to bind and/or modulate the activity of the proteins encoded by these genes, and/or decrease or increase the level of expression of mRNA corresponding to or protein encoded by these genes. The candidate agent may function as an antagonist or agonist. Examples of various candidate agents include, but are not limited to, natural or synthetic molecules such as antibodies, proteins or fragments thereof, antisense nucleotides, double-stranded RNA, ribozymes, organic or inorganic compounds, etc. Methods for identifying such candidate agents can be carried out in cell-based systems and in animal models.

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[0146] For example, proteins encoding these genes expressed in a recombinant host cell such as CHO or COS may be used to identify candidate agents that bind to and/or modulate the activity of the protein, or that increase or decrease the level of expression of mRNA corresponding to or encoded by these genes. In this regard, the specificity of the binding of a candidate agent showing affinity for the protein can be shown by measuring the affinity of the agents for cells expressing the receptor or membranes from these cells. This can be achieved by measuring the specific binding of labeled, e.g., radioactive agent to the cell, cell membranes or isolated protein, or by measuring the ability of the candidate agent to displace the specific binding of standard labeled ligand.

[0147] Cells expressing proteins encoded by these genes can also be utilized to identify agents that modulate the protein's activity. For example, one method for identifying compounds useful for treating pain, or for use as a flavor or fragrance, comprises, providing a cell that expresses one of these proteins, e.g., TRPV3, TRPV4 or TRPM8, combining a candidate agent with the cell and measuring the effect of the candidate agent on the protein's activity. The cell can be a mammalian cell, a yeast cell, bacterial cell, insect cell or any other cell expressing the TRPV3 protein. The candidate compound is evaluated for its ability to elicit an appropriate response, e.g., the stimulation of cellular depolarization or increase in intracellular calcium ion levels due to calcium ion influx.

[0148] The level of intracellular calcium can be assessed using a calcium ionsensitive fluorescent indicator such as a calcium ion-sensitive fluorescent dye, including, but not limited to, quin-2 (see, e.g., Tsien et al., *J. Cell Biol.*, 94:325 (1982)), fura-2 (see, e.g., Grynkiewicz et al., *J. Biol. Chem.*, 260:3440 (1985)), fluo-3 (see, e.g., Kao et al., *J. Biol.*

Chem., 264:8179 (1989)) and rhod-2 (see, e.g., Tsien et al., J. Biol. Chem., Abstract 89a (1987)).

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[0149] Membrane depolarization of recombinant cells expressing the above proteins can be monitored using a fluorescent dye that is sensitive to changes in membrane potential, including, but not limited to, carbocyanaines such as 3,3'-dipentyloxacarbocyanine iodide (DiOC₅) and 3,3'-dipropylthiadicarbocyanine iodide (DiSC₃), oxonols, such as bis-(1,3-dibutylbarbituric acid) pentamethine oxonol (DiBAC₄ (Biotrend Chemikalien GmbH, Cologne, Germany)) or bis-(1,3-dibutylbarbituric acid) pentamethine oxonol, etc. Cellular fluorescence can be monitored using a fluorometer.

[0150] The assays to identify antagonists of ion channel activity are preferably performed under conditions in which the particular ion channel is active. Conversely, when seeking to identify an agonist, one would preferably perform the screening under conditions in which the ion channel is not active in the absence of the agonist. For example, TRPV3 is activated (i.e., mediates ion passage through a membrane) at temperatures of about 33°C and above. Accordingly, it is preferred to screen for antagonists of TRPV3 at a temperature of above about 33°C (e.g., 35°, 40°, 45°, or above), and to screen for agonists of TRPV3 at a temperature below 33°C (e.g., 30°, 25°, 20°C, or below). In some assays, it is desirable to discriminate between TRPV3-mediated ion transport and ion transport mediated by a different TRP ion channel. For example, to discriminate between TRPV3-mediated cation transport and cation transport mediated by, for example, TRPV1 or TRPV2, the assay can be conducted at a temperature above the activation threshold of TRPV3 but below the activation threshold of the other receptor (e.g., below about 43°C or below about 52°C, respectively, for TRPV1 and TRPV2). Thus, an assay temperature of between about 35°C and about 40°C would result in active TRPV3, but inactive TRPV1 and TRPV2.

[0151] Similarly, assays to identify antagonists of TRPM8 cation channel activity are preferably conducted under conditions in which the TRPM8 conducts cations in the absence of an antagonist. For example, since the threshold activation temperature of TRPM8 is approximately 15°C, one could screen for antagonists at a temperature below 15°C (e.g., 10°, 5°, 0°C, and the like). TRPM8 also is activated by menthol, so instead of or in addition to regulating activity by temperature, one could conduct the assay for antagonists in the presence of menthol. To identify an agonist of TRPM8, it is preferred to conduct the assay under conditions in which TRPM8 does not exhibit significant ion channel activity, such as a

temperature above 15°C (e.g., 20°C, 25°C, 30°C, etc.). To distinguish between TRPM8-mediated cation channel activity and that of other TRP ion channels, the assay for agonists can be conducted at a temperature below 33°C (the activation temperature of TRPV3). For example, a temperature between 20°C and 30°C would result in TRPM8 being inactive in the absence of an agonist, and TRPV3, TRPV1 and TRPV2 also being inactive.

[0152] The TRPV3, TRPV4, and TRPM8 cation channels function to transport not only divalent cations (e.g., Ca²⁺⁺), but also monovalent cations (e.g., Na⁺, K⁺).

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[0153] The assay can be carried out manually or using an automated system. For high throughput screening assays to identify ligands of such proteins, an automated system is preferred. For example, one type of automated system provides a 96-well, 384-well, or 1536-well, culture plate wherein a recombinant cell comprising a nucleotide sequence encoding such a protein is cultured to express the protein. The plate is loaded into a fluorescence imaging plate reader (e.g., "FLIPR®", commercially available from Molecular Devices Corp., Sunnyvale, CA) which measure the kinetics of intracellular calcium influx in each of the wells. The FLIPR® can quantitatively transfer fluids into and from each well of the plate and thus can be utilized to add the calcium-ion sensitive fluorescent indicator dye, a candidate agent, etc. Membrane potential dyes suitable for high throughput assays include the FLIPR® Membrane Potential Assay Kit as sold by Molecular Devices Corp.

[0154] Once a candidate compound is identified as an agonist, such agonists can be added to cells expressing such proteins followed by the addition of various candidate agents to determine which agents function as antagonists.

[0155] The nucleic acids and polypeptides of the present invention can also be utilized to identify candidate agents that modulate, i.e., increase or decrease the level of expression of mRNA and proteins in cells expressing these proteins. For example, expression of the TRPV4 gene is shown to be up-regulated in a rat injury model (see Example 3). The level of expression of mRNA and protein can be detected utilizing methods well-known to those skilled in the art as described above.

[0156] After initial screening assays have identified agents that inhibit the protein's activity or level of expression of mRNA or protein, these agents can then be assayed in conventional live animal models of pain to assess the ability of the agent to ameliorate the pathological effects produced in these models and/or inhibit expression levels of mRNA or protein. For example, in the case of the TRPV4 gene which is shown to be up-

regulated in a rat injury model, one method for identifying an agent useful in the treatment of pain comprises:

a) administering a candidate agent, e.g., an antisense nucleotide derived from the sequence of the TRPV4 gene, to a subject such as a rat model of pain; and

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b) determining reversal of established pain in the animal. Various animal models utilized in neuropathic pain are well-known in the art, e.g., the partial sciatic ligation model, i.e., the Seltzer model, the chronic constriction injury model, i.e., the CCI model and the spinal nerve ligation model, i.e., the Chung model.

[0157] For example, in the partial sciatic ligation (see, the Seltzer model as described in Seltzer et al., *Pain*, 43:205-218 (1990)), rats are anesthetized and a small incision made mid-way up one thigh (usually the left) to expose the sciatic nerve. The nerve is carefully cleared of surrounding connective tissues at a site near the trochanter just distal to the point at which the posterior biceps semitendinosus nerve branches off the common sciatic nerve. A 7-0 silk suture is inserted into the nerve with a 3/8 curved, reversed-cutting mini-needle, and tightly ligated so that the dorsal 1/3 to 1/2 of the nerve thickness is held within the ligature. The muscle and skin are closed with sutures and clips and the wound dusted with antibiotic powder. In sham animals the sciatic nerve is exposed but not ligated and the wound closed as before.

[0158] In the chronic constriction model (the CCI model as described in Bennett et al., *Pain*, 33:87-107 (1988)) rats are anesthetized and a small incision is made midway up one thigh to expose the sciatic nerve. The nerve is freed of surrounding connective tissue and four ligatures of chromic gut are tied loosely around the nerve with approximately 1 mM between each, so that the ligatures just barely construct the surface of the nerve. The wound is closed with sutures and clips. In sham animals the sciatic nerve is exposed but not ligated and the wound is closed.

[0159] In the spinal nerve ligation (see, the Chung model as described in Kim et al., *Pain*, 50:355-363 (1992)) rats are anesthetized and placed into a prone position and an incision made to the left of the spine at the L4-S2 level. A deep dissection through the paraspinal muscles and separation of the muscles from the spinal processes at the L4-S2 level will reveal part of the sciatic nerve as it branches to form the L4, L5 and L6 spinal nerves. The L6 transverse process is carefully removed with a small rongeur enabling visualization of these spinal nerves. The L5 spinal nerve is isolated and tightly ligated with

7-0 silk suture. The wound is closed with a single muscle suture (6-0 silk) and one or two skin closure clips and dusted with antibiotic powder. In sham animals the L5 nerve is exposed as before but not ligated and the wound closed as before.

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[0160] Male Wistar rats (120-140 g) are used for each of the three models. Mechanical hyperalgesia is then assessed in rat by measuring paw withdrawal thresholds of both hindpaws to an increasing pressure stimulus using an Analgesymeter (Ugo-Basile, Milan). Thermal hyperalgesia is assessed by measuring withdrawal latencies to a noxious thermal stimuls applied to the underside of each hindpaw. With all models, mechanical hyperalgesia and allodynia and thermal hyperalgesis develop within 1-3 days following surgery and persist for at least 50 days. Reversal of mechanical hyperalgesia and allodynia and thermal hyperalgesia is assessed following administration of the agent, e.g., the antisense nucleotide specific for the TRPV4 gene.

[0161] Another example of a method for identifying agents useful in treating pain comprises:

- a) administering a candidate agent to a subject such as a rat model of pain;
- b) detecting a level of expression of a protein encoded by or mRNA corresponding to one of genes described herein, e.g., TRPV4, in a sample obtained from the subject; and
- c) comparing the level of expression of the protein or mRNA in the sample in the presence of the agent with a level of expression of the protein or mRNA obtained from the sample of the subject in the absence of the agent, wherein a decreased level of expression of the protein or mRNA in the sample in the presence of the agent relative to the level of expression of the protein or mRNA in the absence of the agent is indicative that the agent is useful in the treatment of pain.

[0162] The present invention also provides a method for identifying an agent useful in the modulation of a mammalian sensory response. The method comprises

- a) contacting a candidate agent with a test system that comprises a receptor polypeptide selected from the group consisting of TRPM8, TRPV3, and TRPV4; and
- b) detecting a change in activity of the receptor polypeptide in the presence of the candidate agent as compared to the activity of the receptor polypeptide in the absence of the agent, thereby identifying an agent that modulates receptor activity.
- [0163] In particularly useful embodiments of this method, the sensory response is response to cold and the polypeptide is a TRPM8 polypeptide preferably having an amino

acid sequence selected from the group consisting of SEQ ID NO: 8 and SEQ ID NO: 11. The method can further include the step of administering the agent that modulates receptor activity to a test subject, and thereafter detecting a change in the sensory response in the test subject.

[0164] The test system that is contacted with a candidate agent can comprise, e.g., a membrane that comprises the receptor polypeptide or a cell that expresses a heterologous polynucleotide that encodes the receptor polypeptide. In a useful embodiment, the heterologous polynucleotide comprises a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7 or as set forth in nucleotides 61-4821 of SEQ ID NO: 10, and the receptor polypeptide is a TRPM8 polypeptide. The cell can be substantially isolated wherein the step of contacting of the cell with the candidate agent is performed *in vitro* or the cell can be present in an organism wherein the step of contacting is performed *in vitro*.

[0165] In particularly useful embodiments, the receptor activity comprises increased or decreased Ca²⁺ passage through the membrane comprising the receptor polypeptide, wherein the membrane can be, e.g., a substantially purified cell membrane or a membrane comprising a liposome.

Pharmaceutical Compositions and Methods

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[0166] The present invention also provides for therapeutic methods of treating a subject suffering from pain utilizing the aforementioned genes, i.e., TRPV3, TRPV4, and TRPM8. Examples of suitable therapeutic agents include, but are not limited to, antisense nucleotides, ribozymes, double-stranded RNAs, antagonists and agonists, as described in detail below.

[0167] As used herein, the term "antisense" refers to nucleotide sequences that are complementary to a portion of an RNA expression product of at least one of the disclosed genes. "Complementary" nucleotide sequences refer to nucleotide sequences that are capable of base-pairing according to the standard Watson-Crick complementary rules. That is, purines will base pair with pyrimidine to form combinations of guanine:cytosine and adenine:thymine in the case of DNA, or adenine:uracil in the case of RNA. Other less common bases, e.g., inosine, 5-methylcytosine, 6-methyladenine, hypoxanthine and others may be included in the hybridizing sequences and will not interfere with pairing.

[0168] When introduced into a host cell, antisense nucleotide sequences specifically hybridize with the cellular mRNA and/or genomic DNA corresponding to the gene(s) so as to inhibit expression of the encoded protein, e.g., by inhibiting transcription and/or translation within the cell.

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[0169] The isolated nucleic acid molecule comprising the antisense nucleotide sequence can be delivered, e.g., as an expression vector, which when transcribed in the cell, produces RNA which is complementary to at least a unique portion of the encoded mRNA of the gene(s). Alternatively, the isolated nucleic acid molecule comprising the antisense nucleotide sequence is an oligonucleotide probe which is prepared *ex vivo* and, which when introduced into the cell results in inhibiting expression of the encoded protein by hybridizing with the mRNA and/or genomic sequences of the gene(s).

[0170] Preferably, the oligonucleotide contains artificial internucleotide linkages which render the antisense molecule resistant to exonucleases and endonucleases, and thus are stable in the cell. Examples of modified nucleic acid molecules for use as antisense nucleotide sequences are phosphoramidate, phosporothioate and methylphosphonate analogs of DNA as described, e.g., in U.S. Patent Nos. 5,176,996; 5,264,564; and 5,256,775. General approaches to preparing oligomers useful in antisense therapy are described, e.g., in Van der Krol., *BioTechniques*, 6:958-976 (1988); and Stein et al., *Cancer Res.*, 48:2659-2668 (1988).

[0171] Typical antisense approaches, involve the preparation of oligonucleotides, either DNA or RNA, that are complementary to the encoded mRNA of the gene. The antisense oligonucleotides will hybridize to the encoded mRNA of the gene and prevent translation. The capacity of the antisense nucleotide sequence to hybridize with the desired gene will depend on the degree of complementarity and the length of the antisense nucleotide sequence. Typically, as the length of the hybridizing nucleic acid increases, the more base mismatches with an RNA it may contain and still form a stable duplex or triplex. One skilled in the art can determine a tolerable degree of mismatch by use of conventional procedures to determine the melting point of the hybridized complexes.

[0172] Antisense oligonucleotides are preferably designed to be complementary to the 5' end of the mRNA, e.g., the 5'untranslated sequence up to and including the regions complementary to the mRNA initiation site, i.e., AUG. However, oligonucleotide sequences that are complementary to the 3' untranslated sequence of mRNA have also been shown to

be effective at inhibiting translation of mRNAs as described e.g., in Wagner, *Nature*, 372:333 (1994). While antisense oligonucleotides can be designed to be complementary to the mRNA coding regions, such oligonucleotides are less efficient inhibitors of translation.

[0173] Regardless of the mRNA region to which they hybridize, antisense oligonucleotides are generally from about 15 to about 25 nucleotides in length.

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[0174] The antisense nucleotide can also comprise at least one modified base moiety, e.g., 3-methylcytosine, 5-methylcytosine, 7-methylguanine, 5-fluorouracil, 5-bromouracil and may also comprise at least one modified sugar moiety, e.g., arabinose, hexose, 2-fluorarabinose and xylulose.

[0175] In another embodiment, the antisense nucleotide sequence is an alpha-anomeric nucleotide sequence. An alpha-anomeric nucleotide sequence forms specific double stranded hybrids with complementary RNA, in which, contrary to the usual beta-units, the strands run parallel to each other as described e.g., in Gautier et al., *Nucl. Acids. Res.*, 15:6625-6641 (1987).

[0176] Antisense nucleotides can be delivered to cells which express the described genes *in vivo* by various techniques, e.g., injection directly into the target tissue site, entrapping the antisense nucleotide in a liposome, by administering modified antisense nucleotides which are targeted to the target cells by linking the antisense nucleotides to peptides or antibodies that specifically bind receptors or antigens expressed on the cell surface.

[0177] However, with the above-mentioned delivery methods, it may be difficult to attain intracellular concentrations sufficient to inhibit translation of endogenous mRNA. Accordingly, in a preferred embodiment, the nucleic acid comprising an antisense nucleotide sequence is placed under the transcriptional control of a promoter, i.e., a DNA sequence which is required to initiate transcription of the specific genes, to form an expression construct. The use of such a construct to transfect cells results in the transcription of sufficient amounts of single-stranded RNAs to hybridize with the endogenous mRNAs of the described genes, thereby inhibiting translation of the encoded mRNA of the gene. For example, a vector can be introduced *in vivo* such that it is taken up by a cell and directs the transcription of the antisense nucleotide sequence. Such vectors can be constructed by standard recombinant technology methods. Typical expression vectors include bacterial plasmids or phage, such as those of the pUC or Bluescript[™] plasmid series, or viral vectors

such as adenovirus, adeno-associated virus, herpes virus, vaccinia virus and retrovirus, adapted for use in eukaryotic cells. Expression of the antisense nucleotide sequence can be achieved by any promoter known in the art to act in mammalian cells. Examples of such promoters include, but are not limited to, the promoter contained in the 3' long terminal repeat of Rous sarcoma virus as described, e.g., in Yamamoto et al., *Cell*, 22:787-797 (1980); the herpes thymidine kinase promoter as described, e.g., in Wagner et al., *Proc. Natl. Acad. Sci. USA*, 78:1441-1445 (1981); the SV40 early promoter region as described e.g., in Bernoist and Chambon, *Nature*, 290:304-310 (1981); and the regulatory sequences of the metallothionein gene as described, e.g., in Brinster et al., *Nature*, 296:39-42 (1982).

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[0178] Ribozymes are RNA molecules that specifically cleave other single-stranded RNA in a manner similar to DNA restriction endonucleases. By modifying the nucleotide sequences encoding the RNAs, ribozymes can be synthesized to recognize specific nucleotide sequences in a molecule and cleave it as described, e.g., in Cech, *J. Amer. Med. Assn.*, 260:3030 (1988). Accordingly, only mRNAs with specific sequences are cleaved and inactivated.

[0179] Two basic types of ribozymes include the "hammerhead" type as described, e.g., in Rossie et al., *Pharmac. Ther.*, 50:245-254 (1991); and the hairpin ribozyme as described, e.g., in Hampel et al., *Nucl. Acids Res.*, 18:299-304 (1999) and U.S. Patent No. 5,254,678. Intracellular expression of hammerhead and hairpin ribozymes targeted to mRNA corresponding to at least one of the disclosed genes can be utilized to inhibit protein encoded by the gene.

[0180] Ribozymes can either be delivered directly to cells, in the form of RNA oligonucleotides incorporating ribozyme sequences, or introduced into the cell as an expression vector encoding the desired ribozymal RNA. Ribozyme sequences can be modified in essentially the same manner as described for antisense nucleotides, e.g., the ribozyme sequence can comprise a modified base moiety.

[0181] Double-stranded RNA, i.e., sense-antisense RNA, corresponding to at least one of the disclosed genes can also be utilized to interfere with expression of at least one of the disclosed genes. Interference with the function and expression of endogenous genes by double-stranded RNA has been shown in various organisms such as *C. elegans* as described e.g., in Fire et al., *Nature*, 391:806-811 (1998); *Drosophila* as described, e.g., in Kennerdell et al., *Cell*, 23;95(7):1017-1026 (1998); and mouse embryos as described, e.g., in Wianni et

al., Nat. Cell Biol., 2(2):70-75 (2000). Such double-stranded RNA can be synthesized by in vitro transcription of single-stranded RNA read from both directions of a template and in vitro annealing of sense and antisense RNA strands. Double-stranded RNA can also be synthesized from a cDNA vector construct in which the gene of interest is cloned in opposing orientations separated by an inverted repeat. Following cell transfection, the RNA is transcribed and the complementary strands reanneal. Double-stranded RNA corresponding to at least one of the disclosed genes could be introduced into a cell by cell transfection of a construct such as that described above.

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[0182] The term "antagonist" with respect to methods of treatment refers to a molecule which, when bound to the protein encoded by the gene, inhibits its activity.

Antagonists can include, but are not limited to, peptides, proteins, carbohydrates and small molecules (generally, a molecule having a molecular weight of about 1000 daltons or less).

[0183] The term "agonist" with respect to methods of treatment refers to a molecule which, when bound to the protein encoded by the gene, activates its activity. Agonists can include, but are not limited to, peptides, proteins, carbohydrates and small molecules.

[0184] In a particularly useful embodiment, the antagonist is an antibody-specific for the cell-surface protein expressed by one of the genes, e.g., TRPV3. Antibodies useful as therapeutics encompass the antibodies as described above, and are preferably monoclonal antibodies. The antibody alone may act as an effector of therapy or it may recruit other cells to actually effect cell killing. The antibody may also be conjugated to a reagent such as a chemotherapeutic, radionuclide, ricin A chain, cholera toxin, pertussis toxin, etc. and serve as a target agent. Alternatively, the effector may be a lymphocyte carrying a surface molecule that interacts, either directly or indirectly, with a tumor target. Various effector cells include, cytotoxic T cells and NK cells.

[0185] Examples of the antibody-therapeutic agent conjugates which can be used in therapy include, but are not limited to: 1) antibodies coupled to radionuclides, such as ¹²⁵I, ¹³¹I, ¹²³I, ¹¹¹In, ¹⁰⁵Rh, ¹⁵³Sm, ⁶⁷Cu, ⁶⁷Ga, ¹⁶⁶Ho, ¹⁷⁷Lu, ¹⁸⁶Re and ¹⁸⁸Re, and as described, e.g., in Goldenberg et al., *Cancer Res.*, 41:4354-4360 (1981); Carrasquillo et al., *Cancer Treat. Rep.*, 68:317-328 (1984); Zalcberg et al., *J. Natl. Cancer Inst.*, 72:697-704 (1984); Jones et al., *Int. J. Cancer*, 35:715-720 (1985); Lange et al., *Surgery*, 98:143-150 (1985); Kaltovich et al., *J. Nucl. Med.*, 27:897 (1986); Order et al., *Int. J. Radiother. Oncol. Biol.*

Phys., 8:259-261 (1982); Courtenay-Luck et al., Lancet, 1:1441-1443 (1984) and Ettinger et al., Cancer Treat. Rep., 66:289-297 (1982); 2) antibodies coupled to drugs or biological response modifiers, such as methotrexate, adriamycin and lymphokines, such as interferon as described, e.g., in Chabner et al., Cancer, Principles and Practice of Oncology, 5 J.B. Lippincott Co., Philadelphia, PA, 1:290-328 (1985); Oldham et al., Cancer, Principles and Practice of Oncology, J.B. Lippincott Co., Philadelphia, PA, 2:2223-2245 (1985); Deguchi et al., Cancer Res., 46:3751-3755 (1986); Deguchi et al., Fed. Proc., 44:1684 (1985); Embleton et al., Br. J. Cancer, 49:559-565 (1984); and Pimm et al., Cancer Immunol. Immunother., 12:125-134 (1982); 3) antibodies coupled to toxins, as described, 10 e.g., in Uhr et al., Monoclonal Antibodies and Cancer, Academic Press, Inc., pp. 85-98 (1983); Vitetta et al., Biotechnology and Bio. Frontiers, P.H. Abelson, Ed., pp. 73-85 (1984) and Vitetta et al., Science, 219:644-650 (1983); 4) heterofunctional antibodies, for example, antibodies coupled or combined with another antibody so that the complex binds both to the carcinoma and effector cells, e.g., killer cells, such as T cells, as described, e.g., in Perez 15 et al., J. Exper. Med., 163:166-178 (1986); and Lau et al., Proc. Natl. Acad. Sci. USA, 82:8648-8652 (1985); and 5) native, i.e., non-conjugated or non-complexed, antibodies, as described in, e.g., in Herlyn et al., Proc. Natl. Acad. Sci. USA, 79:4761-4765 (1982); Schulz et al., Proc. Natl. Acad. Sci. USA, 80:5407-5411 (1983); Capone et al., Proc. Natl. Acad. Sci. USA, 80:7328-7332 (1983); Sears et al., Cancer Res., 45:5910-5913 (1985); Nepom et al., 20 Proc. Natl. Acad. Sci. USA, 81:2864-2867 (1984); Koprowski et al., Proc. Natl. Acad. Sci. USA, 81:216-219 (1984); and Houghton et al., Proc. Natl. Acad. Sci. USA, 82:1242-1246

[0186] Methods for coupling an antibody or fragment thereof to a therapeutic agent as described above are well-known in the art and are described, e.g., in the methods provided in the references above. In yet another embodiment, the antagonist useful as a therapeutic for treating disorders can be an inhibitor of a protein encoded by one of the disclosed genes.

(1985).

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[0187] In the case of treatment with an antisense nucleotide, the method comprises administering a therapeutically effective amount of an isolated nucleic acid molecule comprising an antisense nucleotide sequence derived from at least one of the disclosed genes, wherein the antisense nucleotide has the ability to decrease the transcription/translation of one of the genes. The term "isolated" nucleic acid molecule

means that the nucleic acid molecule is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring nucleic acid molecule is not isolated, but the same nucleic acid molecule, separated from some or all of the coexisting materials in the natural system, is isolated, even if subsequently reintroduced into the natural system. Such nucleic acid molecules could be part of a vector or part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

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[0188] With respect to treatment with a ribozyme or double-stranded RNA molecule, the method comprises administering a therapeutically effective amount of a nucleotide sequence encoding a ribozyme, or a double-stranded RNA molecule, wherein the nucleotide sequence encoding the ribozyme/double-stranded RNA molecule has the ability to decrease the transcription/translation of one of the genes.

[0189] In the case of treatment with an antagonist, the method comprises administering to a subject a therapeutically effective amount of an antagonist that inhibits a protein encoded by one of these genes.

[0190] In the case of treatment with an agonist, the method comprises administering to a subject a therapeutically effective amount of an agonist that inhibits a protein encoded by one of these genes. In particularly useful embodiments, the gene is TRPV8 and the agonist can include compounds that are derivatives of menthol and other compounds known to be cool-feeling agents including, but not limited to, camphor, thymol, peppermint oil, thymol and the like. Such compounds can be particular useful in alleviating pain associated with skin inflammation by providing a cool sensation to the skin.

[0191] A "therapeutically effective amount" of an isolated nucleic acid molecule comprising an antisense nucleotide, nucleotide sequence encoding a ribozyme, double-stranded RNA, agonist or antagonist, refers to a sufficient amount of one of these therapeutic agents to treat a subject suffering from pain. The determination of a therapeutically effective amount is well within the capability of those skilled in the art. For any therapeutic, the therapeutically effective dose can be estimated initially either in cell culture assays, or in animal models, usually mice, rats, rabbits, dogs or pigs. The animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

[0192] The present invention also provides for methods of treating pain, wherein the method comprises identifying a patient suffering from a TRPV3-, TRPV4- or TRPM8-mediated pain by measuring expression of protein encoded by or mRNA corresponding to the TRPV3, TRPV4 or TRPM8 gene, and then administering to such a patient an analgesically effective amount of an agent which decreases or increases the activity or expression of one of these genes. The agent can be a therapeutic agent as described above.

[0193] An "analgesically effective amount" can be a therapeutically effective amount as described above.

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pharmaceutical procedures in cell cultures or experimental animals, e.g., ED₅₀ (the dose therapeutically effective in 50% of the population) and LD₅₀ (the dose lethal to 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index, and it can be expressed as the ratio, LD₅₀/ED₅₀. Antisense nucleotides, ribozymes, double-stranded RNAs, agonists, antagonists and other agents which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies is used in formulating a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, sensitivity of the patient and the route of administration.

[0195] The exact dosage will be determined by the practitioner, in light of factors related to the subject that requires treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, general health of the subject, age, weight and gender of the subject, diet, time and frequency of administration, drug combination(s), reaction sensitivities and tolerance/response to therapy.

[0196] Normal dosage amounts may vary from 0.1-100,000 mg, up to a total dose of about 1 g, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for antagonists.

[0197] For therapeutic applications, the antisense nucleotides, nucleotide sequences encoding ribozymes, double-stranded RNAs (whether entrapped in a liposome or

contained in a viral vector), antibodies or other agents are preferably administered as pharmaceutical compositions containing the therapeutic agent in combination with one or more pharmaceutically acceptable carriers. The compositions may be administered alone or in combination with at least one other agent, such as stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier, including, but not limited to, saline, buffered saline, dextrose and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs or hormones.

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[0198] The pharmaceutical compositions may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intraarticular, intraarterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual or rectal means.

[0199] In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences, Maack Publishing Co., Easton, PA.

[0200] Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well-known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions and the like, for ingestion by the patient.

[0201] Pharmaceutical preparations for oral use can be obtained through combination of active compounds with solid excipient, optionally grinding a resulting mixture, and processing the mixture of granules, after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are carbohydrate or protein fillers, such as sugars, including lactose, sucrose, mannitol or sorbitol; starch from corn, wheat, rice, potato, or other plants; cellulose, such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethylcellulose; gums including arabic and tragacanth; and proteins, such as gelatin and collagen. If desired, disintegrating or solubilizing agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, alginic acid or a salt thereof, such as sodium alginate.

[0202] Dragee cores may be used in conjunction with suitable coatings, such as concentrated sugar solutions, which may also contain gum arabic, talc, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for product identification or to characterize the quantity of active compound, i.e., dosage.

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[0203] Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating, such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with a filler or binders, such as lactose or starches, lubricants, such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid or liquid polyethylene glycol with or without stabilizers.

[0204] Pharmaceutical formulations suitable for parenteral administration may be formulated in aqueous solutions, preferably in physiologically compatible buffers, such as Hank's solution, Ringer's solution or physiologically buffered saline. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils, such as sesame oil or synthetic fatty acid esters, such as ethyl oleate or triglycerides or liposomes. Non-lipid polycationic amino polymers may also be used for delivery. Optionally, the suspension may also contain suitable stabilizers or agents which increase the solubility of the compounds to allow for the preparation of highly concentrated solutions.

[0205] For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

[0206] The pharmaceutical compositions of the present invention may be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping or lyophilizing processes.

[0207] The pharmaceutical composition may be provided as a salt and can be formed with many acids, including but not limited to, hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, etc. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preferred preparation may be a lyophilized powder which may contain any or all of the following: 1-50 mM histidine, 0.1-2% sucrose, and 2-7% mannitol, at a pH range of 4.5-5.5, that is combined with buffer prior to use.

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[0208] After pharmaceutical compositions have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. For administration of the antisense nucleotide or antagonist, such labeling would include amount, frequency and method of administration. Those skilled in the art will employ different formulations for antisense nucleotides than for antagonists, e.g., antibodies or inhibitors. Pharmaceutical formulations suitable for oral administration of proteins are described, e.g., in U.S. Patent Nos. 5,008,114; 5,505,962; 5,641,515; 5,681,811; 5,700,486; 5,766,633; 5,792,451; 5,853,748; 5,972,387; 5,976,569; and 6,051,561.

[0209] In another aspect, the treatment of a subject, e.g., a rat injury model, with a therapeutic agent such as those described above, can be monitored by detecting the level of expression of mRNA or protein encoded by at least one of the disclosed genes, or the activity of the protein encoded by the gene. These measurements will indicate whether the treatment is effective or whether it should be adjusted or optimized. Accordingly, one or more of the genes described herein can be used as a marker for the efficacy of a drug during clinical trials.

[0210] In a particularly useful embodiment, a method for monitoring the efficacy of a treatment of a subject suffering from pain with an agent (e.g., an antagonist, protein, nucleic acid, small molecule or other therapeutic agent or candidate agent identified by the screening assays described herein) is provided comprising:

- a) obtaining a pre-administration sample from a subject prior to administration of the agent;
- b) detecting the level of expression of mRNA or protein encoded by the gene, or activity of the protein encoded by the gene in the pre-administration sample;
 - c) obtaining one or more post-administration samples from the subject;

d) detecting the level of expression of mRNA or protein encoded by the gene, or activity of the protein encoded by the gene in the post-administration sample or samples;

- e) comparing the level of expression of expression of mRNA or protein encoded by the gene, or activity of the protein encoded by the gene in the pre-administration sample with the level of expression of mRNA or protein encoded by the gene, or activity of the protein encoded by the gene in the post-administration sample or samples; and
 - f) adjusting the administration of the agent accordingly.

[0211] For example, increased administration of the agent may be desirable to decrease the level of expression or activity of the gene to lower levels than detected, i.e., to increase the effectiveness of the agent. Alternatively, decreased administration of the agent may be desirable to increase expression or activity of the gene to higher levels than detected, i.e., to decrease the effectiveness of the agent.

EXAMPLES

[0212] The following examples are offered to illustrate, but not to limit the present invention.

EXAMPLE 1

Identification of New VRs

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A. VR searching

[0213] Strategy: Known VR sequences are downloaded (GI Nos. 6782444, 5305598, 7106445, 4589143, 6635238, 2570933, 5263196 and 4589141) from NCBI and assembled using Clustal (Megalign--DNAstar, Madison, WI) with the following parameters: Gap Penalty 10, GapLength Penalty 10, Ktuple 1, Window 5 and Diagonals Saved 5. This alignment is saved as a *.MSF file.

[0214] This *.MSF file is converted to a hidden Markov model using

HMMBUILD 2.0 (Sean Eddy, Washington University, St. Louis) then calibrated using

HMMCALIBRATE 2.0 (Sean Eddy), and used to search 6 frame translations (Feb 20 release) of the Celera human genome data using the default parameters. The protein sequences of these files are retrieved and used as subjects in a BLASTP search of NR. This file is manually inspected identifying three novel candidates for VRs.

B. Identification of VR TRPV3

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[0215] Mechanical and thermal stimuli activate specialized sensory neurons that terminate in the skin at receptor structures like hair follicles or as free nerve endings. Pain and temperature sensitive neurons belong to the latter category and are thus thought to directly sense stimuli. A TRP channel that is expressed in pain neurons, VR1 is partially responsible for the detection of noxious heat. This Example describes the cloning of TRPV3, a close relative of VR1 that is also activated by noxious heat. Surprisingly, TRPV3 is most highly-expressed in skin cells. Keratinocytes that express TRPV3 show a calcium influx in response to noxious heat. Therefore, skin cells possess molecular tools similar to those of sensory neurons to "sense" heat.

[0216] VR1 (TRPV1), the best-characterized receptor in the somatic sensory system, is directly gated by noxious heat. VR1 is expressed in small-diameter, nociceptive DRG neurons that terminate in the skin as free nerve endings to detect noxious heat. Analysis of VR1 knockout mice has demonstrated that this channel is partially responsible for heat sensitivity. VR1 belongs to the family of six transmembrane-containing TRP non-selective cation-channels that function in mechanosensation, osmoregulation and replenishment of intracellular calcium stores. This TRPV family includes at least five members, three of which are expressed in DRG neurons. One of these, VRL1 (TRPV2), is also gated by heat, but has a higher threshold than VR1 (52°C instead of 43°C) and is not co-expressed with VR1. Recent experiments have implied that VRL1 expression does not correlate with the heat-sensitive neurons in VR1 knockout mice, suggesting the existence of yet another heat-sensing channel.

[0217] Public and Celera databases for VR1-related TRP channels are searched by constructing a Hidden Markov Model (HMM) of the VR1 and VRL1 protein sequences from different mammalian species. With this model, the 6-frame translation of human sequence is queried and has identified multiple new putative exons with a great degree of sequence similarity to the ankyrin and transmembrane domains of VR1. These exons map to two genes, one of which is TRPV4, as described, e.g., in Liedtke et al., Cell, 103:525-35 (2000); and Strotmann et al., supra). The other novel gene is known as TRPV3.

[0218] The full-length sequence of mouse TRPV3 is derived from a combination of exon-prediction software, PCR and RACE amplification from newborn mouse DRG and skin cDNA. For PCR cloning, primers (5'-TGACATGATCCTGCTGAGGAGTG-3'

(SEQ ID NO: 19) and 5'-ACGAGGCAGGCAGGCAGGTATTCTT-3' (SEQ ID NO: 20)) are designed from the HMM sequences for TRPV3 as a result of blast hits to the ankyrin and transmembrane domains and used to amplify a 699-nucleotide fragment of TRPV3 from newborn DRG cDNA. From this initial fragment, Rapid Amplification of cDNA Ends (RACE) PCR (Clontech) is used to obtain the 5' and 3' ends of TRPV3 from mouse newborn skin and DRG cDNA. In order to characterize the genomic locus of VR1 and TRPV3, primers are designed from predicted HMM TRPV3 exon sequences and used to screen a genomic BAC Mouse (RPCI22) library (Roswell Park Cancer Institute). Primers utilized are shown in Table 1. Additionally, mouse VR1 BACs are identified by hybridizing a 320 bp probe spanning the mouse VR1 ankyrin region to the same BAC library. Positive BAC clones are further characterized by restriction digest mapping, pulse field gel electrophoresis, and Southern blotting as previously described using probes specific to the 5' and 3' ends of the VR1 and TRPV3 genes. BAC clones positive for TRPV3 included 5J3. BAC clones that were positive for both VR1 and TRPV3 included 9e22, 27I14, 82c1 and 112g17. BACs positive for VR1 included 137N13, 137O13, 234J23, 246D9 and 285G11.

Table 1: TRPV3 Primers

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		SEQ ID NO:
5' RACE		
AP40	CAGCGTATGCAGAGGCTCCAGGGTCAG	21
AP4	TTGAAGTCCTCAGCCACCGTCACCA	22
Mvr4ANK	CACCAGCGCGTGCAGGATGT	23
AP105 RACE-rev	tcgttctcctcagcgaaggcaagcaga	24
AP110R (nested)	CCTTCTATCTCCAGGAAGAAGTGTGC	25
ap113r (race)	GTCACCAGCGCGTGCAGGATGTTGT	26
ap36	AGGCCCATACGCCCAGTCCGTGAAC	27
ap33R	CATGCCCATAGACTGGAAGCC	28
ap71	GATGGCGATGTTCAGCGCTGTCTGC	29
3' RACE		
AP37	GCTGCCAAGATGGGCAAGGCTGAGA	30
Ap31	CCTGGGCTGGCGAACATGCTCTA	31
TM6VR4RACE	GCGCCAGATGCGTTCACTTTCTTTGGA	32
Primers to amplify partial and/or full-length TRPV transcript		SEQ ID NO:
mVR4-F	TGACATGATCCTGCTGAGGAGTG	33
mVR4-R	ACGAGGCAGGCGAGGTATTCTT	34

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AP72 F	TCCAAGCTGTGCTTGTGATA	35
AP73R	CTTGAGCATGTAGTTTCACACAAA	36
AP74R	GTGTTTTCCATTCCGTCCAC	37
AP75R	CGACGTTTCTGGGAATTCAT	38
AP76R	CTTGAGCATGTAGTTTCACACAAA	39
AP77F	TCCTCCTCAACATGCTC	40
AP78R	TGGAAATCAAAACAGTATTTCAATG	41
AP79F	CTCTTCAAGCTCACCATAGGC	42
AP80R	CGACGTTTCTGGGAATTCAT	43
AP81R	GTGTTTTCCATTCCGTCCAC	44
AP82R	CCCTCTGTTACCGCAGACAC	45
AP83F	ACTCCAGCCTGGGTGACA	46
AP84R	ATGGTCTCCAGCTCCCAGTT	47
AP85R	AGGAGGACGAAGGTGAGGAT	48
AP86F	AGCCTCAGGTCTGAAGTGGA	49
AP87R	GCCAGATGCGTTCACTTTCT	50
AP88R	GGCAAATTTCTTCCATTTCG	51
AP89R	AGATGCGTTCGCTCTCTT	52
AP102F	TGCACACTTCTTCCTGGAGAT	53
AP103F	TTCCTCATGCACAAGCTGAC	54
AP104F	TCTTCCTGGAGATAGAAGGGATT	55
AP106R	CGATGATTTCCAGCACAGAG	56
AP107F	CTCACCAATGTAGACACAACGAC	57
AP108F	TACCAGCATGAAGGCTTCTATTT	58
AP109R	ATAAGCACTGCTGTGATGTCTCC	59
AP111R	GTCAGCTTGTGCATGAGGAA	60
AP112F	TGACAGAGACCCCATCCAATCCCAACA	61
AP114F	CTCTTGTGATATGGCTTTCTGG	62
AP115F	GAGAAGGAGTGGGTGAGCTG	63
AP116R	CCTTCTCCCAGAGTCCACAG	64
AP117F	AGCAGGCAGGAAAATGAGAG	65
AP118R	CCAAAGATGGTCCAGAAAGC	66
AP115F	CTCTTGTGATATGGCTTTCTGG	67
AP116F	AACTGTGATGACATGGACTCTCCCCAG	68

AP118F	AACTGTGATGACATGGACTC	69	
AP119F	CAGGATGATGTGACAGAGACCCCATC	70	
AP128F	ATGATCCTGCTGAGGAGTGG	71	
AP129R	AGGATGACACAGGCCCATAC	72	
AP130F	ATCCTCACCTTCGTCCTCCT	73	
AP131R	CATTCCGTCCACTTCACCTC	74	
AP204R (3'UTR)	TGGTTTTGCTGTTGTTCCTG	75	
AP205R (POLYA)CATGTAAATCAACGCAGAAGTCA 76			

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[0219] Several murine ESTs from skin tissues contain 3' UTR TRPV3 sequence (BB148735, BB148088, BB151430 and AI644701), and recently the human TRPV3 sequence has been annotated (see GI: 185877, 18587705 and Peng et al., *Genomics*, 76:99-109 (2001)).

[0220] As predicted from the nucleotide sequence, TRPV3 is composed of 791 amino acid residues. The overall sequence of mouse TRPV3 has 43% identity to TRPV1 (VR1) and TRPV4; 41% to TRPV2 (VRL1); and 20% to TRPV5 (ECAC) and TRPV6 (see Figure 2C). TRPV3 has four, instead of the usual three, predicted N-terminal ankyrin domains that are thought to be involved in protein-protein interactions, TM6 domains and a pore loop region between the last two membrane spanning regions. Two coiled-coil domains N-terminus to the ankyrin domains in TRPV3 are also identified (see Figure 2F). Coiled-coil domains are implicated in oligomerization of GABA-B channels, and have been previously reported to be present in some TRP channels, but not for TRPVs. Further examination shows that VR1, but not the other members of the TRPV family, also has putative coiled-coil domains in the same N-terminal location. Phylogenetic analysis illustrates that TRPV3 is indeed a member of the OTRP/TRPV sub-family, which is part of the larger TRP ion channel family (see Figure 2A). The same BAC genomic clone in the public database contains the sequence of TRPV3 and VR1. Both genes map to human chromosome 17p13 and mouse chromosome 11B4. Mapping analysis of these BAC clones, and later the assembled human and mouse genome sequences reveals the distance between the two genes to be about 10 kb (see Figure 2B). This suggests that TRPV3 and VR1 are derived from a single duplication event.

EXAMPLE 2

Localization of TRPV3 Expression

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A. Northern blot analysis

[0221] For Northern blot analyses approximately 3 μg of polyA⁺ RNA extracted from adult mouse and newborn tissue are electrophoresed on 1% glyoxal gels, transferred and hybridized at high-stringency with a ³²P labeled probe representing the entire full-length TRPV3 sequence. Commercial Northern blots (Clontech) are hybridized with the same TRPV3 full-length probe. For human skin specific expression, Northern blots are prepared from 20 μg of total RNA from primary keratinocytes and cell lines CRL-2309 and CRL-2404 (ATCC) or from 2 μg of polyA⁺ adult and fetal skin RNA (Stratagene). Blots are hybridized with a probe corresponding to the ankryin 1-TM2 region of the TRPV3 human sequence. For VR1 hybridizations, a probe corresponding to nucleotides 60-605, encoding the amino terminus of rat VR1 are used on mouse blots. The entire coding sequence of human VR1 are used as a probe on human Northern blots.

[0222] As stated above, to determine the overall tissue distribution of TRPV3, the full-length mouse TRPV3 sequence is used as a probe for Northern blot analysis. No TRPV3 expression is detected using commercial Northern blots. Blots from adult rat are then used that include tissues relevant to somatic sensation, including DRG, spinal cord and different sources of skin. A mRNA of approximately 6.5 kb is present in tissues derived from skin but not in DRGs. Probing the same adult blot with a TRPV1-specific probe confirms its strong expression in DRG while demonstrating a lack of expression in skin tissues. Northern blot analysis of human adult and fetal skin also shows expression of TRPV3. Cultured primary mouse keratinocytes as well as several epidermal cell lines do not show any TRPV3 expression by Northern blots. These finding suggest that TRPV3 expression may get down regulated after tissue dissociation and long-term culture. Northern blots from newborn and adult mice that include tissues relevant for somatic sensation, including DRG, spinal cord and different sources in skin also show TRPV3 expression in skin tissues with weak expression in the DRG.

B. In situ hybridization

[0223] For *in situ* hybridizations, newborn and adult tissues are dissected, fixed in 4% paraformaldehyde in PBS, cryoprotected and frozen in liquid nitrogen in OCT mounting

medium. Cryostat sections (10 μm) are processed and probed with either a digoxygenin cRNA probe or a ³⁵S-labeled probe generated by *in vitro* transcription as described in Wilkinson, in *Essential Developmental Biology, A Practical Approach*, C. Stern, P. Holland, eds., Oxford Univ. Press, NY, pp. 258-263 (1993). Two mouse TRPV3-specific antisense riboprobes are used, one corresponding to nucleotides 235-1020 encoding the amino terminus and the other spanning nucleotides 980-1675 corresponding to the region between the third ankyrin and TM4 domains.

[0224] Digoxygenin-labeled probes show specific expression in specialized skin tissues, such as hair follicles in both newborn and adult mice. Expression in epidermis is difficult to assess, because of high background observed in this tissue with the sense probe. To circumvent this problem, and to gain more sensitivity, ³⁵S-radioactive *in situ* hybridizations are carried out on cross-sections of newborn mice. Clear expression is detected in the epidermis and hair follicles. No significant expression is detected in DRGs.

C. Immunohistochemical staining assays

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[0225] For immunohistochemistry, rabbits are immunized (AnimalPharm Services, Healdsburg, CA) with KLH conjugated peptide corresponding to either the N-terminus of mouse TRPV3 (CDDMDSPQSPQDDVTETPSN (SEQ ID NO: 77)) or a C-terminus peptide (KIQDSSRSNSKTTL (SEQ ID NO: 78)). Affinity purified antiserum recognizes a band of relative molecular mass ~85 kDa in whole-cell extracts prepared from CHO cells stably transfected with mouse TRPV3 (not shown). For peptide competition, diluted antibody solutions (1:5000) of TRPV3 are pre-incubated (room temperature, 2 hours) with TRPV3 antigenic peptide (9 μgmL⁻¹) prior to incubation with tissue sections. Immunofluorescence are performed on fixed frozen and paraffin sections using rabbit anti-TRPV3 (1:5000), pan cytokeratin (Abcam) cytokeratin (1:300, Abcam), cytokeratin 10 (K8.60, Sigma), pan-basal Cytokeratin (Abcam), PGP9.5 (Abcam) followed by FITC-labeled goat anti-rabbit (10 μg/mL⁻¹) and Cy-3-labeled donkey anti-mouse (Jackson Immunoresearch) antibodies.

[0226] Using polyclonal antibodies produced against TRPV3 peptides from either the N-terminus or the C-terminus, intense TRPV3 immunoreactivity is observed in most keratinocytes at the epidermal layer and in hair follicles from newborn and adult rodent tissues. In the epidermis, staining is absent in the outermost layers (stratum corneum and

lucidum) as well as the basement membrane. In hair follicles, expression is localized to the outer root sheath and absent from the matrix cells, inner root sheath and sebaceous glands. Developmentally, expression in hair follicles increases from newborn to adult stages. High magnification of these images indicates staining in the cytoplasm and at high levels in the plasma membrane.

[0227] Coexpression with various keratinocyte-specific markers shows that TRPV3 is expressed in the basal keratinocytes, which *in vitro* require low calcium concentrations to maintain their undifferentiated state, as well as in some of the more differentiated suprabasal layers of the epidermis. Temperature-sensing neurons are thought to terminate as free nerve endings mainly at the level of dermis, but some processes do extend into the epidermis (see Hilliges et al., *supra*; and Cauna, *supra*. Cutaneous termini can be labeled with the immunohistochemical marker protein gene product 9.5 (PGP 9.5), and it is observed that these epidermal endings indeed co-localize with TRPV3.

D. GFP-fusion constructs

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[0228] The full-length mouse TRPV3 is amplified and subcloned into pcDNA3.1/CT-GFP-TOPO (Invitrogen). *In vitro* transcription/translation (TnT System, Promega) confirms the integrity of the constructs. Cells are viewed live or fixed in 4% paraformaldehyde 48-72 hours after transfection, counterstained with propidium iodide and mounted in Slowfade (Molecular probes).

[0229] Confocal fluorescence microscopy on cells transiently transfected with a C-terminally GFP-tagged TRPV3 protein construct also finds the protein mainly localized at the plasma membrane. This pattern of expression at the cell membrane is consistent with TRPV3 having a role as an ion channel. In sum, the expression analysis suggests that TRPV3 is most prominently expressed in plasma membrane of keratinocytes in both rodents and humans.

EXAMPLE 3

Activation of TRPV3 Protein by Heat

A. Effect of heat, capsazepine and ruthenium red upon conductance
[0230] Given the high degree of homology of TRPV3 to TRPV family members,
TRPV3 is tested to determine whether it responds to stimuli known to activate other closely-

related family members. Accordingly, the effects of heat upon TRPV3 activity in mediating conductance are examined using whole-cell patch-clamp analysis of transfected CHO cell lines expressing TRPV3.

[0231] Mouse TRPV3 and rat TRPV1 cDNA are subcloned into pcDNA5 (Invitrogen) and transfected into CHO-K1/FRT cells using Fugene 6 (Roche). The transfected cells are selected by growth in MEM medium containing 200 μg/mL hygromycin (Gibco BRL). Populations are frozen at early passages and these stocks are used for further studies. Stable clones that express the mRNAs are identified by Northern blot analysis as well as Southern blotting to confirm integration site. Long-term cultures are subsequently maintained at 33°C.

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[0232] TRPV3 expressing CHO cells are assayed electrophysiologically using whole cell voltage clamped techniques. Currents are recorded via pCLAMP8 suite of software via an Axopatch 200A and filtered at 5 kHz. Series-resistance compensation for all experiments is 80% using 2-5 M Ω resistance, fire-polished pipettes. Unless stated, the holding potential for most experiments is -60 mV, apart from the current-voltage relationship studies (2 second ramp from -100 to +80 mV). Cells are normally bathed in a medium containing (mM): NaCl, 140; KCl, 5; Glucose; 10, HEPES, 10; CaCl₂, 2; MgCl₂ 1; titrated to pH 7.4 with NaOH, apart from the monovalent permeability studies, when NaCl is replaced by equimolar KCl or CsCl with the omission of KCl, 5 mM. For the divalent permeability studies, the solutions either contain 1 mM Ca²⁺ or Mg²⁺ and (mM) NaCl, 100; Glucose, 10; Hepes, 10; sucrose, 80 or 30 mM test ion, in the above solution minus sucrose. The experiments in calcium free media have no added CaCl₂ with the addition of 100 μ M EGTA. Pipette solution is always (mM) CsCl, 140; CaCl₂, 1; EGTA, 10; HEPES, 10; MgATP, 2; titrated to pH7.4 with CsOH. For the permeability, ratios for the monovalent cations relative to Na (P_X/P_{Na}) are calculated as follows:

$$P_X/P_{Na} = E_{shift} = \{RT/F\} \log (P_X/P_{Na} [X]_O / [Na]_O)$$

where F is Faraday's constant, R is the universal gas constant, and T is absolute temperature. For the divalent ions, P_{Ca} or P_{Mg}/P_{Na} is calculated as follows:

$$E_{\text{shift}} = \{RT/F\} \log \{[Na]_O + 4B', [X]_{O(2)}\} / \{[Na]_O 4B', [X]_{O(1)}\}$$

where B' = P'_X/P_{Na} and $P'_X = P_X/(1 + e^{EF/RT})$ and $[X]_{O(1)}$ and $[X]_{O(2)}$ refer to the two different concentrations of the divalent ion tested.

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[0233] The results from transfected cells assayed electrophysiologically via whole cell voltage clamped techniques are described below. Capsaicin (1 µM), an activator of TRPV1, does not evoke a response in TRPV3-expressing cells. Similarly no current responses are seen when TRPV3-expressing cells are challenged with a hypo-osmotic solution containing 70 mM NaCl or with low pH (5.4). However, raising the temperature of superperfused external solution from room temperature to 45°C evokes currents in TRPV3 expressing cells. Analysis of currents evoked by temperature ramps from ~15°C to ~48°C (see Figure 3A) shows that little current is elicited until temperatures rise above ~33°C and that the current continues to increase in the noxious temperature range (>42°C). With these findings, TRPV3-expressing cells are subsequently maintained at 33°C to avoid constitutive activation. The current amplitude is influenced by the presence or absence of Ca2+ in the external medium, with reduced current amplitudes in the presence of 2 mM Ca²⁺ after a prior challenge in Ca²⁺-free solution (see Figure 3B). This finding is reminiscent of the channel properties of TRPV5 and TRPV6 (see Nilius et al., J. Physiol., 527:239-248 (2000)). As shown in Figure 3C, the heat evoked current in TRPV3-expressing CHO cells increases exponentially at temperatures above 35°C with an e-fold increase per 5.29 ± 0.35 °C (n=12), corresponding to a mean Q_{10} of 6.62. This temperature dependence is considerably greater than that seen for most ion channel currents, which typically have Q₁₀ values in the range 1.5-2.0, but is less than the values noted for TRPV1 (VR1, Q10 = 17.8) (see Vyklicky et al., J. Physiol., 517:181-192 (1999)). In some cells it is difficult to see a sharp threshold temperature. However, measurable temperature dependent currents below 30°C show an efold increase for a 22.72 \pm 3.31°C (n=12) increase in temperature (Q₁₀ = 1.69).

[0234] The elevated temperature evoked currents, in TRPV3-expressing cells, shows a pronounced outward rectification (see Figure 3D) with a reversal potential in the standard recording solution of -1.22 ± 1 mV. Reducing the NaCl in the external solution to 70 mM (from 140 mM) shifts the reversal potential by -19mV as expected for a cation selective conductance (shift = -17.5 mV). Differences in reversal potentials are also used to determine the ionic selectivity of TRPV3 channels. In simplified external solutions, the reversal potentials of the heat activated currents are very similar when NaCl ($E_{rev} = -1.22 \pm 1.08$ mV, n=5) is replaced with either KCl ($E_{rev} = -0.40 \pm 0.77$ mV, n=6) or CsCl ($E_{rev} = -1.14 \pm 0.53$ mV, n=6), which yields relative permeability ratios P_K/P_{Na} and P_{Cs}/P_{Na} close to 1 (see Funayama et al., *Brain Res. Mol. Brain Res.*, 43:259-266 (1996)). The relative

permeability of Ca^{2+} and Mg^{2+} are estimated from the shift in reversal potentials when their concentrations are raised from 1 mM to 30 mM in a 100 mM NaCl solution containing the divalent cation under investigation. The reversal potential shifts (from -9.1 +1.40 mV to +11.29 + 0.38 mV for Ca^{2+} and from -8.41 ± 0.50 mV to +10.34 ± 2.38 mV for Mg^{2+}) correspond to $P_{\text{Ca}}/P_{\text{Na}} = 2.57$ and $P_{\text{Mg}}/P_{\text{Na}} = 2.18$. These data show that TRPV3 is a non-selective cation channel that discriminates poorly between the tested monovalent cations and has significant divalent cation permeability.

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[0235] Heat activation of TRPV3 shows a marked sensitization with repeated heat stimulation. This is studied at a steady membrane potential of -60 mV and with voltage ramps. The first response to a step increase from room temperature to ~48°C is often very small, but the current response grew with repeated heat steps (see Figure 4A). Sensitization to heat has also been observed for TRPV1 and TRPVL (see Caterina et al., supra and Jordt et al., Cell, 108:421-430 (2002)). Application of voltage ramps shows that sensitization is associated with an increase in outward rectification (see Figure 4B). A protocol of repeated temperature challenges is used to investigate if antagonists of TRPV1 (VR1) are inhibitors of TRPV3. Under normal conditions, a heat challenge delivered 2 minutes after 4-5 sensitizing heat steps evokes a current that is 1.57 ± 0.25 (n=4) times the amplitude of the preceding response (see Figure 4C). Application of 10 µM capsazepine, a competitive capsaicin antagonist at TRPV1, for 2 minutes prior to the test heat challenge does not reduce the current amplitude (2.31 \pm 0.36 times the amplitude of the preceding response, n=4). In contrast, a similar exposure to 1 µM ruthenium red, a non-competitive inhibitor of other TRPV channels, reduces the relative amplitude of the heat response to 0.34 ± 0.03 , n=5 (see Figure 4D). Taken together, these results indicate that TRPV3 is a cation permeable channel activated by warm and hot temperatures and has channel properties reminiscent of other TRPV channels.

EXAMPLE 4

Gene Expression Analysis of TRPV3 in the Rat Chung Model

[0236] These studies discussed below measure relative levels of RNA expression for TRPV3 in the Chung neuropathic pain model using RT-PCR.

A. Spinal nerve ligation (Chung) model

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[0237] This model is established according to the methods described by Kim and Chung, *supra*, 1992. Rats are anesthetized and placed into a prone position and an incision made to the left of the spine at the L4-S2 level. A deep dissection through the paraspinal muscles and separation of the muscles from the spinal processes at the L4-S2 level will reveal part of the sciatic nerve as it branches to form the L4, L5 and L6 spinal nerves. The L6 transverse process is carefully removed with a small rongeur enabling visualization of these spinal nerves. The L5 spinal nerve is isolated and tightly ligated with 7-0 silk suture. The wound is closed with a single muscle suture (6-0 silk) and one or two skin closure clips and dusted with antibiotic powder. In sham animals the L5 nerve is exposed as before but not ligated and the wound closed as before.

[0238] Male Wistar rats (120-140 g) are used for each procedure. Mechanical hyperalgesia is assessed by measuring paw withdrawal thresholds of both hindpaws to an increasing pressure stimulus using an Analgesymeter (Ugo-Basile, Milan). Mechanical allodynia is assessed by measuring withdrawal thresholds to non-noxious mechanical stimuli applied with von Frey hairs to the plantar surface of both hindpaws. Thermal hyperalgesia is assessed by measuring withdrawal latencies to a noxious thermal stimulus applied to the underside of each hindpaw. With all models, mechanical hyperalgesia and allodynia and thermal hyperalgesia develop within 1-3 days following surgery and persist for at least 50 days. Drugs may be applied before and after surgery to assess their effect on the development of hyperalgesia, or approximately 14 days following surgery to determine their ability to reverse established hyperalgesia.

B. RT-PCR mRNA analysis

[0239] One microgram of total RNA samples from the Chung model (L4 and L5 DRG) and sham-operated animals are used for first-strand cDNA synthesis using 50 pmol of oligo (dt) 24 primer in a 20 μL total reaction with 200 units Superscript II (LTI). The cDNA is then diluted to 100 μL with Tris-EDTA buffer (10 mM TrisCl, pH 8.0 and 1 mM EDTA). Three μL of the diluted cDNA is used to amplify the message for TRPV3 with gene-specific primers (sequences in 5' to 3' orientation: TRPV3 forward primer,

30 CTCATGCACAAGCTGACAGCCT (SEQ ID NO: 79); TRPV3 reverse primer, AGGCCTCTTCCGTGTACTCAGCGTTG (SEQ ID NO: 80)) in a 15 μL PCR reaction

using NotStart Taq DNA polymerase (Qiagen) for 25-38 cycles. Neuropeptide Y (NPY) is used as positive control.

[0240] For normalization 1 μL of the diluted cDNA is used to amplify actin using the following primers:

5'actin primer: ATC TGG CAC CAC ACC TTC TAC AA (SEQ ID NO: 81) 3'actin primer: GCC AGC CAG GTC CAG ACG CA (SEQ ID NO: 82)

[0241] A portion of the samples are then analyzed on a 4-20 TBE Criterion polyacrylamide gel (BioRad), stained with SYBR GREEN I (Molecular Probes) and

visualized on a Phosphorimager.

[0242] Figure 1A shows the average fold regulation of TRPV3 (VRLx) in L4 and L5 DRG neurons from the Chung model from three independent experiments. As shown in Figure 1A the positive control, NPY and TRPV3 message are elevated in the injured DRG relative to sham and non-ligated DRGs.

EXAMPLE 5

15 Identification of TRPV4

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[0243] Primers are designed to amplify distinct regions of the candidate genes that had been identified through the computer model. Based on the human sequence obtained, PCR primers are designed to also amplify the mouse homologue of TRPV4 (mTRPV4) (TRPV4 forward: CTCATGCACAAGCTGACAGCCT (SEQ ID NO: 83); TRP4 reverse: AGGCCTCTTCCGTGTACTCAGCGTTG (SEQ ID NO: 84)). These PCR products are subsequently sequenced and the mouse EST database is searched using these sequences. One EST clone (ID No. AI510567) is identified and obtained from the IMAGE consortium. The EST is further characterized and found to contain a ~2.4 kb insert which is sequenced. Primers are designed from this sequence and used to obtain the full length cDNA using the RACE protocol (Clontech). Both 5' and 3' RACE products are obtained and sequenced. This approach results in the amplification of the full length cDNA of mTRPV4 from mouse kidney and DRG cDNA using primers designed from the very 5' and 3' end of the RACE products. All primers utilized in the characterization of mTRPV4 are shown in Table 2. A novel full length cDNA of ~3.2 kb is identified, which includes an open-reading frame of ~2.5 kb, a 5' UTR consisting of ~145 bp and a 3' UTR encompassing ~400-500 nucleotides. The gene encodes a 3.4 kb transcript that contains three ankryin-repeat regions and TM6

domains. The protein sequence includes ~1000 amino acids and is set forth in SEQ ID NO: 14. Clustal W alignments to the rat VR (GenBank Ascession No. AF029310) reveals 34% identity and 64% similarity to VR1 in the region spanning the Ank2 through the TM4 region.

Table 2: TRPV4 Primers

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		SEQ ID NO:
Primers used for	RACE	
3' RACE	CCCTGGGCTGGGCGAACATGCTCTA	85
VR3RACE5'	CTTGGCAGCCATCATGAGAGGCGAA	86
Primers to ampli	ify partial/full length TRPV4	
AP19	GCAGTGGTAACAACGCAGAG	87
AP20	AGGTCAGATCTGTGGCAGGT	88
AP21	CGTGAGGTGACAGATGAGGA	89
AP32	CCAGTATGGCAGATCCTGGT	90
AP25	ATGGCAGATCCTGGTGATG	91
1706 66		92
AP26 CC	CCAGGCACTACTGAGGACT	93
AP27 AC	GGGCTACGCTCCCAAGT	94
		95
AP28_G]	<u> </u>	
AP22	TGAACTTGCGAGACAGATGC	

[0244] A combination of RT-PCR and Northern blot analyses are utilized to characterize expression of TRPV4. Total RNA is prepared from adult mouse kidney, newborn DRG and adult trigeminal tissue. RT-PCR is carried out using cDNA prepared from these three mouse tissues and primers within the ankyrin and the TM domain of mTRPV4. The expected 403 bp product is observed in all three tissues. This PCR product also serves as a probe on Northern blots (Clontech MTN blots). The expected 3.4 kb transcript is observed in kidney and other tissues.

[0245] The genomic structure of hTRPV4 is predicted from the high throughput genomic sequence database (GenBank Accession No. AC007834). HVR3 encompasses ~17 exons. A comparison of the amino acid sequence of the rat VR1 sequence (GenBank Accession No. AF029310) and the mouse VR3 protein reveals 34% identity and 64% similarity in the sequence spanning the ankryin 2 region and the TM4 domain. The nucleotide and amino acid sequences of hTRPV4 are shown in SEQ ID NO: 16 and SEQ ID NO: 17, respectively.

EXAMPLE 6

Gene Expression Analysis of TRPV4 in the Rat Chung Model

[0246] These studies discussed below measure relative levels of RNA expression for TRPV4 in the Chung neuropathic pain model using RT-PCR.

A. Spinal nerve ligation (Chung) model

[0247] This model is established according to the methods described by Kim and Chung, *supra*, and is described in Example 4.

B. RT-PCR mRNA analysis

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[0248] One microgram of total RNA samples from the Chung model (L4 and L5 DRG) and sham-operated animals are used for first-strand cDNA synthesis using 50 pmol of oligo (dt) 24 primer in a 20 μL total reaction with 200 units Superscript II (LTI). The cDNA is then diluted to 100 μL with Tris-EDTA buffer (10 mM TrisCl, pH 8.0 and 1 mM EDTA). Three μL of the diluted cDNA is used to amplify the message for TRPV4 with gene-specific primers (Sequences in 5' to 3' orientation: TRPV4 forward primer, 99

TGAGGATGACATAGGTGATGAG 120 (SEQ ID NO: 96), TRPV4 reverse primer, 255 CCAAGGACAAAAAGGACTGC 236 (SEQ ID NO: 97)) in a 15 μL PCR reaction using NotStart Taq DNA polymerase (Qiagen) for 25-38 cycles. NPY is used as positive control.

[0249] For normalization 1 μL of the diluted cDNA is used to amplify actin using the following primers:

5'actin primer: ATC TGG CAC CAC ACC TTC TAC AA (SEQ ID NO: 81)

3'actin primer: GCC AGC CAG GTC CAG ACG CA (SEQ ID NO: 82)

[0250] A portion of the samples are then analyzed on a 4-20 TBE Criterion polyacrylamide gel (BioRad), stained with SYBR GREEN I (Molecular Probes) and visualized on a Phosphorimager.

[0251] First-strand cDNA from the Chung model (50 days post-ligation) is normalized using a house-keeping gene; beta-actin. Figures 1A and 1B shows the expression of TRPV4 and NPY in the Chung Model (50- and 28-day post-ligation, respectively). The positive control, NPY and TRPV4 message are elevated in the injured DRG relative to sham and non-ligated DRGs. Accordingly, TRPV4 serves as a target for neuropathic pain.

EXAMPLE 7

Identification of VR TRPM8

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[0252] To identify novel TRP channels, genomic DNA databases are searched by constructing a HMM from the known TRP protein sequences of different mammalian species. With this model, the 6-frame translation of all available human sequences is queried and identifies multiple novel putative exons with similarity to the TM4 and TM6 domains of VR1. A fragment of the mouse homologue of one novel TRP channel is amplified by RT-PCR from mouse DRG RNA. Full-length sequence of this gene is derived from a combination of exon-prediction software, PCR and RACE amplification from newborn mouse DRGs.

[0253] For PCR cloning, primers 163f (5'-CAAGTTTGTCCGCCTCTTTC (SEQ ID NO: 98)) and 164r (5'-AACTGTCTGGAGCTGGCAGT (SEQ ID NO: 99)) are designed from the HMM sequences for TRPM8 as a result of blast hits and used to amplify a 699-nucleotide fragment of TRPM8 from newborn DRG cDNA. From this initial sequence and exon prediction programs, RACE PCR (Clontech) is used to obtain the 5' and 3' ends of TRPM8 from mouse newborn DRG cDNA following the manufacturer's protocol. Primers used in these experiments are shown in Table 3.

Table 3: Primers to Amplify Mouse TRPM8 cDNA

Table 3. Trimers to A	mpmy Mouse TRI Mo CDIVA	SEQ ID NO:
Putative trp candidate		
2KMHMR5R44-MOD C	ELERA HUMAN CONTIG	
FOR MOUSE:		
Probes designed for in situ	hyb analysis	
AP163F	CAAGTTTGTCCGCCTCTTTC	100
AP164R	ACTGCCAGCTCCAGACAGTT	101
Rapid amplification of cDN	NA ends (RACE)	
5' RACE primers		
5' RACE (nested)	ccttcgatgtgctggctctgggcataa	102
5' RACE	CCTTGCCTTTCTTCCCCAGAGTCTCAA	103
AP220 5' RACE	GCAAAGTTTTTGGCTCCACCCGTCA	104
AP2215' RACE (nested)	GCCAGTGCTGGGTCAGCAGTTCGTA	105
3' RACE primers		
3' RACE I	TTCAGGAGGTCATGTTCACGGCTCTCA	106
3' RACE I (nested)	GTACCGGAACCTGCAGATCGCCAAGA	107
AP218 3'RACE TRPXII	GCAAGATCCCTTGTGTGGTGGTGGA	108
AP219 3' (nested)	CAGCCTGGTGGAGGTGGAGGATGTT	109
3' RACE #3	CGGAACCTGCAGATCGCCAAGAACT	110
3' RACE primer in TM5 regu	ion of TRPM8	
AP225	GCGTGGCCAGACAGGGGATCCTAAG	111
3' REVERSE primer in TM5	region of TRPM8	
AP226	CCACACAGCAAAGAGGAACA	112
To amplify longer piece of m	ouse TRPM8	
216F	GGAGCCGCAGAAATGGTACT	113
Primers used for Northern p	robe	
Amplifies around 1.2 kB ba	<u>ınd</u>	
AP258	TCTCATTGGCCTCATTTCTG	114
AP247	ATATGAGACCCGAGCAGTGG	115

[0254] The protein TRPM8, has been named following the nomenclature suggested in Clapham et al., *Cell*, 108:595-598 (2001). Several human ESTs, many of which have been isolated from various cancer tissues, contain fragments of TRPM8 (Genbank GI Nos. 8750489, 9149390, 9335992 and 2223353).

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[0255] Translation of the nucleotide sequence of TRPM8 predicts a protein composed of 1104 amino acid residues (see SEQ ID NO: 8). The overall sequence of mouse

TRPM8 is 93% identical to that of the human gene (see Figure 6A). Its closest relative is TRPM2 (42% identity) (see Figures 6A and 6B). TRPM8 belongs to the "long" or Melastatin subfamily of TRP channels, a group of TRPs characterized by their lack of ankyrin domains in the N-terminus. TRP channels are predicted to contain TM6 domains, although at least one is predicted to have seven membrane-spanning domains (see Nagamine et al., *Genomics*, 54:124-131 (1998)). A Kyte-Doolittle plot suggests the presence of eight distinct hydrophobic peaks in TRPM8 sequence, which could represent six to eight predicted transmembrane domains. Overall, the predicted transmembrane domains are within amino acids 695-1024 of TRPM8. Outside of this region, the only predicted secondary structures are coiled-coil domains present both in the N- and C-terminal portion of the protein (data not shown) (see Burkhard et al., *Trends Cell. Biol.*, 11:82-88 (2001)). Coiled-coil domains are implicated in oligomerization of GABA-B channels, and have been previously predicted in some TRP channels (see Funayama et al., *supra*; and Margeta-Mitrovic et al., *Neuron*, 27:97-106 (2000)).

15 EXAMPLE 8

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Localization of TRPM8 expression

A. Northern blot analysis

[0256] Northern blots are made as followed: Total RNA are purified from mouse newborn and adult tissues using TRIzol LS (Invitrogen/Gibco Life technologies), followed by polyA⁺ purification with Oligotex (Qiagen) according to the manufacturer's protocols. Approximately 3 mg of sample are electrophoresed on 1% glyoxal gels, transferred and hybridized at high-stringency with a ³²P-labeled probe representing nucleotides 1410-1980 of the mouse full-length TRPM8 sequence. Commercial Northern blots (Clontech) are hybridized with the same TRPM8 probe. Blots are hybridized for 3 hours at 68°C in ExpressHyb hybridization solution (Clontech) and washed according to the manufacturer's high-stringency washing protocol and exposed to a phosphoimager screen for 1-3 days.

[0257] The results from this analysis are described below. No TRPM8 expression is detected using commercial Northern blots. Blots from newborn and adult mice are used that include tissues relevant for somatic sensation, including DRG, spinal cord and different

sources of skin. One mRNA species of approximately 6.3 kb is present predominantly in DRGs.

B. In situ hybridization

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[0258] For *in situ* hybridizations, newborn and adult tissues are dissected, fixed in 4% paraformaldehyde in PBS, cryoprotected and frozen in liquid nitrogen in OCT mounting medium. Cryostat sections (10 μm) are processed and hybridized with a digoxygenin cRNA probe generated by *in vitro* transcription (Roche Biochemicals). The mouse TRPM8 mRNA-specific antisense riboprobe corresponds to nucleotides 1410-1980 of the mTRPM8 sequence. Fluorescence detection and double-labeling experiments are carried out with the tyramide signal amplification kit (TSA; NEN) essentially as previously described (see Dong et al., *Cell*, 106:619-632 (2001)).

[0259] Digoxygenin-labeled probes show specific expression in DRG and trigeminal ganglia (cranial sensory neurons innervating the mouth and jaw) in newborn and adult mouse, but not in day 13 embryos. TRPM8 expression is restricted to approximately 5-10% of adult DRG neurons. The average size of the neurons positive for TRPM8 is $18 \pm$ 3.1 µm (mean ± standard deviation, n=69), and can be classified as small-diameter c-fibercontaining neurons, which in mouse are defined as smaller than 25 µm. TRPM8 is not expressed in heavily-myelinated neurons marked by Neurofilament (NF) antibodies, which correlates well with TRPM8 expression in small-sized neurons. TRPM8⁺ neurons thus appear to belong to a subset of nociceptive or thermoceptive neurons that express trkA, an NGF receptor, during development (see Huang and Reichardt, Ann. Rev. Neurosci., 24:677-736 (2001)). In the absence of NGF or trkA, DRG neurons that normally express this receptor die through apoptosis during embryonic development (Huang and Reichardt, supra). To prove that TRPM8 is expressed in trkA-dependent neurons, TRPM8 expression is evaluated in DRGs from newborn trkA-null mice. The expression of TRPM8 is completely abolished in the mutant ganglia. In addition, TRPM8 is not co-expressed with VR1, which marks a class of nociceptors that respond to capsaicin and noxious heat. This observation is confirmed by the lack of TRPM8 co-expression with either CGRP or IB4, two well-characterized antigenic markers found on nociceptive neurons (see Snider and McMahon, Neuron, 20:629-632 (1998); Tominaga et al., Neuron, 21:531-543 (1998)). This data strongly indicates that TRPM8 is expressed in a subpopulation of

thermoceptive/nociceptive neurons distinct from the well-characterized heat and pain sensing neurons marked by VR1, CGRP or IB4.

[0260] Following *in situ* hybridization, immunofluorescence is performed with anti-CGRP (1:100; Biogenesis), IB-4 (10 μg/mL; Sigma), anti-VR1 (1/2000; Abcam), anti-NF150 (1/1000; Chemicon) and detected with FITC or CY3 (10 μg/mL; Jackson Immunoresearch). Although all panels shown in these studies demonstrate lack of coexpression, this is not due to technical issues since additional probes/antibodies are used as controls to ensure our double-labeling protocol with the TRPM8 *in situ* probe is working.

EXAMPLE 9

10 Activation of TRPM8 Protein by Cold and Menthol

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A. Effect of heat, capsaicin, cold and menthol upon intracellular calcium

[0261] Given the similarity of TRPM8 protein to TRPV family members and its unique expression pattern, the effects of heat, capsaicin, cold and menthol in mediating calcium influx are examined using transfected CHO-K1/FRT cells expressing TRPM8 protein and a fluorescent calcium imaging method as described in detail below.

[0262] To generate mouse TRPM8-expressing CHO cell lines, mouse TRPM8 cDNA are subcloned in pcDNA5 (Invitrogen), transfected into CHO-K1/FRT cells using Fugene 6 (Roche). The transfected cells are selected by growth in MEM medium containing 200 μ g/ μ L⁻¹ hygromycin (Gibco BRL). Populations are frozen at early passage numbers and these stocks are used for further studies. Stable clones that express the mRNAs are identified by Northern blot analysis as well as Southern blotting to confirm integration site (not shown). CHO cells do not express an endogenous TRPM8 isoform and therefore serve as a control along with a cell line stably transfected with a VR1-expressing plasmid.

[0263] Calcium imaging experiments are performed essentially as previously described (see Savidge et al., *Neuroscience*, 102:177-184 (2001)). Briefly, cells are plated on glass coverslips and loaded with Fura-2 acetoxymethyl ester (2.5-5 mM) and incubated for 30-60 minutes at room temperature in 1.5 mM of pluronic acid (Molecular Probes, Eugene, OR) in a HEPES-buffered saline (2 mM Ca²⁺). Coverslips are placed in a laminar flow perfusion chamber (Warner Instrument Corp.) and constantly perfused with HEPES-buffered saline (2 mM Ca²⁺) via a local perfusion pipette through which buffer and chilled

solutions are also applied. Chilled stimulations consist of a linear decrease (~1-1.5°C sec⁻¹) in perfusate temperature from 33°C to 10°C. Perfusate temperature is controlled by a regulated Peltier device and is monitored in the cell chamber by a miniature thermocouple. Alternatively, cells are plated on 24-well tissue culture plates, loaded with Fura-2 and application of solutions is performed with a 3 cc syringe over a period of 10 seconds. Images of Fura-2 loaded cells with the excitation wavelength alternating between 340 and 380 nM are captured with a cooled CCD camera. Following subtraction of background fluorescence, the ratio of fluorescence intensity at the two wavelengths is calculated. Ratio levels in groups of 20-40 individual cells are analyzed using MetaFluor (Universal Imaging Corporation). All graphs are averaged responses from groups of 20-30 individual cells from representative single experiments. All experiments are repeated on three separate occasions and similar results obtained. Hanks balanced salt solution (HBSS), phosphate buffered saline (PBS) and all cell culture reagents are obtained from Gibco BRL. Ruthenium red, capsaicin and menthol are obtained from Sigma.

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[0264] The results of the above calcium imaging experiments are described below. Capsaicin (10 μM), an activator of VR1, does not evoke a response in TRPM8 expressing cells. Neither hypo-osmotic solutions, known to generate Ca²⁺ responses in TRPV3-expressing cells, or hypertonic buffer elicit a response in TRPM8 expressing cell lines (see Liedtke et al., *supra*; and Strotmann et al., *supra*)). An increase in temperature (25-50°C), a potent stimulus for VR1, also does not alter intracellular calcium levels. However, when the temperature is lowered from 25°C to 15°C, an increase in intracellular calcium is observed in TRPM8 expressing cells (Figures 7A and 8A). This response is not observed in non-transfected CHO cells or the VR1-expressing cell line (Figures 7A and 8A). Addition of a 10°C stimulus also evokes an influx of Ca²⁺. This response is dependent on Ca²⁺ in the buffer, because removal of extracellular calcium suppresses the temperature response (Figures 7A and 8A). The dependence on outside calcium is indicative of a cation-permeable channel localized at the plasma membrane. A potent blocker of the heat response for VR1, ruthenium red (at 5 μM), does not suppress the temperature response.

[0265] Since TRPM8 responds to a decrease in temperature, additional experiments are carried out to investigate the temperature threshold at which intracellular calcium ([Ca²⁺]_i) begins to rise in TRPM8 expressing cells. Cells are incubated at 35°C (normal skin temperature) for several minutes followed by a decrease in temperature to

13°C. The temperature response in mouse TRPM8-CHO cells shows a threshold of 22-25°C at which $[Ca^{2+}]_i$ starts to increase (Figure 7B), followed by a marked increase when the temperature of the buffer reached ~15°C. These experiments indicate that at physiological relevant temperatures, the upper activation threshold for TRPM8 is ~23°C (Figure 7C).

[0266] Menthol, a compound commonly used for its cooling properties, is tested as a stimulus on TRPM8 expressing CHO cells. Non-transfected CHO cells are completely insensitive to menthol (tested up to 1 mM) (Figure 7D). However, upon treatment of TRPM8 cells (incubated at 25°C), intracellular fluorescence increases significantly within seconds in response to menthol concentrations of 10 and 100 μM (Figure 7D). Additionally, as with the temperature stimulus, depletion of calcium from the extracellular buffer suppresses the calcium response (Figure 7D). The effect that menthol has at different temperatures is also examined. Incubation of TRPM8 expressing cells at 33°C, reveals that 10 μM menthol does not induce a calcium response as observed at 25°C, but upon lowering the temperature to 30°C, intracellular calcium levels increases (Figure 7E). Menthol thus appears to mimic the effect of lowering the temperature on TRPM8 expressing cells.

B. Effect of cold and menthol upon conductance

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[0267] To investigate the membrane responses to cold and menthol, voltage clamp experiments are carried out on TRPM8 expressing cells which are prepared as described above.

[0268] Cells are plated onto poly-D-lysine coated cover-slips for recording purposes and recordings undertaken 18-24 hours later. Experiments are carried out at room temperature using whole-cell voltage clamp technique, with an Axopatch 2B amplifier, filtered at 5 kHz and pClamp suite of software (Axon Instruments). Series resistant compensation is 80% for all experiments, using 2-5 MΩ fire-polished pipettes. Recording solutions are as follows; pipette solution for all experiments is (mM) CsCl, 140; CaCl₂, 1; EGTA, 10; HEPES, 10; MgATP, 2; titrated to pH 7.4 with CsOH. For menthol and cold activated currents the bath solution is (mM): NaCl, 140; KCl, 5; Glucose; 10, HEPES, 10; CaCl₂, 2; MgCl₂, 1; titrated to pH 7.4 with NaOH. Current-voltage relationships are used to evaluate reversal potentials with voltage ramps from -100 to +60 mV (2 second duration). For the permeability studies for the monovalent ions the NaCl in a simplified bath solution (mM): NaCl, 140; Glucose; 10, HEPES, 10; CaCl₂, 2; MgCl₂, 1, is substituted by either

equimolar CsCl or KCl (titrated with CsOH or KOH). For calcium permeability estimates, the bath solutions contains (mM) NaCl, 100; Glucose, 10 mM; Hepes, 10 mM (titrated with NaOH) plus 1 or 30 mM CaCl₂. Osmolarity of solutions are adjusted by addition of sucrose. Permeability ratios for the monovalent cations to Na (P_X/P_{Na}) are calculated as follows:

 $P_X/P_{Na} = E_{shift} = \{RT/F\} \log (P_X/P_{Na}[X]_O/[Na]_O)$

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where F is Faraday's constant, R is the universal gas constant and T is absolute temperature. For measurements of calcium permeability P_{Ca}/P_{Na} is calculated as follows:

 $E_{\text{shift}} = \{RT/F\} \log \{[\text{Na}]_{\text{O}} + 4B'[\text{Ca}]_{\text{O}(2)}\} / \{[\text{Na}]_{\text{O}} 4B'[\text{Ca}]_{\text{O}(1)}\}$

where B'= P'_{Ca}/P_{Na} and $P'_{Ca}=P_{Ca}/(1+e^{EF/RT})$ and $[Ca]_{O(1)}$ and $[Ca]_{O(2)}$ refer to the two different calcium concentrations. Local perfusion of menthol is via a TC^2 bip temperature controller. A Marlow temperature controller is used for the cooling ramps.

[0269] The results of the voltage clamp studies carried out on TRPM8 expressing cells are described below. Temperature ramps from 35°C to 7-13°C evoke inward currents at a holding potential of -60 mV and outward currents at +40 or +60 mV. Currents increase in amplitude as the temperature is lowered and usually show some degree of desensitization at the coldest temperatures tested <10°C (Figure 9A). The temperature threshold for current activation shows no dependence on membrane potential and individual cells activated at temperatures between 19°C and 25°C, with a mean threshold of 21.79 \pm 0.64°C (n=5). Analysis of the current-voltage relationships of the response to a cold stimulus with CsCl filled recording pipettes and a typical NaCl-based external solution reveals an outwardly rectifying current with a reversal potential (E_{rev}) close to 0 mV which is typical of a non-selective cation channel (Figure 9B).

[0270] Application of menthol evokes rapidly activating currents in TRPM8 expressing, but not in non-transfected CHO cells at temperatures above the threshold for cold activation (>23°C, Figure 10A). The menthol activated current shows pronounced outward rectification (Figure 10B) with an E_{rev} of -9.28 \pm 0.75 mV (n=12) that is similar to the E_{rev} for the cold-activated current under the same ionic conditions. These currents could be inactivated by raising the temperature (see Figure 10A) suggesting that menthol shifts the threshold for activation to higher temperatures, which agrees with the calcium imaging experiments. To test this idea further, concentration-response curves for menthol-evoked currents at two temperatures (22°C and 35°C) are obtained using positive membrane potentials to increase the size of the currents (Figures 11A and 11B). The concentration-

response relationship is shifted to the left at the lower temperature with a marked increase in the maximum amplitudes (Figures 11A and 11B). Changes in E_{rev} are used to determine the ion selectivity of the menthol activated current. Isotonic replacement of the NaCl in the solution with KCl or CsCl, causes small positive shifts in E_{rev} indicating that the TRPM8 channel discriminates poorly between these cations (data not shown). From the changes in E_{rev} measured on individual cells (external NaCl to KCl gives a shift of +7.38 ± 1.43 mV, n=7; NaCl to CsCl gives a shift of +9.09 ± 0.36 mV, n=5) a permeability sequence of Cs>K>Na is calculated with P_{Cs}/P_{Na} = 1.43 and P_K/P_{Na} = 1.34. Relative calcium permeability is calculated from the E_{rev} values measured with different external calcium concentrations. Increasing the external calcium from 1-30 mM, in the absence of external Mg^{2+} ions, shifts E_{rev} by +11.67 ± 1.20 mV, which corresponds to P_{Ca}/P_{Na} = 0.97. Thus TRPM8 is permeable to the monovalent cations, Na, K and Cs as well as the divalent cation calcium.

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[0271] It is understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and scope of the appended claims. All publications, patents, and patent applications cited herein are hereby incorporated by reference for all purposes.

WE CLAIM:

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1. An isolated TRPV3 nucleic acid molecule comprising a member selected from the group consisting of:

- a) a polynucleotide that encodes a mouse TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO: 2;
- b) a polynucleotide that encodes a mouse TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO: 2;
- a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a mouse TRPV3 protein;
- d) a polynucleotide that encodes a human TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO 5;
- e) a polynucleotide that encodes a human TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO 5;
- f) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a human TRPV3 protein; and
- g) a polynucleotide that is complementary to a polynucleotide of a) through f).
- 2. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule is a polydeoxyribonucleic acid (DNA).
- 20 3. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule is a polyribonucleic acid (RNA).
 - 4. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule is a) or b) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 3.
- 5. The TRPV3 nucleic acid molecule of claim 4, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 65-2440 of SEQ ID NO: 1.

6. The TRPV3 nucleic acid molecule of claim 4, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 65-2440 of SEQ ID NO: 1.

- 7. The TRPV3 nucleic acid molecule of claim 4, wherein the first
 5 polynucleotide comprises a nucleotide sequence as set forth in nucleotides 65-2440 of SEQ
 ID NO: 1.
 - 8. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 6.
- 9. The TRPV3 nucleic acid molecule of claim 8, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 57-2432 of SEQ ID NO: 4.
 - 10. The TRPV3 nucleic acid molecule of claim 9, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 57-2432 of SEQ ID NO: 4.
 - 11. The TRPV3 nucleic acid molecule of claim 9, wherein the first polynucleotide comprises a nucleotide sequence as set forth in nucleotides 57-2432 of SEQ ID NO: 4.
- 12. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule is c) or f) and the polypeptide comprises one or more functional domains selected from the group consisting of:
 - a) an ankyrin domain;

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- b) a transmembrane region;
- c) a pore loop region; and
- d) a coiled-coil domain.
- 13. The TRPV3 nucleic acid molecule of claim 12, wherein the polypeptide comprises a pore loop region flanked by two transmembrane regions.

14. The TRPV3 nucleic acid molecule of claim 12, wherein the polypeptide comprises four ankyrin domains.

- 15. The TRPV3 nucleic acid molecule of claim 1, wherein the nucleic acid molecule further comprises a heterologous nucleic acid.
- 5 16. The TRPV3 nucleic acid molecule of claim 15, wherein the heterologous nucleic acid comprises a promoter operably linked to the TRPV3 polynucleotide.
 - 17. The TRPV3 nucleic acid molecule of claim 15, wherein the heterologous nucleic acid comprises an expression vector.

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- 18. A host cell that comprises a TRPV3 nucleic acid molecule of claim 15.
- 19. An isolated TRPV3 polypeptide comprising a member selected from the group consisting of:
 - a mouse TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO: 2;
 - b) a mouse TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO: 2;
 - c) one or more functional domains of a mouse TRPV3 protein;
 - a human TRPV3 protein comprising amino acid residues 1-791 of SEQ ID NO 5;
 - e) a human TRPV3 protein comprising amino acid residues 2-791 of SEQ ID NO 5; and
 - f) one or more functional domains of a human TRPV3 protein.
- 20. The TRPV3 polypeptide of claim 19, wherein the TRPV3 polypeptide is c) or f) and comprises one or more functional domains selected from the group consisting of:
 - a) an ankyrin domain;
 - b) a transmembrane region;
 - c) a pore loop region; and

d) a coiled-coil domain.

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- 21. The TRPV3 polypeptide of claim 20, wherein the polypeptide comprises a pore loop region flanked by two transmembrane regions.
- 22. The TRPV3 polypeptide of claim 20, wherein the polypeptidecomprises four ankyrin domains.
 - 23. An antibody that specifically binds to a TRPV3 polypeptide of claim 19.
 - 24. A method for identifying an agent that modulates TRPV3-mediated cation passage through a membrane, the method comprising:
 - a) providing a membrane that comprises a TRPV3 polypeptide of claim
 19;
 - b) contacting the membrane with a candidate agent; and
 - c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent.
 - 25. The method of claim 24, wherein the membrane comprises a cell and cation passage through the membrane is detected by measuring cation influx across the membrane into the cell.
- 26. The method of claim 25, wherein the cell comprises a promoter20 operably linked to a heterologous polynucleotide that encodes the TRPV3 polypeptide.
 - 27. The method of claim 24, wherein cation passage through the membrane is detected by voltage clamping.
 - 28. The method of claim 24, wherein cation passage through the membrane is detected by an ion sensitive dye or a membrane potential dye.
- 25. The method of claim 24, wherein the assay is conducted at a temperature of at least 33°C.

30. The method of claim 24, wherein the assay is conducted at a temperature of less than 52°C.

- 31. The method of claim 30, wherein the assay is conducted at a temperature of less than 43°C.
- 5 32. The method of claim 24, wherein the membrane is contacted with the candidate modulating agent in a well of a multiwell plate.
 - 33. The method of claim 32, wherein the multiwell plate is a 96-, 384- or 1536-well plate.
- 34. The method of claim 24, wherein a candidate agent that reduces cation
 10 passage is further tested for ability to treat pain by administering the candidate agent to a test
 animal and determining whether the candidate agent decreases the test animal's response to a
 pain stimulus.
 - 35. The method of claim 34, wherein the pain stimulus is exposure to a temperature above 33° C.
- 36. A method of reducing pain associated with TRPV3 activity, the method comprising administering to a subject suffering from pain an analgesically effective amount of a compound that reduces TRPV3-mediated cation passage through a membrane or reduces signal transduction from a TRPV3 polypeptide to a DRG neuron.
- 37. The method of claim 36, wherein the pain is associated with one or20 more of heat exposure, inflammation, or tissue damage.

- 38. The method of claim 36, wherein the compound is selected from the group consisting of:
 - a) an antibody that specifically binds to a TRPV3 polypeptide;
 - b) an antisense polynucleotide, ribozyme, or an interfering RNA that reduces expression of a TRPV3 polypeptide; and
 - c) a chemical compound that reduces cation passage through a membrane that comprises a TRPV3 polypeptide.

39. The method of claim 38, wherein the chemical compound has a molecular weight of 1000 daltons or less.

40. A method for determining whether pain in a subject is mediated by TRPV3, the method comprising:

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- a) obtaining a sample from a region of the subject at which the pain is felt; and
- b) testing the sample to determine whether a TRPV3 polypeptide or TRPV3 polynucleotide is present in the sample.
- 41. The method of claim 40, wherein the presence of a TRPV3 polypeptide

 in the sample is detected by determining whether cation passage across membranes of cells in the sample is mediated by a TRPV3 polypeptide.
 - 42. The method of claim 41, wherein TRPV3 involvement in mediating cation passage across membranes of the cells is determined by detecting an increase in cation passage across membranes of the cells when assayed above 33°C compared to cation passage when assayed below 33°C.
 - 43. The method of claim 40, wherein the presence of a TRPV3 polypeptide in the sample is detected by contacting the sample with a reagent that specifically binds to a TRPV3 polypeptide.
 - 44. The method of claim 43, wherein the reagent comprises an antibody.
- 20 45. The method of claim 40, wherein the presence of a TRPV3 polynucleotide in the sample is detected by contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPV3 polynucleotide.
 - 46. The method of claim 45, wherein the test polynucleotide comprises an oligonucleotide at least 10 nucleotides in length.
- 25 47. The method of claim 45, wherein the method comprises amplification of a TRPV3 polynucleotide, if present in the sample.

48. The method of claim 47, wherein the amplification comprises polymerase chain reaction or ligase chain reaction.

- 49. The method of claim 45, wherein the test polynucleotide is attached to a solid support.
- 50. The method of claim 49, wherein the solid support comprises a microchip.

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- 51. An isolated TRPV4 nucleic acid molecule comprising a member selected from the group consisting of:
 - a) a polynucleotide that encodes a mouse TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO: 14;
 - b) a polynucleotide that encodes a mouse TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO: 14;
 - c) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a mouse TRPV4 protein;
 - d) a polynucleotide that encodes a human TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO 17;
 - e) a polynucleotide that encodes a human TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO 17;
 - f) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a human TRPV4 protein; and
 - g) a polynucleotide that is complementary to a polynucleotide of a) through f).
- **52.** The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule is a polydeoxyribonucleic acid (DNA).
- 25 53. The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule is a polyribonucleic acid (RNA).

54. The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule is a) or b) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 15.

55. The TRPV4 nucleic acid molecule of claim 54, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 156-2771 of SEQ ID NO: 13.

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- 56. The TRPV4 nucleic acid molecule of claim 54, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 156-2771 of SEQ ID NO: 13.
- 57. The TRPV4 nucleic acid molecule of claim 56, wherein the first polynucleotide comprises a nucleotide sequence as set forth in nucleotides 156-2771 of SEQ ID NO: 13.
 - 58. The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 18.
 - 59. The TRPV4 nucleic acid molecule of claim 58, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 16.
 - 60. The TRPV4 nucleic acid molecule of claim 58, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 16.
 - 61. The TRPV4 nucleic acid molecule of claim 60, wherein the first polynucleotide comprises a nucleotide sequence as set forth in SEQ ID NO: 16.
- 62. The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule is c) or f) and the polypeptide comprises one or more functional domains selected from the group consisting of:
 - a) an ankyrin domain:

- b) a transmembrane region;
- c) a pore loop region; and
- d) a coiled-coil domain.

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- 63. The TRPV4 nucleic acid molecule of claim 62, wherein the polypeptide5 comprises a pore loop region flanked by two transmembrane regions.
 - 64. The TRPV4 nucleic acid molecule of claim 62, wherein the polypeptide comprises three ankyrin domains.
 - 65. The TRPV4 nucleic acid molecule of claim 51, wherein the nucleic acid molecule further comprises a heterologous nucleic acid.
 - 66. The TRPV4 nucleic acid molecule of claim 65, wherein the heterologous nucleic acid comprises a promoter operably linked to the TRPV4 polynucleotide.
 - 67. The TRPV4 nucleic acid molecule of claim 65, wherein the heterologous nucleic acid comprises an expression vector.
 - 68. A host cell that comprises a TRPV4 nucleic acid molecule of claim 65.
 - 69. An isolated TRPV4 polypeptide comprising a member selected from the group consisting of:
 - a) a mouse TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO: 14;
 - b) a mouse TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO: 14;
 - c) one or more functional domains of a mouse TRPV4 protein;
 - a human TRPV4 protein comprising amino acid residues 1-871 of SEQ ID NO 17;
 - e) a human TRPV4 protein comprising amino acid residues 2-871 of SEQ ID NO 17; and
 - f) one or more functional domains of a human TRPV4 protein.

70. The TRPV4 polypeptide of claim 69, wherein the polypeptide is c) or f) and comprises one or more functional domains selected from the group consisting of:

a) an ankyrin domain;

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- b) a transmembrane region;
- c) a pore loop region; and
- d) a coiled-coil domain.
- 71. The TRPV4 polypeptide of claim 70, wherein the polypeptide comprises a pore loop region flanked by two transmembrane regions.
- 72. The TRPV4 polypeptide of claim 70, wherein the polypeptide comprises three ankyrin domains.
 - 73. An antibody that specifically binds to a TRPV4 polypeptide of claim 69.
 - 74. A method for identifying an agent that modulates TRPV4-mediated cation passage through a membrane, the method comprising:
 - a) providing a membrane that comprises a TRPV4 polypeptide of claim 69;
 - b) contacting the membrane with a candidate agent; and
 - c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent.
 - 75. The method of claim 74, wherein the membrane comprises a cell and cation passage through the membrane is detected by measuring cation influx across the membrane into the cell.
- 76. The method of claim 75, wherein the cell comprises a promoter25 operably linked to a heterologous polynucleotide that encodes the TRPV4 polypeptide.
 - 77. The method of claim 74, wherein cation passage through the membrane is detected by voltage clamping.

78. The method of claim 74, wherein cation passage through the membrane is detected by an ion sensitive dye or a membrane potential dye.

- 79. The method of claim 74, wherein the membrane is contacted with the candidate modulating agent in a well of a multiwell plate.
- 5 80. The method of claim 79, wherein the multiwell plate is a 96-, 384- or 1536-well plate.

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- 81. The method of claim 74, wherein a candidate agent that reduces cation passage is further tested for ability to treat pain by administering the candidate agent to a test animal and determining whether the candidate agent decreases the test animal's response to a pain stimulus.
 - 82. The method of claim 81, wherein the pain is neuropathic pain.
- 83. A method of reducing pain associated with TRPV4 activity, the method comprising administering to a subject suffering from pain an analysesically effective amount of a compound that reduces TRPV4-mediated cation passage through a membrane or reduces signal transduction from a TRPV4 polypeptide to a DRG neuron.
 - 84. The method of claim 83, wherein the pain is neuropathic pain.
- 85. The method of claim 83, wherein the compound is selected from the group consisting of:
 - a) an antibody that specifically binds to a TRPV4 polypeptide;
 - b) an antisense polynucleotide, ribozyme, or an interfering RNA that reduces expression of a TRPV4 polypeptide; and
 - c) a chemical compound that reduces cation passage through a membrane that comprises a TRPV4 polypeptide.
- 86. The method of claim 85, wherein the chemical compound has a molecular weight of 1000 daltons or less.
 - 87. A method for determining whether pain in a subject is mediated by TRPV4, the method comprising:

a) obtaining a sample from a region of the subject at which the pain is felt; and

- b) testing the sample to determine whether a TRPV4 polypeptide or TRPV4 polynucleotide is present in the sample.
- 5 88. The method of claim 87, wherein the presence of a TRPV4 polypeptide in the sample is detected by determining whether cation passage across membranes of cells in the sample is mediated by a TRPV4 polypeptide.
- 89. The method of claim 87, wherein the presence of a TRPV4 polypeptide in the sample is detected by contacting the sample with a reagent that specifically binds to a TRPV4 polypeptide.
 - 90. The method of claim 89, wherein the reagent comprises an antibody.
 - 91. The method of claim 87, wherein the presence of a TRPV4 polynucleotide in the sample is detected by contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPV4 polynucleotide.
- 15 92. The method of claim 91, wherein the test polynucleotide comprises an oligonucleotide at least 10 nucleotides in length.
 - 93. The method of claim 91, wherein the method comprises amplification of a TRPV4 polynucleotide, if present in the sample.
- 94. The method of claim 93, wherein the amplification comprises polymerase chain reaction or ligase chain reaction.
 - 95. The method of claim 91, wherein the test polynucleotide is attached to a solid support.
 - 96. The method of claim 95, wherein the solid support comprises a microchip.
- 25 97. An isolated TRPM8 nucleic acid molecule comprising a member selected from the group consisting of:

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a) a polynucleotide that encodes a mouse TRPM8 protein comprising amino acid residues 1-1104 of SEQ ID NO: 8;

- b) a polynucleotide that encodes a mouse TRPM8 protein comprising amino acid residues 2-1104 of SEQ ID NO: 8;
- c) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a mouse TRPM8 protein;
- d) a polynucleotide that encodes a human TRPM8 protein comprising amino acid residues 1-1268 of SEQ ID NO 11;
- e) a polynucleotide that encodes a human TRPM8 protein comprising amino acid residues 2-1268 of SEQ ID NO 11;
- f) a polynucleotide that encodes a polypeptide that comprises one or more functional domains of a human TRPM8 protein; and
- g) a polynucleotide that is complementary to a polynucleotide of a) through f).
- 15 98. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleic acid molecule is a polydeoxyribonucleic acid (DNA).
 - 99. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleic acid molecule is a polyribonucleic acid (RNA).
 - 100. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleic acid molecule is a) or b) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 9.
 - 101. The TRPM8 nucleic acid molecule of claim 100, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7.
- 25 102. The TRPM8 nucleic acid molecule of claim 100, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7.

103. The TRPM8 nucleic acid molecule of claim 102, wherein the first polynucleotide comprises a nucleotide sequence as set forth in nucleotides 448-3762 of SEQ ID NO: 7.

104. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleic acid molecule is d) or e) and comprises a first polynucleotide 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in SEQ ID NO: 12.

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- 105. The TRPM8 nucleic acid molecule of claim 104, wherein the first polynucleotide is 80% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 61-4821 of SEQ ID NO: 10.
- 106. The TRPM8 nucleic acid molecule of claim 104, wherein the first polynucleotide is 90% or more identical to a second polynucleotide having a nucleotide sequence as set forth in nucleotides 61-4821 of SEQ ID NO: 10.
 - 107. The TRPM8 nucleic acid molecule of claim 106, wherein the first polynucleotide comprises a nucleotide sequence as set forth in nucleotides 61-4821 of SEQ ID NO: 10.
 - 108. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleic acid molecule is c) or f) and the polypeptide comprises one or more functional domains selected from the group consisting of:
 - a) a transmembrane region;
 - b) a pore loop region; and
 - c) a coiled-coil domain.
 - 109. The TRPM8 nucleic acid molecule of claim 108, wherein the polypeptide comprises a pore loop region flanked by two transmembrane regions.
- 110. The TRPM8 nucleic acid molecule of claim 97, wherein the nucleicacid molecule further comprises a heterologous nucleic acid.

111. The TRPM8 nucleic acid molecule of claim 110, wherein the heterologous nucleic acid comprises a promoter operably linked to the TRPM8 polynucleotide.

- 112. The TRPM8 nucleic acid molecule of claim 110, wherein the5 heterologous nucleic acid comprises an expression vector.
 - 113. A host cell that comprises a TRPM8 nucleic acid molecule of claim 97.
 - 114. An isolated TRPM8 polypeptide comprising a member selected from the group consisting of:
 - a) a mouse TRPM8 protein comprising amino acid residues 1-1104 of SEQ ID NO: 8;
 - b) a mouse TRPM8 protein comprising amino acid residues 2-1104 of SEQ ID NO: 8;
 - c) one or more functional domains of a mouse TRPM8 protein;
 - d) a human TRPM8 protein comprising amino acid residues 1-1268 of SEQ ID NO 11;
 - e) a human TRPM8 protein comprising amino acid residues 2-1268 of SEQ ID NO 11; and
 - f) one or more functional domains of a human TRPM8 protein.
- 115. The TRPM8 polypeptide of claim 114, wherein the nucleic acid
 20 molecule is c) or f) and the functional domains comprise one or more members selected from the group consisting of:
 - a) a transmembrane region;
 - b) a pore loop region; and
 - c) a coiled-coil domain.

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- 25 116. The TRPM8 polypeptide of claim 115, wherein the polypeptide comprises a pore loop region flanked by two transmembrane regions.
 - 117. An antibody that specifically binds to a TRPM8 polypeptide of claim 114.

118. A method for identifying an agent that modulates TRPM8-mediated cation passage through a membrane, the method comprising:

- a) providing a membrane that comprises a TRPM8 polypeptide of claim 114:
- b) contacting the membrane with a candidate agent; and

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- c) determining whether passage of one or more cations through the membrane is increased in the presence of the candidate agent compared to passage in the absence of the candidate agent.
- 119. The method of claim 118, wherein the membrane comprises a cell and cation passage through the membrane is detected by measuring cation influx across the membrane into the cell.
 - 120. The method of claim 119, wherein the cell comprises a promoter operably linked to a heterologous polynucleotide that encodes the TRPM8 polypeptide.
 - 121. The method of claim 118, wherein cation passage through the membrane is detected by voltage clamping.
 - 122. The method of claim 118, wherein cation passage through the membrane is detected by an ion sensitive dye or a membrane potential dye.
 - 123. The method of claim 118, wherein the membrane is contacted with the candidate modulating agent in a well of a multiwell plate.
- 20 **124.** The method of claim 123, wherein the multiwell plate is a 96-, 384- or 1536-well plate.
 - 125. The method of claim 118, wherein the assay is to identify antagonists of TRPM8-mediated cation passage and is conducted at a temperature of less than 20°C and/or in the presence of menthol.
- 25 126. The method of claim 125, wherein a candidate agent that reduces cation passage is further tested for ability to treat pain by administering the candidate agent to a test

animal and determining whether the candidate agent decreases the test animal's response to a pain stimulus.

- 127. The method of claim 126, wherein the pain stimulus is cold.
- 128. The method of claim 118, wherein the assay is to identify agonists of TRPM8-mediated cation passage and is conducted at a temperature of greater than 20°C.
 - 129. The method of claim 128, wherein an agonist of TRPM8-mediated cation passage is used as a fragrance or a flavor enhancer.
 - 130. A method of reducing pain associated with TRPM8 activity, the method comprising administering to a subject suffering from pain an analysesically effective amount of a compound that reduces TRPM8-mediated cation passage through a membrane or reduces signal transduction from a TRPM8 polypeptide to a DRG neuron.

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- 131. The method of claim 130, wherein the pain is associated with one or more of cold exposure, inflammation, or tissue damage.
- 132. The method of claim 130, wherein the compound is selected from the group consisting of:
 - a) an antibody that specifically binds to a TRPM8 polypeptide;
 - b) an antisense polynucleotide, ribozyme, or an interfering RNA that reduces expression of a TRPM8 polypeptide; and
 - c) a chemical compound that reduces cation passage through a membrane that comprises a TRPM8 polypeptide.
 - 133. The method of claim 132, wherein the chemical compound has a molecular weight of 1000 daltons or less.
 - 134. A method for determining whether pain in a subject is mediated by TRPM8, the method comprising:
- 25 a) obtaining a sample from a region of the subject at which the pain is felt; and

b) testing the sample to determine whether a TRPM8 polypeptide or TRPM8 polynucleotide is present in the sample.

135. The method of claim 134, wherein the presence of a TRPM8 polypeptide in the sample is detected by determining whether cation passage across membranes of cells in the sample is mediated by a TRPM8 polypeptide.

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- 136. The method of claim 135, wherein TRPM8 involvement in mediating cation passage across membranes of the cells is determined by detecting an increase or decrease in cation passage across membranes of the cells when assayed below 20°C and/or in the presence of menthol, compared to cation passage when assayed above 20°C and/or in the absence of menthol.
- 137. The method of claim 134, wherein the presence of a TRPM8 polypeptide in the sample is detected by contacting the sample with a reagent that specifically binds to a TRPM8 polypeptide.
 - 138. The method of claim 137, wherein the reagent comprises an antibody.
- 139. The method of claim 134, wherein the presence of a TRPM8 polynucleotide in the sample is detected by contacting nucleic acids from the sample with a test polynucleotide that can hybridize to a TRPM8 polynucleotide.
 - 140. The method of claim 139, wherein the test polynucleotide comprises an oligonucleotide at least 10 nucleotides in length.
- 20 141. The method of claim 139, wherein the method comprises amplification of a TRPM8 polynucleotide, if present in the sample.
 - 142. The method of claim 141, wherein the amplification comprises polymerase chain reaction or ligase chain reaction.
- 143. The method of claim 139, wherein the test polynucleotide is attached to 25 a solid support.

144. The method of claim 143, wherein the solid support comprises a microchip.

145. A method for identifying an agent useful in the modulation of a mammalian sensory response, the method comprising:

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- a) contacting a candidate agent with a test system that comprises a receptor polypeptide selected from the group consisting of TRPM8, TRPV3 and TRPV4; and
- b) detecting a change in activity of the receptor polypeptide in the presence of the candidate agent as compared to the activity of the receptor polypeptide in the absence of the agent, thereby identifying an agent that modulates receptor activity.
- 146. The method of claim 145, wherein the sensory response is response to cold and the polypeptide is a TRPM8 polypeptide.
- 147. The method of claim 146, wherein the TRPM8 polypeptide comprises an amino acid sequence as set forth in SEQ ID NO: 8 or SEQ ID NO: 11.
 - 148. The method of claim 145, wherein the sensory response is response to warm or hot temperatures and the polypeptide is a TRPV3 polypeptide.
 - 149. The method of claim 148, wherein the TRPV3 polypeptide comprises an amino acid sequence as set forth in SEQ ID NO: 2 or SEQ ID NO: 5.
- 20 **150.** The method of claim 145, wherein the sensory response neuropathic pain and the polypeptide is a TRPV4 polypeptide.
 - 151. The method of claim 150, wherein the TRPV4 polypeptide comprises an amino acid sequence as set forth in SEQ ID NO: 14 or SEQ ID NO: 17.
- 152. The method of claim 145, wherein the method further comprises
 administering the agent that modulates receptor activity to a test subject, and thereafter detecting a change in the sensory response in the test subject.

153. The method of claim 145, wherein the test system comprises a membrane that comprises the receptor polypeptide.

- 154. The method of claim 153, wherein the test system comprises a cell that expresses a heterologous polynucleotide that encodes the receptor polypeptide.
- 5 155. The method of claim 154, wherein the cell is substantially isolated and the contacting is performed *in vitro*.
 - 156. The method of claim 154, wherein the cell is present in an organism and the contacting is performed *in vivo*.
- 157. The method of claim 145, wherein the receptor activity comprises increased or decreased Ca²⁺ passage through the membrane that comprises the receptor polypeptide.
 - 158. The method of claim 157, wherein the membrane comprises a substantially purified cell membrane.
 - 159. The method of claim 157, wherein the membrane comprises a liposome.
- 15 160. A method for monitoring the efficacy of a treatment of a subject suffering from pain, the method comprising:

- a) obtaining, at two or more time points in the course of treatment for pain, a sample from a region of the subject at which the pain is felt;
 and
- b) testing the samples to determine whether a reduction is observed from one time point to another in amount or activity of one or more members selected from the group consisting of: a TRPV3 polypeptide, a TRPV3 mRNA, a TRPV4 polypeptide, a TRPV4 mRNA, a TRPM8 polypeptide, and a TRPM8 mRNA.
- 25 161. The method of claim 160, wherein one of the time points is prior to administration of the treatment for pain.

162. An assay capable of detecting the expression of one or more of TRPV3, TRPV4 or TRPM8 in human tissue, the assay selected from the group consisting of:

- an assay comprising contacting a human tissue sample with monoclonal antibodies binding to TRPV3, TRPV4 or TRPM8 and determining whether the monoclonal antibodies bind to polypeptides in the sample; and
- b) an assay comprising contacting a human tissue sample with an oligonucleotide that is capable of hybridizing to a nucleic acid that encodes TRPV3, TRPV4 or TRPM8.
- 163. The assay of claim 162, wherein the assay comprises contacting a human tissue sample with a pair of oligonucleotides that are capable of hybridizing to a nucleic acid that encodes TRPV3, TRPV4 or TRPM8 and subjecting the sample to polymerase chain reaction.

- 164. The assay of claim 162, wherein the assay comprises contacting a human tissue sample with an oligonucleotide array that comprises one or more oligonucleotides that are capable of hybridizing to a nucleic acid that encodes TRPV3, TRPV4 or TRPM8.
 - 165. The assay of claim 162, wherein the human tissue sample is obtained from a site of pain.
- 20 166. A method of treating pain, the method comprising identifying a patient suffering from pain mediated by one or more polypeptides selected from the group consisting of TRPV3, TRPV4 and TRPM8 by measuring expression of the polypeptide in tissue from such patient, and administering to such patient an analgesically effective amount of an agent which inhibits the polypeptide.
- 25 **167.** A method for identifying an agent useful in the treatment of pain, the method comprising:
 - a) administering a candidate agent to a mammal suffering from pain;
 - b) in a sample obtained from the mammal, detecting an activity or amount of one or more members selected from the group consisting

of: a TRPV3 polypeptide, a TRPV3 mRNA, a TRPV4 polypeptide, a TRPV4 mRNA, a TRPM8 polypeptide, and a TRPM8 mRNA; and

- c) comparing the amount or activity of the member in the presence of the candidate agent with the amount or activity of the member in a sample obtained from the mammal in the absence of the candidate agent, wherein a decrease in amount or activity of the member in the sample in the presence of the candidate agent relative to the amount or activity in the absence of the candidate agent is indicative of an agent useful in the treatment of pain.
- 168. A method of identifying an agent that binds to and/or modulates the activity of an mRNA or polypeptide encoded by a TRPV3, TRPV4, or TRPM8 nucleic acid, the method comprising:
 - a) contacting an isolated cell which expresses a heterologous TRPV3,
 TRPV4, or TRPM8 nucleic acid encoding a polypeptide with the agent; and
 - b) determining binding and/or modulation of the activity of the mRNA or polypeptide by the agent, to identify agents which bind with and/or modulate the activity of the polypeptide.

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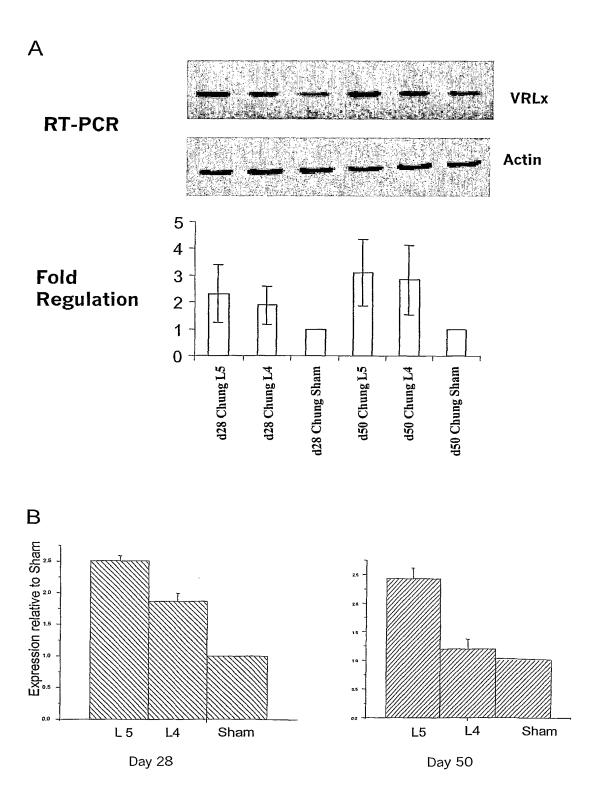
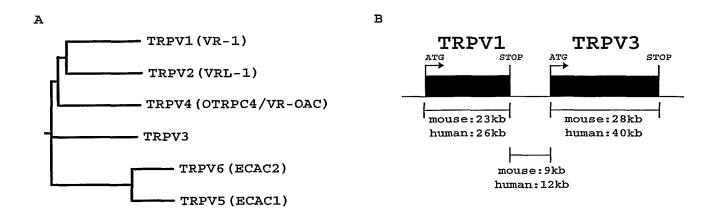


Figure 1



C

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TRPV4 206 ASYTEEARECGTALHIAIERRGCHYWELLYAQGADVHAQARGREEGCPKDEGGYEYEGELDLAAACTNOPHIYVGEHERD
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TRPV5 102 ESTLCEPFVGGTALHIAVINQNVNLVRALLARGASASARATGSAEHRS-SHNLIMYGEHPLSFAACVGSEELVRLIIERG TRPV1 272 WQPADISARDSVENTVLHALVEVADNTVDNIKFVISHKNEIJILGAKLHEILKEELITRKEITEIALAASSEKIEVLAV
TRPV2 229 HQPASLEATDSLEGHTVLHALVEVADNTVDNIKFVISHKNEIJILGAKLHEILKEELITRKEITEIALAASSEKIEVLAV
TRPV4 309 HKKADMERQDSRENTVLHALVATADNITENTKFVIKKVDLLLKCSREFEDSNLEIVLNNDEUSPLIMMAAATERIEVET
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Figure 2

Figure 2D

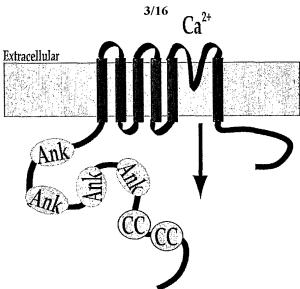


Figure 2E

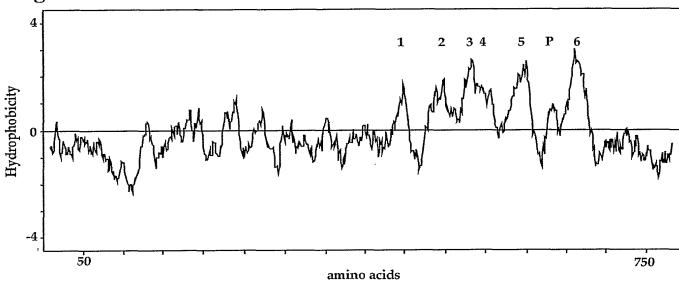
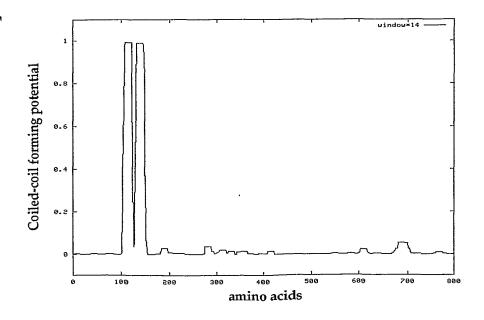
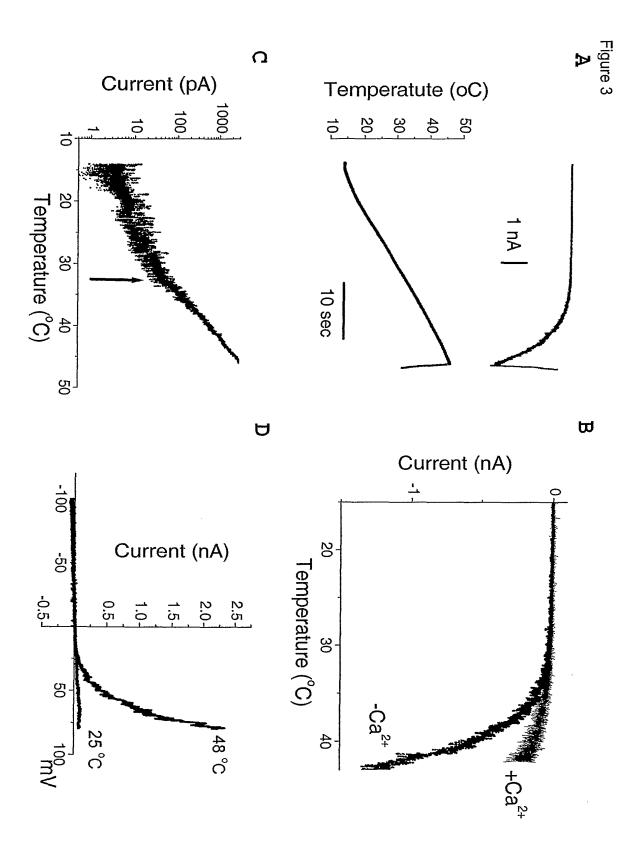


Figure 2F





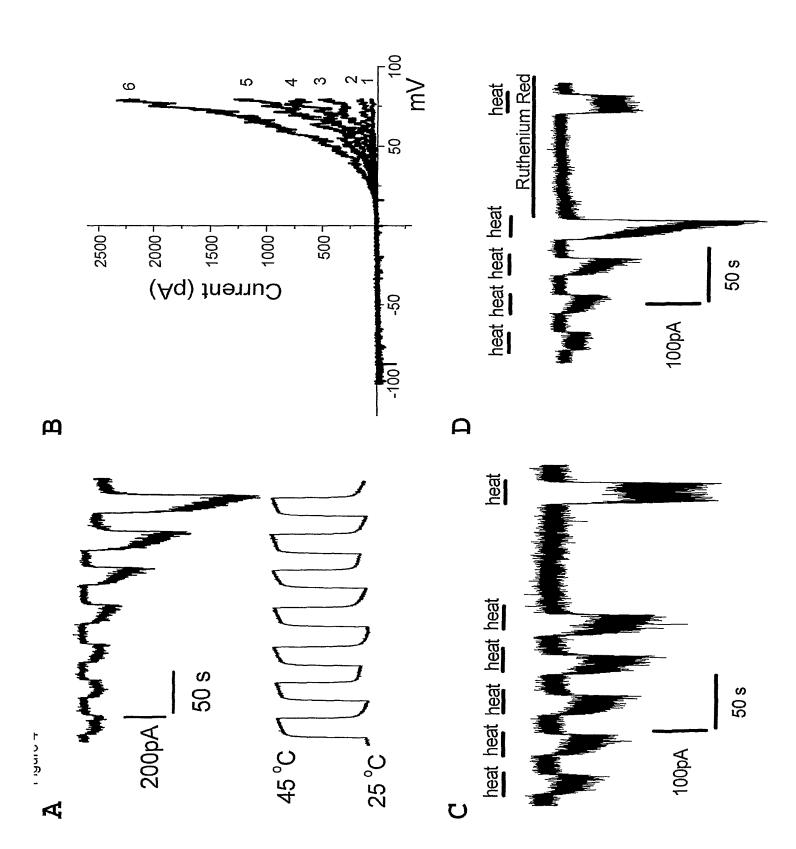


Figure 5

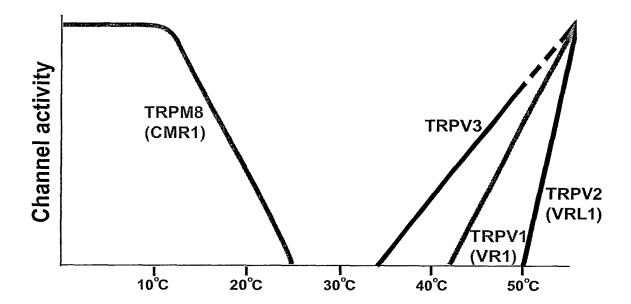


Figure 6A

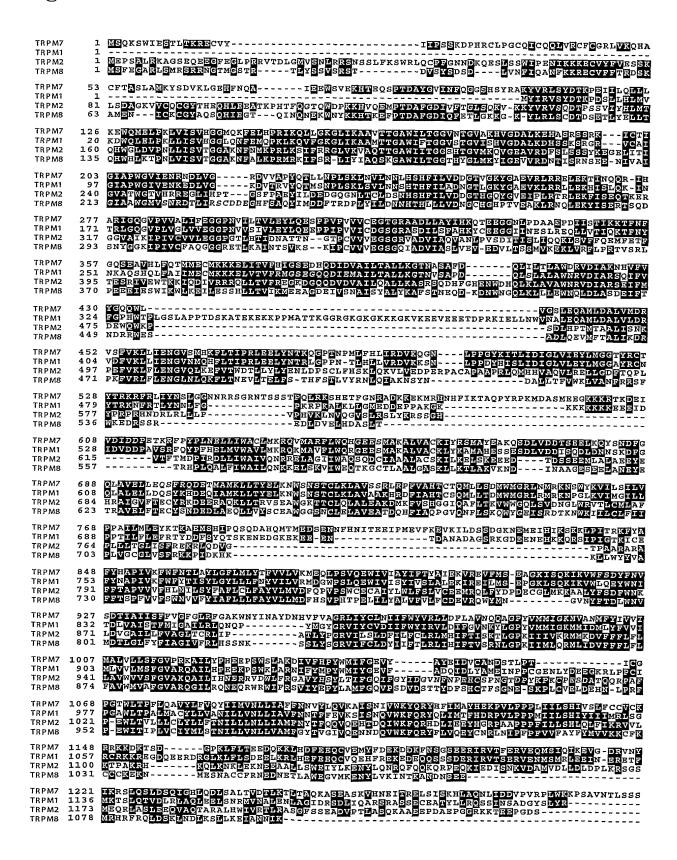


Figure 6B

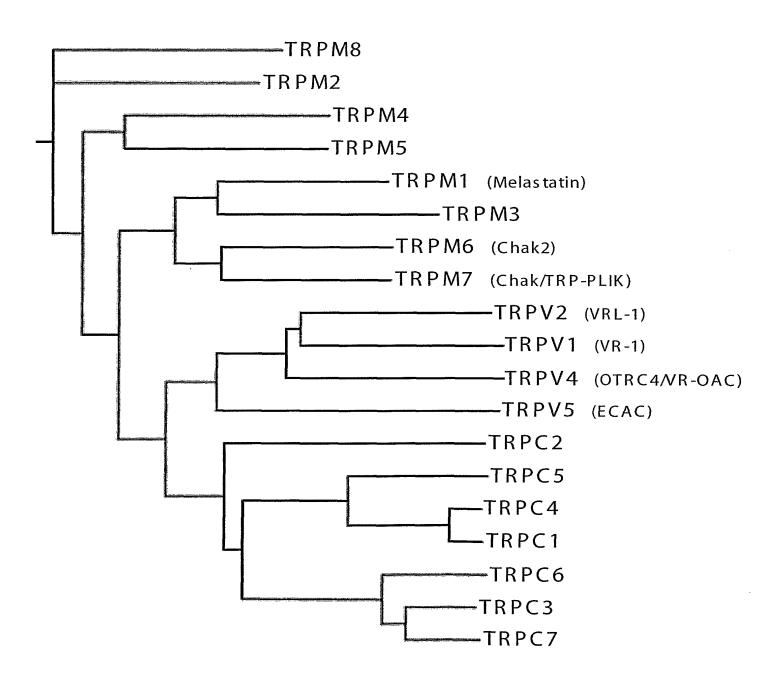


Figure 6C

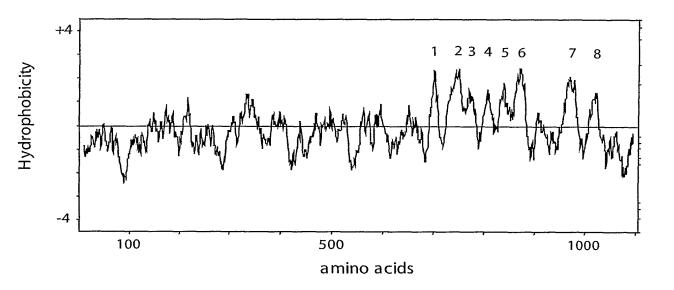


Figure 6D

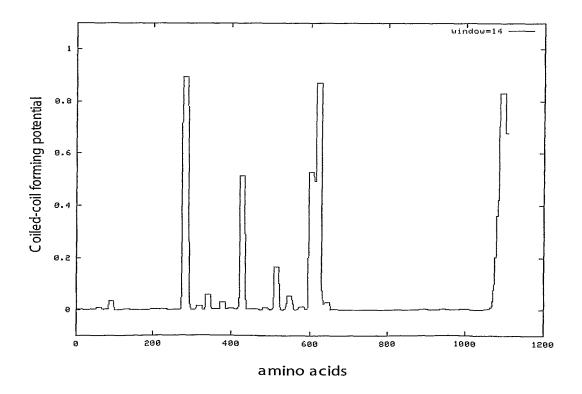


Figure 7A

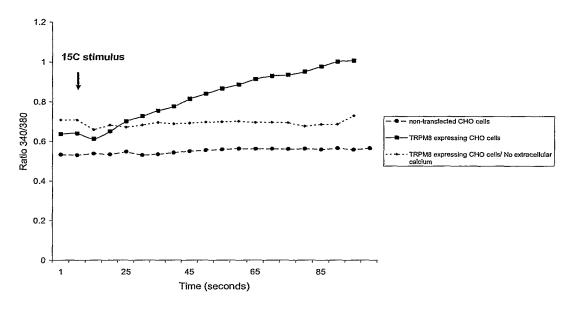


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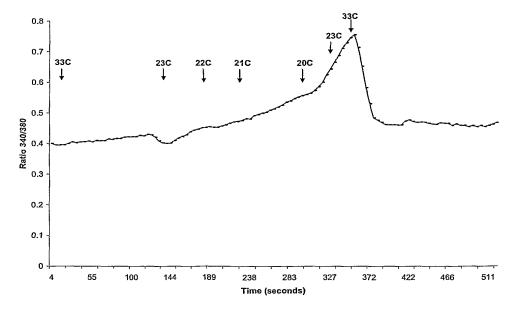


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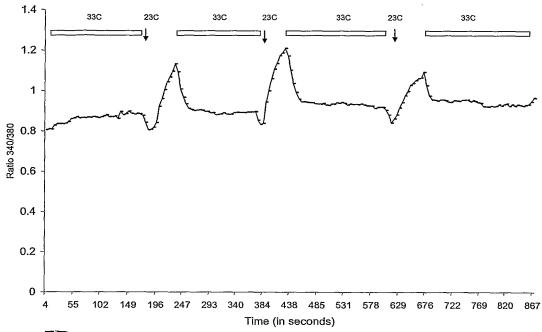


Figure 7D

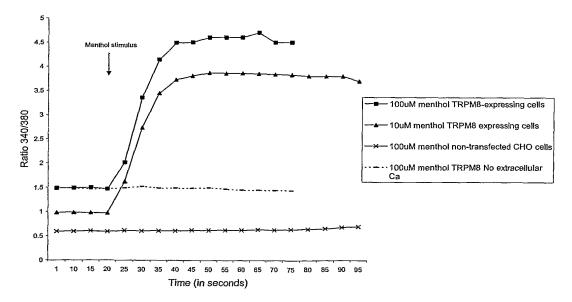


Figure 7E

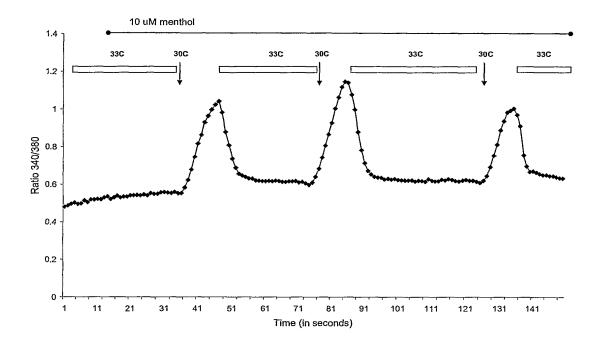
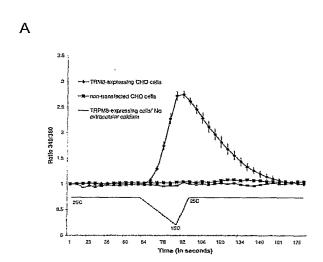


Figure 8



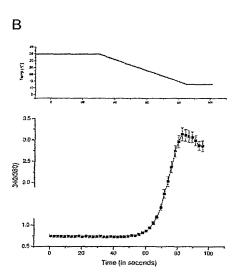
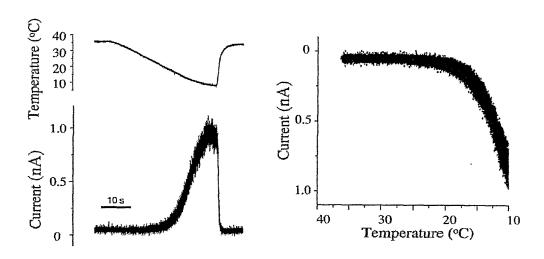


Figure 9





В

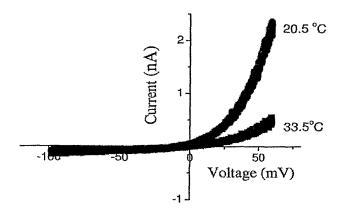
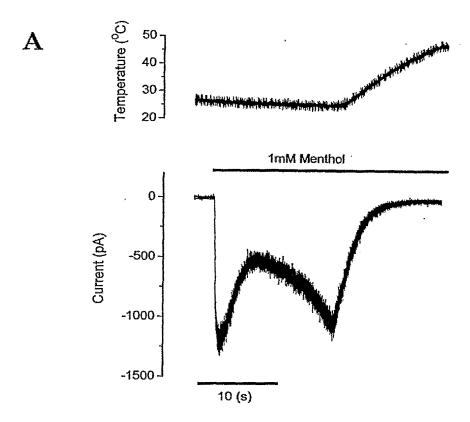


Figure 10



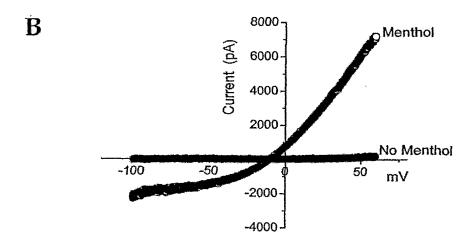
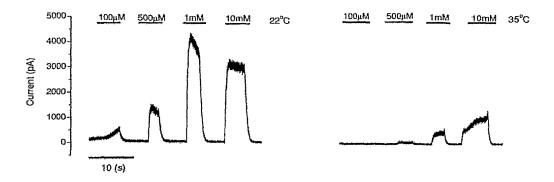
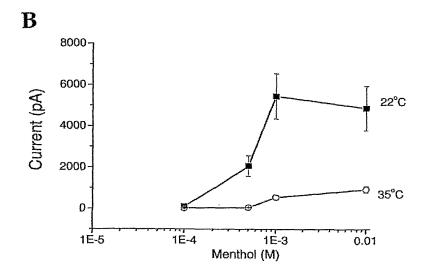


Figure 11







WO 02/101045 PCT/EP02/06520 1/75

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	cta Leu															2077
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	gag Glu												Leu			2173
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ctg Leu 720	tgc Cys	aaa Lys	gta Val	gca Ala	gat Asp 725	gag Glu	gac Asp	ttc Phe	cgg Arg	ctg Leu 730	tgt Cys	ctg Leu	cgg Arg	atc Ile	aac Asn 735	2269
	gtg Val															2317
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Gly	Phe 50		Pro	Asn	Pro	Thr 55		Thr	Lys	Thr	Ser 60		Pro	Ile	Phe	
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Pro	Ser	Asn	Pro 100		Ser	Pro	Ser	Ala 105		Leu	Ala	Lys	Glu 110		Gln	
Arg	Gln	Lys 115		Lys	Arg	Leu	Lys 120		Arg	Ile	Phe	Ala 125		Val	Ser	
Glu	Gly		Val	Glu	Glu	Leu		Glu	Leu	Leu	Gln		Leu	Gln	Asp	

								~,.	_						
T	130	70	7	70	3	135			** 7	_	140		_		1
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									Asn			aar Lys		Glu		336
												gcn Ala 125				384
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ytn Leu 145	tgy Cys	mgn Arg	mgn Arg	mgn Arg	mgn Arg 150	ggn	ytn Leu	gay Asp	gtn Val	ccn Pro 155	gay Asp	tty Phe	ytn Leu	atg Met	cay His 160	480
aar Lys	ytn Leu	acn Thr	gcn Ala	wsn Ser 165	gay Asp	acn Thr	ggn	aar Lys	acn Thr 170	tgy Cys	ytn Leu	atg Met	aar Lys	gcn Ala 175	ytn Leu	528
ytn Leu	aay Asn	ath Ile	aay Asn 180	ccn Pro	aay Asn	acn Thr	aar Lys	gar Glu 185	ath Ile	gtn Val	mgn Arg	ath Ile	ytn Leu 190	ytn Leu	gcn Ala	576
tty Phe	gcn Ala	gar Glu 195	gar Glu	aay Asn	gay Asp	ath Ile	ytn Leu 200	gay Asp	mgn Arg	tty Phe	ath Ile	aay Asn 205	gcn Ala	gar Glu	tay Tyr	624
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acn Thr 305	gtn Val	gcn Ala	gar Glu	gay Asp	tty Phe 310	aar Lys	acn Thr	car Gln	aay Asn	gay Asp 315	tty Phe	gtn Val	aar Lys	mgn Arg	atg Met 320	960
tay Tyr	gay Asp	atg Met	ath Ile	ytn Leu 325	ytn Leu	mgn Arg	wsn Ser	ggn Gly	aay Asn 330	tgg Trp	gar Glu	ytn Leu	gar Glu	acn Thr 335	atg Met	1008
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345

340

8/75

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						cay His										1440
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ath Ile	wsn Ser	gtn Val	aar Lys 500	gar Glu	ggn Gly	ath Ile	gcn Ala	ath Ile 505	tty Phe	ytn Leu	ytn Leu	mgn Arg	ccn Pro 510	wsn Ser	gay Asp	1536
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						ath Ile										1776
						ytn Leu										1824
ath Ile						gay Asp										1872

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Leu Leu Ile T			ytn ytn ytn aay Leu Leu Leu Asn 670	
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			ath ytn gar tty Ile Leu Glu Phe 700	
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			ytn mgn ath aay Leu Arg Ile Asn 735	
Val Lys Trp Tl			tty ytn aay gar Phe Leu Asn Glu 750	
ccn ggn ccn at Pro Gly Pro II 755	th mgn mgn acn le Arg Arg Thr	gcn gay ytn aay Ala Asp Leu Asn 760	aar ath car gay Lys Ile Gln Asp 765	wsn 2304 Ser
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	11

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Asn Met Leu Tyr Tyr Thr Arg Gly Phe Gln Ser Met Gly Met Tyr Ser

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Val Tyr Ile Val Phe Leu Leu Gly Phe Gly Val Ala Leu Ala Ser Leu
        595
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Ile Glu Lys Cys Pro Lys Asp Asn Lys Asp Cys Ser Ser Tyr Gly Ser
                          615
                                               620
Phe Ser Asp Ala Val Leu Glu Leu Phe Lys Leu Thr Ile Gly Leu Gly
                     630
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Asp Leu Asn Ile Gln Gln Asn Ser Lys Tyr Pro Ile Leu Phe Leu Phe
                 645
                                      650
Leu Leu Ile Thr Tyr Val Ile Leu Thr Phe Val Leu Leu Leu Asn Met
                                  665
Leu Ile Ala Leu Met Gly Glu Thr Val Glu Asn Val Ser Lys Glu Ser
                              680
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Glu Arg Ile Trp Arg Leu Gln Arg Ala Arg Thr Ile Leu Glu Phe Glu
                         695
                                               700
Lys Met Leu Pro Glu Trp Leu Arg Ser Arg Phe Arg Met Gly Glu Leu
705
                     710
                                          715
Cys Lys Val Ala Glu Asp Asp Phe Arg Leu Cys Leu Arg Ile Asn Glu
                 725
                                      730
Val Lys Trp Thr Glu Trp Lys Thr His Val Ser Phe Leu Asn Glu Asp
                                  745
Pro Gly Pro Val Arg Arg Thr Ala Asp Phe Asn Lys Ile Gln Asp Ser
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Ser Arg Asn Asn Ser Lys Thr Thr Leu Asn Ala Phe Glu Glu Val Glu
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Glu Phe Pro Glu Thr Ser Val
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<211> 2373
<212> DNA
<213> Artificial Sequence
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<223> Generic sequence that encompasses all nucleotide
      sequences that encode human TRPV3 having an amino
      acid sequence as shown in SEQ ID NO:5
<221> misc feature
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984,1086,1116,1122,1158,1161,1164,1206,1332,1377,1494,1533,1545,1554,1608,
1713, 1728, 1821, 1860, 1863, 1872, 1878, 1944, 2055,
2064,2139,2241,2304,2307,2319,2370
<223> n = A,T,C or G if after TC;
      n = T or C if after AG
<221> misc_feature
<222> 45,90,219,339,342,351,354,366,441,444,447,564,606,675,678,
876,885,957,981,1011,1089,1107,1113,1125,1248,1386,1392,
1461,1527,1701,2070,2079,2088,2136,2142,2148,2187,2199,2271,2274,
<223> n = A,T,C or G if after CG;
      n = A or G if after AG
<221> misc_feature <222> all "n" not specified above
<223> n = A,T,C or G
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atg aar gen eay een aar gar atg gtn een ytn atg ggn aar mgn gtn
                                                                          48
Met Lys Ala His Pro Lys Glu Met Val Pro Leu Met Gly Lys Arg Val
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                                       10
```

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									Leu						gcn Ala	96	
															gar Glu	144	
						acn Thr 55									tty Phe	192	
						aay Asn									tgy Cys 80	240	
gay Asp	gay Asp	atg Met	gay Asp	wsn Ser 85	ccn Pro	car Gln	wsn Ser	ccn Pro	car Gln 90	gay Asp	gay Asp	gtn Val	acn Thr	gar Glu 95	acn Thr	288	
						ccn Pro										336	
						ytn Leu										384	
						ytn Leu 135										432	
ytn Leu 145	tgy Cys	mgn Arg	mgn Arg	mgn Arg	cay His 150	gay Asp	gar Glu	gay Asp	gtn Val	ccn Pro 155	gay Asp	tty Phe	ytn Leu	atg Met	cay His 160	480	
						acn Thr										528	
						acn Thr										576	
tty Phe	gcn Ala	gar Glu 195	gar Glu	aay Asn	gay Asp	ath Ile	ytn Leu 200	ggn Gly	mgn Arg	tty Phe	ath Ile	aay Asn 205	gcn Ala	gar Glu	tay Tyr	624	
acn Thr	gar Glu 210	gar Glu	gcn Ala	tay Tyr	gar Glu	ggn Gly 215	car Gln	acn Thr	gcn Ala	ytn Leu	aay Asn 220	ath Ile	gcn Ala	ath Ile	gar Glu	672	
mgn Arg 225	mgn Arg	car Gln	gly	gay Asp	ath Ile 230	gcn Ala	gcn Ala	ytn Leu	ytn Leu	ath Ile 235	gcn Ala	gcn Ala	ggn Gly	gcn Ala	gay Asp 240	720	
gtn Val	aay Asn	gcn Ala	cay His	gcn Ala 245	aar Lys	ggn Gly	gcn Ala	tty Phe	tty Phe 250	aay Asn	ccn Pro	aar Lys	tay Tyr	car Gln 255	cay His	768	
						gar Glu										816	
aay Asn	car Gln	ccn Pro 275	gar Glu	ath Ile	gtn Val	car Gln	ytn Leu 280	ytn Leu	atg Met	gar Glu	cay His	gar Glu 285	car Gln	acn Thr	gay Asp	864	

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		16/75		
ath acn wsn m Ile Thr Ser A 290	mgn gay wsn mgr Arg Asp Ser Arg 295	Gly Asn Asn Ile	n ytn cay gcn ytn e Leu His Ala Leu 300	gtn 912 Val
			y tty gtn aar mgn o Phe Val Lys Arg o	
tay gay atg a Tyr Asp Met I	ath ytn ytn mgr Ile Leu Leu Arc 325	wsn ggn aay tgg Ser Gly Asn Trp 330	g gar ytn gar acn o Glu Leu Glu Thr 335	acn 1008 Thr
Arg Asn Asn A	gay ggn ytn acr Asp Gly Leu Thi 340	ccn ytn car ytn Pro Leu Gln Leu 345	n gcn gcn aar atg n Ala Ala Lys Met 350	ggn 1056 Gly
aar gcn gar a Lys Ala Glu I 355	ath ytn aar tay Ile Leu Lys Tyr	ath ytn wsn mgn Ile Leu Ser Arg 360	n gar ath aar gar g Glu Ile Lys Glu 365	aar 1104 Lys
		Lys Phe Thr Asp	tgg gcn tay ggn Trp Ala Tyr Gly 380	
gtn wsn wsn w Val Ser Ser S 385	vsn ytn tay gay Ser Leu Tyr Asp 390	ytn acn aay gtn Leu Thr Asn Val 395	gay acn acn acn Asp Thr Thr Thr	gay 1200 Asp 400
aay wsn gtn y Asn Ser Val L	otn gar ath acr beu Glu Ile Thr 405	gtn tay aay acn Val Tyr Asn Thr 410	aay ath gay aay Asn Ile Asp Asn 415	mgn 1248 Arg
His Glu Met L			ytn ytn cay atg Leu Leu His Met 430	
tgg aar aar t Trp Lys Lys P 435	ty gcn aar cay Phe Ala Lys His	atg tty tty ytn Met Phe Phe Leu 440	wsn tty tgy tty Ser Phe Cys Phe 445	tay 1344 Tyr
			tay tay mgn ccn Tyr Tyr Arg Pro 460	
gar gar gar g Glu Glu Glu A 465	cn ath ccn cay la Ile Pro His 470	ccn ytn gcn ytn Pro Leu Ala Leu 475	acn cay aar atg Thr His Lys Met	ggn 1440 Gly 480
tgg ytn car y Trp Leu Gln L	tn ytn ggn mgn eu Leu Gly Arg 485	atg tty gtn ytn Met Phe Val Leu 490	ath tgg gcn atg Ile Trp Ala Met 495	tgy 1488 Cys
Ile Ser Val L	ar gar ggn ath ys Glu Gly Ile 00	gcn ath tty ytn Ala Ile Phe Leu 505	ytn mgn ccn wsn Leu Arg Pro Ser 510	gay 1536 Asp
			tty gtn tty tty Phe Val Phe Phe 525	
car gcn gtn y Gln Ala Val Le 530	tn gtn ath ytn eu Val Ile Leu 535	wsn gtn tty ytn Ser Val Phe Leu	tay ytn tty gcn Tyr Leu Phe Ala 540	tay 1632 Tyr
aar gar tay y Lys Glu Tyr Le 545	tn gcn tgy ytn eu Ala Cys Leu 550	gtn ytn gcn atg Val Leu Ala Met 555	gcn ytn ggn tgg Ala Leu Gly Trp	gcn 1680 Ala 560

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aay Asn	atg Met	ytn Leu	tay Tyr	tay Tyr 565	Thr	mgn Arg	ggn	tty Phe	car Gln 570	Ser	ato Met	ggn Gly	atg Met	tay Tyr 575	wsn Ser	1728
gtn Val	atg Met	ath Ile	car Gln 580	aar Lys	gtn Val	ath Ile	ytn Leu	cay His 585	Asp	gtn Val	ytn Leu	ı aar ı Lys	tty Phe 590	Leu	tty Phe	1776
gtn Val	tay Tyr	ath Ile 595	Val	tty Phe	ytn Leu	ytn Leu	ggn Gly 600	tty Phe	ggn	gtn Val	gcn Ala	ytn Leu 605	Ala	wsn Ser	ytn Leu	1824
ath Ile	gar Glu 610	aar Lys	tgy Cys	ccn Pro	aar Lys	gay Asp 615	aay Asn	aar Lys	gay Asp	tgy Cys	wsn Ser 620	wsn Ser	tay Tyr	ggn Gly	wsn Ser	1872
tty Phe 625	wsn Ser	gay Asp	gcn Ala	gtn Val	ytn Leu 630	gar Glu	ytn Leu	tty Phe	aar Lys	ytn Leu 635	acn Thr	ath Ile	ggn	ytn Leu	ggn Gly 640	1920
gay Asp	ytn Leu	aay Asn	ath Ile	car Gln 645	car Gln	aay Asn	wsn Ser	aar Lys	tay Tyr 650	ccn Pro	ath Ile	ytn Leu	tty Phe	ytn Leu 655	tty Phe	1968
ytn Leu	ytn Leu	ath Ile	acn Thr 660	tay Tyr	gtn Val	ath Ile	ytn Leu	acn Thr 665	tty Phe	gtn Val	ytn Leu	ytn Leu	ytn Leu 670	aay Asn	atg Met	2016
ytn Leu	ath Ile	gcn Ala 675	ytn Leu	atg Met	ggn Gly	gar Glu	acn Thr 680	gtn Val	gar Glu	aay Asn	gtn Val	wsn Ser 685	aar Lys	gar Glu	wsn Ser	2064
gar Glu	mgn Arg 690	ath Ile	tgg Trp	mgn Arg	ytn Leu	car Gln 695	mgn Arg	gcn Ala	mgn Arg	acn Thr	ath Ile 700	ytn Leu	gar Glu	tty Phe	gar Glu	2112
aar Lys 705	atg Met	ytn Leu	ccn Pro	gar Glu	tgg Trp 710	ytn Leu	mgn Arg	wsn Ser	mgn Arg	tty Phe 715	mgn Arg	atg Met	ggn Gly	gar Glu	ytn Leu 720	2160
tgy Cys	aar Lys	gtn Val	gcn Ala	gar Glu 725	gay Asp	gay Asp	tty Phe	mgn Arg	ytn Leu 730	tgy Cys	ytn Leu	mgn Arg	ath Ile	aay Asn 735	gar Glu	2208
gtn Val	aar Lys	tgg Trp	acn Thr 740	gar Glu	tgg Trp	aar Lys	acn Thr	cay His 745	gtn Val	wsn Ser	tty Phe	ytn Leu	aay Asn 750	gar Glu	gay Asp	2256
ccn Pro	ggn Gly	ccn Pro 755	gtn Val	mgn Arg	mgn Arg	Thr	gcn Ala 760	gay Asp	tty Phe	aay Asn	aar Lys	ath Ile 765	car Gln	gay Asp	wsn Ser	2304
wsn Ser	mgn Arg 770	aay Asn	aay Asn	wsn Ser	Lys	acn Thr 775	acn Thr	ytn Leu	aay Asn	Ala	tty Phe 780	gar Glu	gar Glu	gtn Val	gar Glu	2352
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<220> <221> CDS

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atg Met 10	agg Arg	agc Ser	cgc Arg	aga Arg	aat Asn 15	ggt Gly	act Thr	atg Met	ggc	agc Ser 20	acc Thr	cgg Arg	acc Thr	ctg Leu	tac Tyr 25	522
tcc Ser	agt Ser	gta Val	tct Ser	cgg Arg 30	agc Ser	aca Thr	gac Asp	gtg Val	tcc Ser 35	tac Tyr	agt Ser	gac Asp	agt Ser	gat Asp 40	ttg Leu	570
														ttc Phe		618
														tat Tyr		666
														aag Lys		714
aac Asn 90	tac Tyr	aaa Lys	aaa Lys	cat His	acc Thr 95	aag Lys	gag Glu	ttt Phe	cca Pro	aca Thr 100	gac Asp	gcc Ala	ttc Phe	gly aaa	gac Asp 105	762
att Ile	cag Gln	ttt Phe	gag Glu	act Thr 110	ctg Leu	gly aaa	aag Lys	aaa Lys	ggc Gly 115	aag Lys	tac Tyr	tta Leu	cgc Arg	ttg Leu 120	tcc Ser	810
tgt Cys	gac Asp	acc Thr	gac Asp 125	tct Ser	gaa Glu	act Thr	ctc Leu	tac Tyr 130	gaa Glu	ctg Leu	ctg Leu	acc Thr	cag Gln 135	cac His	tgg Trp	858
cac Hís	ctc Leu	aaa Lys 140	aca Thr	ccc Pro	aac Asn	ctg Leu	gtc Val 145	att Ile	tca Ser	gtg Val	acg Thr	ggt Gly 150	gga Gly	gcc Ala	aaa Lys	906
aac Asn	ttt Phe 155	gct Ala	ttg Leu	aag Lys	cca Pro	cgc Arg 160	atg Met	cgc Arg	aag Lys	atc Ile	ttc Phe 165	agc Ser	agg Arg	ctg Leu	att Ile	954
tac Tyr 170	atc Ile	gca Ala	cag Gln	tct Ser	aaa Lys 175	ggt Gly	gcg Ala	tgg Trp	att Ile	ctc Leu 180	act Thr	gga Gly	ggc Gly	act Thr	cac His 185	1002
tac Tyr	ggc Gly	ctg Leu	atg Met	aag Lys 190	tac Tyr	ata Ile	ggc Gly	gag Glu	gtg Val 195	gtg Val	aga Arg	gac Asp	aac Asn	acc Thr 200	atc Ile	1050
agc Ser	agg Arg	aac Asn	tca Ser 205	gaa Glu	gag Glu	aac Asn	atc Ile	gtg Val 210	gcc Ala	att Ile	ggc Gly	atc Ile	gca Ala 215	gca Ala	tgg Trp	1098
ggc Gly	atg Met	gtc Val	tcc Ser	aac Asn	agg Arg	gac Asp	acc Thr	ctc Leu	atc Ile	agg Arg	agc Ser	tgt Cys	gat Asp	gat Asp	gag Glu	1146

		220					225					230)			
gga Gly	cat His 235	ttt Phe	tca Ser	gct Ala	caa Gln	tac Tyr 240	Ile	atg Met	gat Asp	gac Asp	ttt Phe 245	Thr	aga Arg	gad J Asp	cct Pro	1194
cta Leu 250	tac Tyr	atc Ile	ctg Leu	gac Asp	aac Asn 255	aac Asn	cat His	acc Thr	cac His	ctg Leu 260	Leu	ctt Leu	gtg Val	gac Asp	aac Asn 265	1242
ggt Gly	tgt Cys	cat His	gga Gly	cac His 270	ccc Pro	aca Thr	gtg Val	gaa Glu	gcc Ala 275	Lys	ctc Leu	cgg Arg	aat Asn	caç Glr 280	ctg Leu	1290
gaa Glu	aag Lys	tac Tyr	atc Ile 285	tct Ser	gag Glu	cgc Arg	acc Thr	agt Ser 290	caa Gln	gat Asp	tcc Ser	aac Asn	tat Tyr 295	Gly	ggt Gly	1338
aag Lys	atc Ile	ccc Pro 300	atc Ile	gtg Val	tgt Cys	ttt Phe	gcc Ala 305	caa Gln	gga Gly	ggt	gga Gly	aga Arg 310	gag Glu	act Thr	cta Leu	1386
aaa Lys	gcc Ala 315	atc Ile	aac Asn	acc Thr	tct Ser	gtc Val 320	aaa Lys	agc Ser	aag Lys	atc Ile	cct Pro 325	tgt Cys	gtg Val	gtg Val	gtg Val	1434
gaa Glu 330	ggc Gly	tcg Ser	gjà aaa	cag Gln	att Ile 335	gct Ala	gat Asp	gtg Val	atc Ile	gcc Ala 340	agc Ser	ctg Leu	gtg Val	gag Glu	gtg Val 345	1482
gag Glu	gat Asp	gtt Val	tta Leu	acc Thr 350	tct Ser	tcc Ser	atg Met	gtc Val	aaa Lys 355	gag Glu	aag Lys	ctg Leu	gta Val	cgc Arg 360	ttt Phe	1530
tta Leu	cca Pro	cgc Arg	act Thr 365	gtg Val	tcc Ser	cgg Arg	ctg Leu	cct Pro 370	gaa Glu	gag Glu	gaa Glu	att Ile	gag Glu 375	agc Ser	tgg Trp	1578
atc Ile	aaa Lys	tgg Trp 380	ctc Leu	aaa Lys	gaa Glu	att Ile	ctt Leu 385	gag Glu	agt Ser	tct Ser	cac His	cta Leu 390	ctc Leu	aca Thr	gta Val	1626
att Ile	aag Lys 395	atg Met	gaa Glu	gag Glu	gct Ala	gga Gly 400	gat Asp	gag Glu	att Ile	gtg Val	agc Ser 405	aac Asn	gcc Ala	att Ile	tcc Ser	1674
tat Tyr 410	gcg Ala	ctg Leu	tac Tyr	aaa Lys	gcc Ala 415	ttc Phe	agc Ser	act Thr	aat Asn	gag Glu 420	caa Gln	gac Asp	aag Lys	gac Asp	aac Asn 425	1722
tgg Trp	aat Asn	gga Gly	cag Gln	ctg Leu 430	aag Lys	ctt Leu	ctg Leu	ctg Leu	gag Glu 435	tgg Trp	aac Asn	cag Gln	ttg Leu	gac Asp 440	ctt Leu	1770
gcc Ala	agt Ser	gat Asp	gag Glu 445	atc Ile	ttc Phe	acc Thr	aat Asn	gac Asp 450	cgc Arg	cgc Arg	tgg Trp	gag Glu	tct Ser 455	gcc Ala	gac Asp	1818
ctt Leu	cag Gln	gag Glu 460	gtc Val	atg Met	ttc Phe	acg Thr	gct Ala 465	ctc Leu	ata Ile	aag Lys	gac Asp	aga Arg 470	ccc Pro	aag Lys	ttt Phe	1866
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acc Thr	ttt Phe	gtc Val	tgg Trp 525	aag Lys	ttg Leu	gtg Val	gca Ala	aac Asn 530	ttc Phe	cgt Arg	cga Arg	agc Ser	ttc Phe 535	Trp	aaa Lys	2058
gag Glu	gac Asp	aga Arg 540	agc Ser	agc Ser	agg Arg	gag Glu	gac Asp 545	ttg Leu	gat Asp	gtg Val	gaa Glu	ctc Leu 550	cat His	gat Asp	gca Ala	2106
tct Ser	ctc Leu 555	acc Thr	acc Thr	cgg Arg	cac His	ccg Pro 560	ctg Leu	caa Gln	gct Ala	ctc Leu	ttc Phe 565	atc Ile	tgg Trp	gcc Ala	att Ile	2154
ctt Leu 570	cag Gln	aac Asn	aag Lys	aag Lys	gaa Glu 575	ctc Leu	tcc Ser	aag Lys	gt <i>c</i> Val	att Ile 580	tgg Trp	gag Glu	cag Gln	acc Thr	aaa Lys 585	2202
ggc Gly	tgt Cys	act Thr	ctg Leu	gca Ala 590	gcc Ala	ttg Leu	gly aaa	gcc Ala	agc Ser 595	aag Lys	ctt Leu	ctg Leu	aag Lys	acc Thr 600	ctg Leu	2250
gcc Ala	aaa Lys	gtt Val	aag Lys 605	aat Asn	gat Asp	atc Ile	aac Asn	gct Ala 610	gct Ala	gjà aaa	gaa Glu	tcg Ser	gag Glu 615	gaa Glu	ctg Leu	2298
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cag Gln	cat His	ttc Phe	atc Ile	gct Ala 670	cag Gln	cct Pro	gly ggg	gtc Val	cag Gln 675	aat Asn	ttc Phe	ctt Leu	tct Ser	aag Lys 680	caa Gln	2490
tgg Trp	tat Tyr	gga Gly	gag Glu 685	att Ile	tcc Ser	cga Arg	gac Asp	acg Thr 690	aag Lys	aac Asn	tgg Trp	aag Lys	att Ile 695	atc Ile	ctg Leu	2538
tgt Cys	cta Leu	ttc Phe 700	att Ile	atc Ile	ccc Pro	tta Leu	gtg Val 705	ggc Gly	tgt Cys	ggc Gly	ctc Leu	gta Val 710	tca Ser	ttt Phe	agg Arg	2586
aag Lys	aaa Lys 715	ccc Pro	att Ile	gac Asp	aag Lys	cac His 720	aag Lys	aag Lys	ctg Leu	ctg Leu	tgg Trp 725	tac Tyr	tat Tyr	gtg Val	gcc Ala	2634
ttc Phe 730	ttc Phe	acg Thr	tcg Ser	ccc Pro	ttc Phe 735	gtg Val	gtc Val	ttc Phe	tcc Ser	tgg Trp 740	aac Asn	gtg Val	gtc Val	ttc Phe	tac Tyr 745	2682
atc Ile	gcc Ala	tt <i>c</i> Phe	ctc Leu	ctg Leu 750	ctg Leu	ttt Phe	gcc Ala	tat Tyr	gtg Val 755	ctg Leu	ctc Leu	atg Met	gac Asp	ttc Phe 760	cac His	2730
tca Ser	gtg Val	cca Pro	cac His	acc Thr	ccc Pro	gag Glu	ctg Leu	atc Ile	ctc Leu	tac Tyr	gcc Ala	ctg Leu	gtc Val	ttc Phe	gtc Val	2778

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														aat Asn		2826
														tac Tyr		2874
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tac Tyr	tct Ser	gjå aaa	cgc Arg	gtc Val 830	att Ile	ttc Phe	tgt Cys	ctg Leu	gat Asp 835	tac Tyr	att Ile	ata Ile	ttc Phe	acg Thr 840	cta Leu	2970
agg Arg	ctc Leu	atc Ile	cac His 845	att Ile	ttc Phe	acc Thr	gtc Val	agc Ser 850	agg Arg	aac Asn	ttg Leu	gga Gly	ccc Pro 855	aag Lys	att Ile	3018
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														tcc Ser		3258
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ctt Leu 970	ctg Leu	gtc Val	aac Asn	ctc Leu	ctg Leu 975	gtc Val	gcc Ala	atg Met	ttt Phe	ggc 980	tac Tyr	acg Thr	gta Val	gly ggc	att Ile 985	3402
gta Val	cag Gln	gag Glu	aac Asn	aac Asn 990	Asp	cag Gln	gt <i>c</i> Val	tgg Trp	aaa Lys 995	Phe	cag Gln	cgg Arg	tac Tyr	ttc Phe 1000	Leu	3450
gtg Val	cag Gln	gag Glu	tac Tyr 1005	Cys	aac Asn	cgc Arg	cta Leu	aac Asn 1010	Ile	ccc Pro	ttc Phe	ccc Pro	ttc Phe 1015	gtt Val	gtc Val	3498
ttc Phe	gct Ala	tat Tyr 1020	Phe	tac Tyr	atg Met	gtg Val	gtg Val 1025	Lys	aag Lys	tgt Cys	ttc Phe	aaa Lys 1030	Cys	tgc Cys	tgt Cys	3546
aaa Lys	gag Glu	aag Lys	aat Asn	atg Met	gag Glu	tct Ser	aat Asn	gcc Ala	tgc Cys	tgt Cys	ttc Phe	aga Arg	aat Asn	gag Glu	gac Asp	3594

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aag atc aac acc Lys Ile Asn Thr	aaa gcc aac ga Lys Ala Asn As 1070	c aac tca gag p Asn Ser Glu 1075	gag atg agg cat cg Glu Met Arg His Ar 1080	gg 3690 Fg
ttt aga caa ctg Phe Arg Gln Leu 108	Asp Ser Lys Le	t aac gac ctc u Asn Asp Leu 1090	aaa agt ctt ctg aa Lys Ser Leu Leu Ly 1095	aa 3738 ⁄s
gag att gct aat Glu Ile Ala Asn 1100		a ggctggcgat g	gcttgtgggg agaaacca	aa 3792
actgctaatg actt tagtcaccct caag aggggtgcaa agga	ctaaag gagacatt ggcata ggtcaggg ccatgt tettetgt agggat taggtgtt atactg c	tt caggtccctg ag caaagtgtac ga aggtgcctgt	atgggacact gatggac agcacagggt ggatgac agaggacttt acacctg gttttctgca tctcaga ccacgactgt gactctg	tct 3912 gaag 3972 gcc 4032
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Phe Lys Lys Arg	Glu Cys Val Phe	e Phe Thr Arg	Asp Ser Lys Ala Me	t
Glu Asn Ile Cys 65	Lys Cys Gly Tys	Ala Gln Ser	Gln His Ile Glu Gl	-
	, -		Lys Lys His Thr Ly	
		Asp Ile Gln	95 Phe Glu Thr Leu Gl	У
			110 Thr Asp Ser Glu Th	r
Leu Tyr Glu Leu			Lys Thr Pro Asn Le	u
130 Val Ile Ser Val	135 Thr Gly Gly Ala	Lys Asn Phe	140 Ala Leu Lys Pro Arg	g
145 Met Arg Lys Ile	150 Phe Ser Arg Lei	155 Ile Tyr Ile	16 Ala Gln Ser Lys Gl	
	165	170	175 Leu Met Lys Tyr Ile	-
180		185	190 Asn Ser Glu Glu Ası	
195	200)	205 Val Ser Asn Arg As	
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225	230	235	Phe Ser Ala Gln Ty:	0
	245	250	Ile Leu Asp Asn Ası 255	
260		265	His Gly His Pro Th	
Val Glu Ala Lys 275	Leu Arg Asn Glr 280	Leu Glu Lys	Tyr Ile Ser Glu Arg	Э
=			Pro Ile Val Cys Phe	9

								_							
	290					295					300				
Ala 305	Gln	Gly	Gly	Gly	Arg 310		Thr	Leu	Lys	Ala 315		Asn	Thr	Ser	Val 320
	Ser	Lys	Ile	Pro 325		Val	Val	Val	Glu 330	Gly	Ser	Gly	Gln	Ile 335	
Asp	Val	Ile	Ala 340		Leu	Val	Glu	Val 345		Asp	Val	Leu	Thr 350		Ser
Met	Val	Lys 355		Lys	Leu	Val	Arg 360		Leu	Pro	Arg	Thr 365		Ser	Arg
Leu	Pro 370		Glu	Glu	Ile	Glu 375		Trp	Ile	Lys	Trp 380		Lys	Glu	Ile
Leu 385		Ser	Ser	His	Leu 390		Thr	Val	Ile	Lys 395		Glu	Glu	Ala	Gly 400
	Glu	Ile	Val	Ser 405		Ala	Ile	Ser	Tyr 410	Ala	Leu	Tyr	Lys	Ala 415	
Ser	Thr	Asn	Glu 420		Asp	Lys	Asp	Asn 425		Asn	Gly	Gln	Leu 430		Leu
Leu	Leu	Glu 435	Trp	Asn	Gln	Leu	Asp 440		Ala	Ser	Asp	Glu 445	Ile	Phe	Thr
Asn	Asp 450	Arg	Arg	Trp	Glu	Ser 455	Ala	Asp	Leu	Gln	Glu 460	Val	Met	Phe	Thr
Ala 465	Leu	Ile	Lys	Asp	Arg 470	Pro	Lys	Phe	Val	Arg 475	Leu	Phe	Leu	Glu	Asn 480
				485					490	Glu				495	
Phe	Ser	Thr	His 500	Phe	Ser	Thr	Leu	Val 505	Tyr	Arg	Asn	Leu	Gln 510	Ile	Ala
Lys	Asn	Ser 515	Tyr	Asn	Asp	Ala	Leu 520	Leu	Thr	Phe	Val	Trp 525	ГЛЗ	Leu	Val
	530					535				Asp	540				
545		_			550		_			Leu 555			_		560
				565					570	Gln				575	
			580					585		Cys			590		
		595					600		•	Lys		605			
	610					615				Asn	620	_			_
625					630					Asn 635					640
				645	_		_		650	Trp	_	_		655	_
			660					665		His			670		
-		675					680			Tyr -	_	685			_
_	690	_			_	695			-	Leu	700				
705					710					Lys 715					720
				725					730	Phe				735	
			740					745		Ala -			750		
		755					760			Val		765			
	770					775				Phe	780				•
785				_	790					Thr 795		_			800
	_			805			-		810	Ala	_			815	•
			820					825	_	Ser			830		
Cys	Leu	Asp	Tyr	Ile	Ile	Phe	Thr	Leu	Arg	Leu	Ile	His	Ile	Phe	Thr

840 845 Val Ser Arg Asn Leu Gly Pro Lys Ile Ile Met Leu Gln Arg Met Leu 855 860 Ile Asp Val Phe Phe Leu Phe Leu Phe Ala Val Trp Met Val Ala 870 875 Phe Gly Val Ala Arg Gln Gly Ile Leu Arg Gln Asn Glu Gln Arg Trp 885 890 895 Arg Trp Ile Phe Arg Ser Val Ile Tyr Glu Pro Tyr Leu Ala Met Phe 905 900 Gly Gln Val Pro Ser Asp Val Asp Ser Thr Thr Tyr Asp Phe Ser His 920 925 915 Cys Thr Phe Ser Gly Asn Glu Ser Lys Pro Leu Cys Val Glu Leu Asp 940 930 935 Glu His Asn Leu Pro Arg Phe Pro Glu Trp Ile Thr Ile Pro Leu Val 955 950 Cys Ile Tyr Met Leu Ser Thr Asn Ile Leu Leu Val Asn Leu Leu Val 970 Ala Met Phe Gly Tyr Thr Val Gly Ile Val Gln Glu Asn Asn Asp Gln 980 985 990 Val Trp Lys Phe Gln Arg Tyr Phe Leu Val Gln Glu Tyr Cys Asn Arg 1005 995 1000 Leu Asn Ile Pro Phe Pro Phe Val Val Phe Ala Tyr Phe Tyr Met Val 1015 1020 1010 Val Lys Lys Cys Phe Lys Cys Cys Lys Glu Lys Asn Met Glu Ser 1035 1030 Asn Ala Cys Cys Phe Arg Asn Glu Asp Asn Glu Thr Leu Ala Trp Glu 1055 1050 1045 Gly Val Met Lys Glu Asn Tyr Leu Val Lys Ile Asn Thr Lys Ala Asn 1065 1070 1060 Asp Asn Ser Glu Glu Met Arg His Arg Phe Arg Gln Leu Asp Ser Lys 1075 1080 1085 Leu Asn Asp Leu Lys Ser Leu Leu Lys Glu Ile Ala Asn Asn Ile Lys 1095 1100

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											tty Phe					144
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											aar Lys					288
											tty Phe					336
											acn Thr					384
											aar Lys 140					432
											gcn Ala					480
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gcn Ala	tgg Trp	ath Ile	ytn Leu 180	acn Thr	ggn Gly	ggn Gly	acn Thr	cay His 185	tay Tyr	ggn Gly	ytn Leu	atg Met	aar Lys 190	tay Tyr	ath Ile	576
											aay Asn					624
ath Ile	gtn Val 210	gcn Ala	ath Ile	ggn Gly	ath Ile	gcn Ala 215	gcn Ala	tgg Trp	ggn Gly	atg Met	gtn Val 220	wsn Ser	aay Asn	mgn Arg	gay Asp	672
acn Thr 225	ytn Leu	ath Ile	mgn Arg	wsn Ser	tgy Cys 230	gay Asp	gay Asp	gar Glu	ggn Gly	cay His 235	tty Phe	wsn Ser	gcn Ala	car Gln	tay Tyr 240	720
ath Ile	atg Met	gay Asp	gay Asp	tty Phe 245	acn Thr	mgn Arg	gay Asp	ccn Pro	ytn Leu 250	tay Tyr	ath Ile	ytn Leu	gay Asp	aay Asn 255	aay Asn	768
cay His	acn Thr	cay His	ytn Leu	ytn Leu	ytn Leu	gtn Val	gay Asp	aay Asn	ggn Gly	tgy Cys	cay His	ggn Gly	cay His	ccn Pro	acn Thr	816

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					mgn Arg											864
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					mgn Arg 310											960
					tgy Cys											1008
					ytn Leu											1056
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					ath Ile											1152
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					aay Asn											1248
wsn Ser	acn Thr	aay Asn	gar Glu 420	car Gln	gay Asp	aar Lys	gay Asp	aay Asn 425	tgg Trp	aay Asn	ggn Gly	car Gln	ytn Leu 430	aar Lys	ytn Leu	1296
ytn Leu	ytn Leu	gar Glu 435	tgg Trp	aay Asn	car Gln	ytn Leu	gay Asp 440	ytn Leu	gcn Ala	wsn Ser	gay Asp	gar Glu 445	ath Ile	tty Phe	acn Thr	1344
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gcn Ala 465	ytn Leu	ath Ile	aar Lys	gay Asp	mgn Arg 470	ccn Pro	aar Lys	tty Phe	gtn Val	mgn Arg 475	ytn Leu	tty Phe	ytn Leu	gar Glu	aay Asn 480	1440
ggn Gly	ytn Leu	aay Asn	ytn Leu	car Gln 485	aar Lys	tty Phe	ytn Leu	acn Thr	aay Asn 490	gar Glu	gtn Val	ytn Leu	acn Thr	gar Glu 495	ytn Leu	1488
					wsn Ser											1536
					gay Asp											1584
gcn Ala	aay Asn	tty Phe	mgn Arg	mgn Arg	wsn Ser	tty Phe	tgg Trp	aar Lys	gar Glu	gay Asp	mgn Arg	wsn Ser	wsn Ser	mgn Arg	gar Glu	1632

535

	550					232					340					
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ytn Leu	car Gln	gcn Ala	ytn Leu	tty Phe 565	ath Ile	tgg Trp	gcn Ala	ath Ile	ytn Leu 570	car Gln	aay Asn	aar Lys	aar Lys	gar Glu 575	ytn Leu	1728
wsn Ser	aar Lys	gtn Val	ath Ile 580	tgg Trp	gar Glu	car Gln	acn Thr	aar Lys 585	ggn	tgy Cys	acn Thr	ytn Leu	gcn Ala 590	gcn Ala	ytn Leu	1776
Gly ggn	gcn Ala	wsn Ser 595	aar Lys	ytn Leu	ytn Leu	aar Lys	acn Thr 600	ytn Leu	gcn Ala	aar Lys	gtn Val	aar Lys 605	aay Asn	gay Asp	ath Ile	1824
aay Asn	gcn Ala 610	gcn Ala	ggn Gly	gar Glu	wsn Ser	gar Glu 615	gar Glu	ytn Leu	gcn Ala	aay Asn	gar Glu 620	tay Tyr	gar Glu	acn Thr	mgn Arg	1872
gcn Ala 625	gtn Val	gar Glu	ytn Leu	tty Phe	acn Thr 630	gar Glu	tgy Cys	tay Tyr	wsn Ser	aay Asn 635	gay Asp	gar Glu	gay Asp	ytn Leu	gcn Ala 640	1920
gar Glu	car Gln	ytn Leu	ytn Leu	gtn Val 645	tay Tyr	wsn Ser	tgy Cys	gar Glu	gcn Ala 650	tgg Trp	ggn Gly	ggn Gly	wsn Ser	aay Asn 655	tgy Cys	1968
ytn Leu	gar Glu	ytn Leu	gcn Ala 660	gtn Val	gar Glu	gcn Ala	acn Thr	gay Asp 665	car Gln	cay His	tty Phe	ath Ile	gcn Ala 670	car Gln	ccn Pro	2016
ggn Gly	gtn Val	car Gln 675	aay Asn	tty Phe	ytn Leu	wsn Ser	aar Lys 680	car Gln	tgg Trp	tay Tyr	ggn Gly	gar Glu 685	ath Ile	wsn Ser	mgn Arg	2064
gay Asp	acn Thr 690	aar Lys	aay Asn	tgg Trp	aar Lys	ath Ile 695	ath Ile	ytn Leu	tgy Cys	ytn Leu	tty Phe 700	ath Ile	ath Ile	ccn Pro	ytn Leu	2112
gtn Val 705	ggn Gly	tgy Cys	ggn Gly	ytn Leu	gtn Val 710	wsn Ser	tty Phe	mgn Arg	aar Lys	aar Lys 715	ccn Pro	ath Ile	gay Asp	aar Lys	cay His 720	2160
aar Lys	aar Lys	ytn Leu	ytn Leu	tgg Trp 725	tay Tyr	tay Tyr	gtn Val	gen Ala	tty Phe 730	tty Phe	acn Thr	wsn Ser	ccn Pro	tty Phe 735	gtn Val	2208
gtn Val	tty Phe	wsn Ser	tgg Trp 740	aay Asn	gtn Val	gtn Val	tty Phe	tay Tyr 745	ath Ile	gcn Ala	tty Phe	ytn Leu	ytn Leu 750	ytn Leu	tty Phe	2256
gcn Ala	tay Tyr	gtn Val 755	ytn Leu	ytn Leu	atg Met	gay Asp	tty Phe 760	cay His	wsn Ser	gtn Val	ccn Pro	cay His 765	acn Thr	ccn Pro	gar Glu	2304
ytn Leu	ath Ile 770	ytn Leu	tay Tyr	gcn Ala	ytn Leu	gtn Val 775	tty Phe	gtn Val	ytn Leu	tty Phe	tgy Cys 780	gay Asp	gar Glu	gtn Val	mgn Arg	2352
car Gln 785	tgg Trp	tay Tyr	atg Met	aay Asn	ggn Gly 790	gtn Val	aay Asn	tay Tyr	tty Phe	acn Thr 795	gay Asp	ytn Leu	tgg Trp	aay Asn	gtn Val 800	2400
atg Met	gay Asp	acn Thr	ytn Leu	ggn Gly	ytn Leu	tty Phe	tay Tyr	tty Phe	ath Ile	gcn Ala	ggn Gly	ath Ile	gtn Val	tty Phe	mgn Arg	2448

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		th ath atg ytn car m le Ile Met Leu Gln A 860	
		rtn tty gcn gtn tgg a weu Phe Ala Val Trp M 875	
Phe Gly Val Ala A		tn mgn car aay gar c eu Arg Gln Asn Glu G 890	
	Arg Ser Val Ile T	ay gar ccn tay ytn g 'yr Glu Pro Tyr Leu A 05 9	
ggn car gtn ccn v Gly Gln Val Pro S 915	wsn gay gtn gay w Ser Asp Val Asp S 920	sn acn acn tay gay t er Thr Thr Tyr Asp P 925	ty wsn cay 2784 he Ser His
		ar ccn ytn tgy gtn g ys Pro Leu Cys Val G 940	
		ar tgg ath acn ath c lu Trp Ile Thr Ile P 955	
Cys Ile Tyr Met I		th ytn ytn gtn aay y le Leu Leu Val Asn L 970	
	Tyr Thr Val Gly I	th gtn car gar aay a le Val Gln Glu Asn A 85 9	
gtn tgg aar tty o Val Trp Lys Phe o 995	car mgn tay tty y Eln Arg Tyr Phe Lo 1000	tn gtn car gar tay t eu Val Gln Glu Tyr C 1005	gy aay mgn 3024 ys Asn Arg
ytn aay ath ccn t Leu Asn Ile Pro I 1010	tty ccn tty gtn g Phe Pro Phe Val Va 1015	tn tty gcn tay tty to al Phe Ala Tyr Phe T 1020	ay atg gtn 3072 yr Met Val
		gy aar gar aar aay a ys Lys Glu Lys Asn M 1035	
Asn Ala Cys Cys F	ty mgn aay gar ga Phe Arg Asn Glu As 1045	ay aay gar acn ytn go sp Asn Glu Thr Leu A 1050	en tgg gar 3168 la Trp Glu 1055
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1080

29/75

1075

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cct ggg ttt gcc tgg Pro Gly Phe Ala Trp 35		Ser Gly Pro Leu Pl	
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gca gca tcg gct cac Ala Ala Ser Ala His 85			
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cag gtt gaa ggg gct Gln Val Glu Gly Ala 115	aca cag gca g Thr Gln Ala G 120	gt gag cac ttg ct Ny Glu His Leu Le 12	u Ser Leu Gly
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gat gag cag gag atg Asp Glu Gln Glu Met 145	aca gct gga g Thr Ala Gly G 150	gg gta tgg gga ag ly Val Trp Gly Ar 155	a ggg ctc tgg 540 g Gly Leu Trp 160
aca gaa gaa aag atg Thr Glu Glu Lys Met 165			
aga agg aat gac act Arg Arg Asn Asp Thr 180	Leu Asp Ser T		

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wo	02/10)1045						30/	75]	PCT/EP	02/0652
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											Gly aaa					876
											ctg Leu					924
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											gcc Ala					1020
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cag Gln	tcc Ser	aaa Lys	ggt Gly 340	gct Ala	tgg Trp	att Ile	ctc Leu	acg Thr 345	gga Gly	ggc gly	acc Thr	cat His	tat Tyr 350	ggc gly	ctg Leu	1116
atg Met	aag Lys	tac Tyr 355	atc Ile	gly aaa	gag Glu	gtg Val	gtg Val 360	aga Arg	gat Asp	aac Asn	acc Thr	atc Ile 365	agc Ser	agg Arg	agt Ser	1164
											gct Ala 380					1212
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Leu Ala Gln Tyr Leu Met Asp Asp Phe Thr Arg Asp Pro Leu Tyr Ile

ctg gac aac cac aca cat ttg ctg ctc gtg gac aat ggc tgt cat

Leu Asp Asn Asn His Thr His Leu Leu Leu Val Asp Asn Gly Cys His

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	31/75	

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acg Thr	gtg Val 530	tcc Ser	cgg Arg	ctg Leu	cct Pro	gag Glu 535	gag Glu	gag Glu	act Thr	gag Glu	agt Ser 540	tgg Trp	atc Ile	aaa Lys	tgg Trp	1692
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tac Tyr	aaa Lys	gcc Ala	ttc Phe 580	agc Ser	acc Thr	agt Ser	gag Glu	caa Gln 585	gac Asp	aag Lys	gat Asp	aac Asn	tgg Trp 590	aat Asn	gly aaa	1836
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						aac Asn										2028
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ctg Leu	cag Gln	atc Ile 675	gcc Ala	aag Lys	aat Asn	tcc Ser	tat Tyr 680	aat Asn	gat Asp	gcc Ala	ctc Leu	ctc Leu 685	acg Thr	ttt Phe	gtc Val	2124
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act Thr	cgg Arg	cac His	ccc Pro	ctg Leu 725	caa Gln	gct Ala	ctc Leu	ttc Phe	atc Ile 730	tgg Trp	gcc Ala	att Ile	ctt Leu	cag Gln 735	aat Asn	2268
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tac Tyr 785	gag Glu	acc Thr	cgg Arg	gct Ala	gtt Val 790	gag Glu	ctg Leu	ttc Phe	act Thr	gag Glu 795	tgt Cys	tac Tyr	agc Ser	agc Ser	gat Asp 800	2460
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	agc Ser															2556
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	gac Asp															2748
	ccc Pro															2796
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	ccc Pro 930															2892
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ctg Leu	tgg Trp	aat Asn	gtg Val	atg Met 965	gac Asp	acg Thr	ctg Leu	gly aaa	ctt Leu 970	ttt Phe	tac Tyr	ttc Phe	ata Ile	gca Ala 975	gga Gly	2988
	gta Val															3036
cga Arg	gtc Val	att Ile 995	Phe	tgt Cys	ctg Leu	gac Asp	tac Tyr 1000	Ile	att Ile	ttc Phe	act Thr	cta Leu 1005	Arg	ttg Leu	atc Ile	3084
cac	att	ttt	act	gta	agc	aga	aac	tta	gga	ccc	aag	att	ata	atg	ctg	3132

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33//5	
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Leu	Thr	Ser 515		Ala	Val	Lys	Glu 520		Leu	Val	Arg	Phe 525		Pro	Arg
Thr	Val 530		Arg	Leu	Pro	Glu 535		Glu	Thr	Glu	Ser 540		Ile	Lys	Trp
Leu 545	Lys	Glu	Ile	Leu	Glu 550		Ser	His	Leu	Leu 555	Thr	Val	Ile	Lys	Met 560
	Glu	Ala	Gly	Asp 565		Ile	Val	Ser	Asn 570			Ser	Tyr	Ala 575	
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Gln	Leu	Lys 595		Leu	Leu	Glu	Trp 600		Gln	Leu	Asp	Leu 605		Asn	Asp
Glu	Ile 610	Phe	Thr	Asn	Asp	Arg 615	Arg	Trp	Glu	Ser	Ala 620	Asp	Leu	Gln	Glu
Val 625	Met	Phe	Thr	Ala	Leu 630	Ile	Lys	Asp	Arg	Pro 635	Lys	Phe	Val	Arg	Leu 640
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	Thr		660					665					670		
	Gln	675					680		_			685			
	Lуs 690					695		_	_		700			_	_
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	Arg			725					730					735	
	Lys		740					745					750	_	
	Ala	755					760					765			
	770					775					780				
785	Glu				790					795					800
	Asp			805					810		_			815	_
	Ser		820					825				_	830		
	Ala	835					840					845			
	Ile 850 Ile					855		_	_		860		_		
865			_		870					875					880
	Asp			885	_			_	890	_				895	
	Leu		900					905				_	910		
	Pro	915					920					925			
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945					950					955					960
_	Trp			965					970					975	
	Val Val		980					985					990		_
	Val	995					1000)				1005			
_	Ile 1010)				1015					1020				
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Val Glu Leu Asp Glu His Asn Leu Pro Arg Phe Pro Glu Trp Ile Thr
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Ile Pro Leu Val Cys Ile Tyr Met Leu Ser Thr Asn Ile Leu Leu Val
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Tyr Cys Ser Arg Leu Asn Ile Pro Phe Pro Phe Ile Val Phe Ala Tyr
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Phe Tyr Met Val Val Lys Lys Cys Phe Lys Cys Cys Cys Lys Glu Lys
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Asn Met Glu Ser Ser Val Cys Cys Phe Lys Asn Glu Asp Asn Glu Thr
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                                      1210
                                                           1215
Leu Ala Trp Glu Gly Val Met Lys Glu Asn Tyr Leu Val Lys Ile Asn
             1220
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Thr Lys Ala Asn Asp Thr Ser Glu Glu Met Arg His Arg Phe Arg Gln
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Asn Lys Ile Lys
1265
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					ytn Leu											192
gtn Val 65	tgg Trp	mgn Arg	ytn Leu	ytn Leu	mgn Arg 70	ccn Pro	tgy Cys	tay Tyr	cay His	tgy Cys 75	gtn Val	tay Tyr	tgy Cys	ggn Gly	ccn Pro 80	240
gcn Ala	gcn Ala	wsn Ser	gcn Ala	cay His 85	ytn Leu	tty Phe	ath Ile	aar Lys	car Gln 90	tgg Trp	ytn Leu	gay Asp	ggn Gly	tgg Trp 95	mgn Arg	288
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					acn Thr											384
					ccn Pro											432
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acn Thr	gar Glu	gar Glu	aar Lys	atg Met 165	wsn Ser	tty Phe	mgn Arg	gcn Ala	gcn Ala 170	mgn Arg	ytn Leu	wsn Ser	atg Met	mgn Arg 175	aay Asn	528
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					aar Lys											672
wsn Ser 225	aar Lys	gcn Ala	acn Thr	gar Glu	aay Asn 230	gtn Val	tgy Cys	aar Lys	tgy Cys	ggn Gly 235	tay Tyr	gcn Ala	car Gln	wsn Ser	car Gln 240	720
cay His	atg Met	gar Glu	ggn Gly	acn Thr 245	car Gln	ath Ile	aay Asn	car Gln	wsn Ser 250	gar Glu	aar Lys	tgg Trp	aay Asn	tay Tyr 255	aar Lys	768
aar Lys	cay His	acn Thr	aar Lys 260	gar Glu	tty Phe	ccn Pro	acn Thr	gay Asp 265	gcn Ala	tty Phe	ggn Gly	gay Asp	ath Ile 270	car Gln	tty Phe	816

gar Glu	acn Thr	ytn Leu 275	ggn Gly	aar Lys	aar Lys	ggn	aar Lys 280	tay Tyr	ath Ile	mgn Arg	ytn Leu	wsn Ser 285	Cys	gay Asp	acn Thr	864
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car Gln	wsn Ser	aar Lys	ggn Gly 340	gcn Ala	tgg Trp	ath Ile	ytn Leu	acn Thr 345	ggn Gly	ggn Gly	acn Thr	cay His	tay Tyr 350	ggn Gly	ytn Leu	1056
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wsn Ser 385	aay Asn	mgn Arg	gay Asp	acn Thr	ytn Leu 390	ath Ile	mgn Arg	aay Asn	tgy Cys	gay Asp 395	gcn Ala	gar Glu	ggn Gly	tay Tyr	tty Phe 400	1200
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ggn Gly	cay His	ccn Pro 435	acn Thr	gtn Val	gar Glu	gcn Ala	aar Lys 440	ytn Leu	mgn Arg	aay Asn	car Gln	ytn Leu 445	gar Glu	aar Lys	tay Tyr	1344
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ath Ile 465	gtn Val	tgy Cys	tty Phe	gcn Ala	car Gln 470	ggn Gly	ggn	ggn Gly	aar Lys	gar Glu 475	acn Thr	ytn Leu	aar Lys	gcn Ala	ath Ile 480	1440
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ggn Gly	car Gln	ath Ile	gcn Ala 500	gay Asp	gtn Val	ath Ile	gcn Ala	wsn Ser 505	ytn Leu	gtn Val	gar Glu	gtn Val	gar Glu 510	gay Asp	gcn Ala	1536
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					gay Asp											1872
					ytn Leu 630											1920
					ytn Leu											1968
					wsn Ser											2016
					aay Asn											2064
					aay Asn											2112
					atg Met 710											2160
					car Gln											2208
					aar Lys											2256
					gcn Ala											2304
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tay Tyr 785	gar Glu	acn Thr	mgn Arg	gcn Ala	gtn Val 790	gar Glu	ytn Leu	tty Phe	acn Thr	gar Glu 795	tgy Cys	tay Tyr	wsn Ser	wsn Ser	gay Asp 800	2400
					car Gln											2448

- -

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					wsn Ser										2976
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Arg					gtn Val)					Phe					3120
				Gly	gtn Val				Gly					Asn	3168
			Arg		ath Ile			Ser					Pro		3216
		Phe			gtn Val		Ser					Thr			3264

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15

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											gtg Val					941
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											tcc Ser					1037

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aaa Lys	gct Ala	gac Asp	atg Met	agg Arg 315	cga Arg	cag Gln	gac Asp	tcg Ser	agg Arg 320	gly ggg	aac Asn	acg Thr	gtg Val	ctg Leu 325	cac His	1133
gcg Ala	ctg Leu	gtg Val	gcc Ala 330	atc Ile	gcc Ala	gac Asp	aac Asn	acc Thr 335	cga Arg	gag Glu	aac Asn	acc Thr	aag Lys 340	ttt Phe	gtc Val	1181
acc Thr	aag Lys	atg Met 345	tac Tyr	gac Asp	ctg Leu	ctg Leu	ctt Leu 350	ctc Leu	aag Lys	tgt Cys	tca Ser	cgc Arg 355	ctc Leu	ttc Phe	ctc Leu	1229
gac Asp	agc Ser 360	aac Asn	ctg Leu	gag Glu	aca Thr	gtt Val 365	ctc Leu	aac Asn	aat Asn	gat Asp	ggc Gly 370	ctt Leu	tcg Ser	cct Pro	ctc Leu	1277
atg Met 375	atg Met	gct Ala	gcc Ala	aag Lys	aca Thr 380	gly	aag Lys	atc Ile	gly aaa	gtc Val 385	ttt Phe	cag Gln	cac His	atc Ile	atc Ile 390	1325
cga Arg	cgt Arg	gag Glu	gtg Val	aca Thr 395	gat Asp	gag Glu	gac Asp	acc Thr	cgg Arg 400	cat His	ctg Leu	tct Ser	cgc Arg	aag Lys 405	ttc Phe	1373
aag Lys	gac Asp	tgg Trp	gcc Ala 410	tat Tyr	gly aaa	cct Pro	gtg Val	tat Tyr 415	tct Ser	tct Ser	ctc Leu	tac Tyr	gac Asp 420	ctc Leu	tcc Ser	1421
tcc Ser	ctg Leu	gac Asp 425	aca Thr	tgc Cys	Gly 999	gag Glu	gag Glu 430	gtg Val	tcc Ser	gtg Val	ctg Leu	gag Glu 435	atc Ile	ctg Leu	gtg Val	1469
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ttc Phe	tac Tyr	atc Ile	Asn	gtg Val 475	⊽al	tcc Ser	Tyr	Leu	Cys	Ala	atg Met	gtc Val	atc Ile	ttc Phe 485	acc Thr	1613
ctc Leu	acc Thr	gcc Ala	tac Tyr 490	tat Tyr	cag Gln	cca Pro	ctg Leu	gag Glu 495	ggc Gly	acg Thr	cca Pro	ccc Pro	tac Tyr 500	cct Pro	tac Tyr	1661
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tta Leu	ctc Leu	tac Tyr	ttc Phe	atc Ile	tac Tyr	tct Ser	gtg Val	ctg Leu	gtg Val	gtt Val	gtc Val	tct Ser	gcg Ala	gcg Ala	ctc Leu	1853

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					aat Asn											1949
acg Thr	600 GJÀ 333	acc Thr	tac Tyr	agc Ser	atc Ile	atg Met 605	att Ile	cag Gln	aag Lys	atc Ile	ctc Leu 610	ttc Phe	aaa Lys	gac Asp	ctc Leu	1997
					gtg Val 620											2045
					ctg Leu											2093
					tgc Cys											2141
					gcc Ala											2189
ggc	atg Met 680	gga Gly	gac Asp	ctg Leu	gag Glu	atg Met 685	ctg Leu	agc Ser	agc Ser	gcc Ala	aag Lys 690	tac Tyr	ccc Pro	gtg Val	gtc Val	2237
ttc Phe 695	atc Ile	ctc Leu	ctg Leu	ctg Leu	gtc Val 700	acc Thr	tac Tyr	atc Ile	atc Ile	ctc Leu 705	acc Thr	ttc Phe	gtg Val	ctc Leu	ctg Leu 710	2285
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					atc Ile											2381
					ttc Phe											2429
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agg Arg 775	tgg Trp	tgc Cys	ttc Phe	agg Arg	gtg Val 780	gac Asp	gag Glu	gtg Val	aac Asn	tgg Trp 785	tct Ser	cac His	tgg Trp	aac Asn	cag Gln 790	2525
aac Asn	ttg Leu	ggc Gly	atc Ile	att Ile 795	aac Asn	gag Glu	gac Asp	cct Pro	ggc Gly 800	aag Lys	agt Ser	gaa Glu	atc Ile	tac Tyr 805	cag Gln	2573
					cac His											2621
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825 830 . 835

gat gaa gtg gtg gta ccc ctg gat aac cta ggg aac ccc aac tgt gac 2717
Asp Glu Val Val Val Pro Leu Asp Asn Leu Gly Asn Pro Asn Cys Asp 840

ggc cac cag cag ggc tac gct ccc aag tgg agg acg gac gat gcc cca 2765
Gly His Gln Gln Gly Tyr Ala Pro Lys Trp Arg Thr Asp Asp Ala Pro 855

ctg tag gggccgtgcc agagctcgca cagatagtcc aggcttggcc ttcgctcca 2821
Leu *

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gaacctggcc agggctgaag ctcatgcagg gacgctgcag ctccagacctg ccacagatct 3181

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 Met
 Lys
 Phe
 Gln
 Gly
 Ala
 Phe
 Arg
 Lys
 Gly
 Val
 Pro

 Asn
 Pro
 Ile
 Asp
 Leu
 Glu
 Ser
 Thr
 Leu
 Tyr
 Glu
 Ser
 Ser
 Val
 Val

 Pro
 Gly
 Pro
 Lys
 Ala
 Pro
 Met
 Asp
 Ser
 Leu
 Phe
 Asp
 Tyr
 Gly
 Thr

 Tyr
 Arg
 His
 Pro
 Ser
 Asp
 Asp
 Ser
 Leu
 Phe
 Asp
 Tyr
 Gly
 Thr

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 Asp
 Trp
 Arg
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 Lys
 Val
 Val

 Tyr
 Arg
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 Pro
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 Asp
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 Arg
 Lys
 Val
 Val

 Tyr
 Arg
 His
 Pro
 Arg
 Arg
 Trp
 Arg
 Lys
 Val
 Val

Glu Lys Gln Pro Gln Ser Pro Lys Ala Pro Ala Pro Gln Pro Pro
130 135 140

Ile Leu Lys Val Phe Asn Arg Pro Ile Leu Phe Asp Ile Val Ser Arg

Ile Leu Lys Val Phe Asn Arg Pro Ile Leu Phe Asp Ile Val Ser Arg
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Gly Ser Thr Ala Asp Leu Asp Gly Leu Leu Ser Phe Leu Leu Thr His

165 170 175
Lys Lys Arg Leu Thr Asp Glu Glu Phe Arg Glu Pro Ser Thr Gly Lys

180

Thr Cys Leu Pro Lys Ala Leu Leu Asn Leu Ser Asn Gly Arg Asn Asp

195 200 205
Thr Ile Pro Val Leu Leu Asp Ile Ala Glu Arg Thr Gly Asn Met Arg

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Glu Phe Ile Asn Ser Pro Phe Arg Asp Ile Tyr Tyr Arg Gly Gln Thr

225 230 235 240 Ser Leu His Ile Ala Ile Glu Arg Arg Cys Lys His Tyr Val Glu Leu

245 250 255 Leu Val Ala Gln Gly Ala Asp Val His Ala Gln Ala Arg Gly Arg Phe

260 265 270

Phe Gln Pro Lys Asp Glu Gly Gly Tyr Phe Tyr Phe Gly Glu Leu Pro
275 280 285

275 280 285 Leu Ser Leu Ala Ala Cys Thr Asn Gln Pro His Ile Val Asn Tyr Leu 290 295 300

Thr Glu Asn Pro His Lys Lys Ala Asp Met Arg Arg Gln Asp Ser Arg

305 Gly	Asn	Thr	Val	Leu	310 His	Ala	Leu	Val	Ala	315 Ile	Ala	Asp	Asn	Thr	320 Arg
Glu	Asn	Thr		325 Phe	Val	Thr	ГЛЗ		330 Tyr	Asp	Leu	Leu	Leu	335 Leu	Lys
Cys	Ser		340 Leu	Phe	Leu	qaA	Ser 360	345 Asn	Leu	Glu	Thr	Val 365	350 Leu	Asn	Asn
Asp	Gly 370	355 Leu	Ser	Pro	Leu	Met 375		Ala	Ala	Lys	Thr 380		Lys	Ile	Gly
Val 385	Phe	Gln	His	Ile	Ile 390		Arg	Glu	۷al	Thr 395		Glu	Asp	Thr	Arg 400
His				405					410					Tyr 415	
			420					425					430	Val	
Val	Leu	Glu 435	Ile	Leu	Val	Tyr	Asn 440	Ser	Lys	Ile	Glu	Asn 445	Arg	His	GLu
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	530					535					540			Leu -	
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			580					585					590	Tyr	
		595					600					605		Gln	
	610					615					620			Leu	
625		_			630					635				Сув	640
				645					650					Pro 655	
-			660					665					670	Leu	
		675					680					685		Ser	
	690					695					700			Ile	
705					710					715				Gly	720
				725					730					Leu 735	
_			740					745					750	Phe	
		755					760					765		Ser	
	770					775					780			Val	
785					790					795				Pro	800
-				805					810					Gly 815	
	_		820					825					830	Glu	
	_	835					840					845		Asn	
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					ccn Pro 140			432
					gay Asp			480
					tty Phe			528
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					aay Asn			624
					acn Thr 220			672
					tay Tyr			720
					cay His			768
					gcn Ala			816
					tty Phe			864
			Asn		ath Ile 300			912
					mgn Arg			960
					gcn Ala			1008
					ytn Leu			1056
					acn Thr			1104
					acn Thr 380			1152

gtn Val 385	Phe	car Gln	cay His	ath Ile	ath Ile 390	Arg	mgn Arg	gar Glu	gtn Val	acr Thr 395	Asp	gar Glu	gay ı Asp	acr Thr	n mgn Arg 400	1200
cay His	ytn Leu	wsn Ser	mgn Arg	aar Lys 405	Phe	aar Lys	gay Asp	tgg Trp	gcn Ala 410	Tyr	ggr Gly	ccr Pro	gtn Val	tay Tyr 415	wsn Ser	1248
wsn Ser	ytn Leu	tay Tyr	gay Asp 420	Leu	wsn Ser	wsn Ser	ytn Leu	gay Asp 425	Thr	tgy Cys	ggn	gar Glu	gar Glu 430	. Val	wsn Ser	1296
gtn Val	ytn Leu	gar Glu 435	ath Ile	ytn Leu	gtn Val	tay Tyr	aay Asn 440	wsn Ser	aar Lys	ath Ile	gar Glu	aay Asn 445	Arg	cay His	gar Glu	1344
atg Met	ytn Leu 450	gcn Ala	gtn Val	gar Glu	ccn Pro	ath Ile 455	aay Asn	gar Glu	ytn Leu	ytn Leu	mgn Arg 460	gay Asp	aar Lys	tgg Trp	mgn Arg	1392
aar Lys 465	tty Phe	ggn Gly	gcn Ala	gtn Val	wsn Ser 470	tty Phe	tay Tyr	ath Ile	aay Asn	gtn Val 475	gtn Val	wsn Ser	tay Tyr	ytn Leu	tgy Cys 480	1440
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gtn Val 545	gay Asp	ggn Gly	wsn Ser	tty Phe	car Gln 550	ytn Leu	ytn Leu	tay Tyr	tty Phe	ath Ile 555	tay Tyr	wsn Ser	gtn Val	ytn Leu	gtn Val 560	1680
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• •

tay Tyr	ccn Pro	gcn Ala	tgy Cys 660	mgn Arg	gay Asp	wsn Ser	gar Glu	acn Thr 665	tty Phe	wsn Ser	gcn Ala	tty Phe	ytn Leu 670	ytn Leu	gay Asp	2016
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gcn Ala	aar Lys 690	tay Tyr	ccn Pro	gtn Val	gtn Val	tty Phe 695	ath Ile	ytn Leu	ytn Leu	ytn Leu	gtn Val 700	acn Thr	tay Tyr	ath Ile	ath Ile	2112
ytn Leu 705	acn Thr	tty Phe	gtn Val	ytn Leu	ytn Leu 710	ytn Leu	aay Asn	atg Met	ytn Leu	ath Ile 715	gcn Ala	ytn Leu	atg Met	ggn Gly	gar Glu 720	2160
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tgg Trp	gcn Ala	acn Thr	acn Thr 740	ath Ile	ytn Leu	gay Asp	ath Ile	gar Glu 745	mgn Arg	wsn Ser	tty Phe	ccn Pro	gtn Val 750	tty Phe	ytn Leu	2256
Arg	Lys	Ala 755	tty Phe	Arg	Ser	Gly	Glu 760	Met	Val	Thr	Val	Gly 765	Lys	Ser	Ser	2304
gay Asp	ggn Gly 770	acn Thr	ccn Pro	gay Asp	mgn Arg	mgn Arg 775	tgg Trp	tgy Cys	tty Phe	mgn Arg	gtn Val 780	gay Asp	gar Glu	gtn Val	aay Asn	2352
tgg Trp 785	wsn Ser	cay His	tgg Trp	aay Asn	car Gln 790	aay Asn	ytn Leu	ggn ggn	ath Ile	ath Ile 795	aay Asn	gar Glu	gay Asp	ccn Pro	ggn 800	2400
aar Lys	wsn Ser	gar Glu	ath Ile	tay Tyr 805	car Gln	tay Tyr	tay Tyr	ggn Gly	tty Phe 810	wsn Ser	cay His	acn Thr	gtn Val	ggn Gly 815	mgn Arg	2448
			gay Asp 820													2496
aay Asn	aar Lys	aay Asn 835	wsn Ser	Ser	gcn Ala	Asp	Glu	Val	gtn Val	gtn Val	Pro	ytn Leu 845	Āsp	aay Asn	ytn Leu	2544
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	52/75	

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tcg Ser	ctt Leu	tat Tyr	gac Asp 420	ctc Leu	tcc Ser	tcc Ser	ctg Leu	gac Asp 425	acg Thr	tgt Cys	Gly 393	gaa Glu	gag Glu 430	gcc Ala	tcc Ser	1296
								agc Ser								1344
atg Met	ctg Leu 450	gct Ala	gtg Val	gag Glu	ccc Pro	atc Ile 455	aat Asn	gaa Glu	ctg Leu	ctg Leu	cgg Arg 460	gac Asp	aag Lys	tgg Trp	cgc Arg	1392
aag Lys 465	tt <i>c</i> Phe	gly ggg	gcc Ala	gtc Val	tcc Ser 470	ttc Phe	tac Tyr	atc Ile	aac Asn	gtg Val 475	gtc Val	tcc Ser	tac Tyr	ctg Leu	tgt Cys 480	1440
gcc Ala	atg Met	gtc Val	atc Ile	ttc Phe 485	act Thr	ctc Leu	acc Thr	gcc Ala	tac Tyr 490	tac Tyr	cag Gln	ccg Pro	ctg Leu	gag Glu 495	ggc Gly	1488
								acg Thr 505								1536
								gly aaa								1584
Ile	aaa Lys 530	gac Asp	ttg Leu	ttc Phe	atg Met	aag Lys 535	aaa Lys	tgc Cys	cct Pro	gga Gly	gtg Val 540	aat Asn	tct Ser	ctc Leu	ttc Phe	1632

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								gca Ala								1728
								ggc 585								1776
								acc Thr								1824
								ttc Phe								1872
								gtc Val								1920
								cag Gln								1968
								acc Thr 665								2016
								gly ggc								2064
								atc Ile								2112
								atg Met								2160
aca Thr	gtg Val	ggc Gly	cag Gln	gtc Val 725	tcc Ser	aag Lys	gag Glu	agc Ser	aag Lys 730	cac His	atc Ile	tgg Trp	aag Lys	ctg Leu 735	cag Gln	2208
								gag Glu 745								2256
								atg Met								2304
								tgc Cys								2352
								ggc Gly								2400
								ggc								2448

							tcc Ser								2496
							gac Asp								2544
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Æ						ccg Pro 870	ctc Leu	tag *							2616
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360 Asp Gly Leu Ser Pro Leu Met Met Ala Ala Lys Thr Gly Lys Ile Gly 370 375 Ile Phe Gln His Ile Ile Arg Arg Glu Val Thr Asp Glu Asp Thr Arg 390 395 His Leu Ser Arg Lys Phe Lys Asp Trp Ala Tyr Gly Pro Val Tyr Ser 405 410 Ser Leu Tyr Asp Leu Ser Ser Leu Asp Thr Cys Gly Glu Glu Ala Ser 420 425 430 Val Leu Glu Ile Leu Val Tyr Asn Ser Lys Ile Glu Asn Arg His Glu 440 Met Leu Ala Val Glu Pro Ile Asn Glu Leu Leu Arg Asp Lys Trp Arg 450 455 Lys Phe Gly Ala Val Ser Phe Tyr Ile Asn Val Val Ser Tyr Leu Cys 470 475 Ala Met Val Ile Phe Thr Leu Thr Ala Tyr Tyr Gln Pro Leu Glu Gly 485 490 Thr Pro Pro Tyr Pro Tyr Arg Thr Thr Val Asp Tyr Leu Arg Leu Ala 500 505 510 Gly Glu Val Ile Thr Leu Phe Thr Gly Val Leu Phe Phe Phe Thr Asn 525 520 Ile Lys Asp Leu Phe Met Lys Lys Cys Pro Gly Val Asn Ser Leu Phe 535 540 Ile Asp Gly Ser Phe Gln Leu Leu Tyr Phe Ile Tyr Ser Val Leu Val 550 555 Ile Val Ser Ala Ala Leu Tyr Leu Ala Gly Ile Glu Ala Tyr Leu Ala 565 570 Val Met Val Phe Ala Leu Val Leu Gly Trp Met Asn Ala Leu Tyr Phe 580 585 590 Thr Arg Gly Leu Lys Leu Thr Gly Thr Tyr Ser Ile Met Ile Gln Lys 595 600 Ile Leu Phe Lys Asp Leu Phe Arg Phe Leu Leu Val Tyr Leu Leu Phe 615 620 Met Ile Gly Tyr Ala Ser Ala Leu Val Ser Leu Leu Asn Pro Cys Ala 630 635 Asn Met Lys Val Cys Asn Glu Asp Gln Thr Asn Cys Thr Val Pro Thr 645 650 655 Tyr Pro Ser Cys Arg Asp Ser Glu Thr Phe Ser Thr Phe Leu Leu Asp 665 Leu Phe Lys Leu Thr Ile Gly Met Gly Asp Leu Glu Met Leu Ser Ser 675 680 685 Thr Lys Tyr Pro Val Val Phe Ile Ile Leu Leu Val Thr Tyr Ile Ile 695 700 Leu Thr Phe Val Leu Leu Asn Met Leu Ile Ala Leu Met Gly Glu 710 715 Thr Val Gly Gln Val Ser Lys Glu Ser Lys His Ile Trp Lys Leu Gln 725 730 Trp Ala Thr Thr Ile Leu Asp Ile Glu Arg Ser Phe Pro Val Phe Leu 745 Arg Lys Ala Phe Arg Ser Gly Glu Met Val Thr Val Gly Lys Ser Ser 760 765 Asp Gly Thr Pro Asp Arg Arg Trp Cys Phe Arg Val Asp Glu Val Asn 775 780 Trp Ser His Trp Asn Gln Asn Leu Gly Ile Ile Asn Glu Asp Pro Gly 790 795 Lys Asn Glu Thr Tyr Gln Tyr Tyr Gly Phe Ser His Thr Val Gly Arg
800
800
800
800
800
800 Leu Arg Arg Asp Arg Trp Ser Ser Val Val Pro Arg Val Val Glu Leu 820 825 830 Asn Lys Asn Ser Asn Pro Asp Glu Val Val Val Pro Leu Asp Ser Met 840 Gly Asn Pro Arg Cys Asp Gly His Gln Gln Gly Tyr Pro Arg Lys Trp 855 Arg Thr Glu Asp Ala Pro Leu

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1266, 1269, 1296, 1323, 1410, 1431, 1626, 1644, 1671, 1689, 1809, 1890, 1902,
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696,711,744,747,807,813,945,948,960,1008,1065,1173,1176,
1200, 1212, 1338, 1380, 1392, 1509, 1530, 1782, 1848, 1983, 2238, 2259, 2271, 2322,
2325, 2337,2448,2454,2457,2463,2484,2556,2586,2595
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Met Ala Asp Ser Ser Glu Gly Pro Arg Ala Gly Pro Gly Glu Val Ala
                                                                                                                    10
gar ytn cen ggn gay gar wsn ggn aen een ggn ggn gar gen tty een
                                                                                                                                                                                                                96
Glu Leu Pro Gly Asp Glu Ser Gly Thr Pro Gly Gly Glu Ala Phe Pro
ytn wsn wsn ytn gcn aay ytn tty gar ggn gar gay ggn wsn ytn wsn
                                                                                                                                                                                                               144
Leu Ser Ser Leu Ala Asn Leu Phe Glu Gly Glu Asp Gly Ser Leu Ser
ccn wsn ccn gcn gay gcn wsn mgn ccn gcn ggn ccn ggn gay ggn mgn
                                                                                                                                                                                                               1.92
Pro Ser Pro Ala Asp Ala Ser Arg Pro Ala Gly Pro Gly Asp Gly Arg
ccn aay ytn mgn atg aar tty car ggn gcn tty mgn aar ggn gtn ccn
                                                                                                                                                                                                               240
Pro Asn Leu Arg Met Lys Phe Gln Gly Ala Phe Arg Lys Gly Val Pro
                                                                  70
aay ccn ath gay ytn ytn gar wsn acn ytn tay gar wsn wsn gtn gtn
                                                                                                                                                                                                               288
Asn Pro Ile Asp Leu Leu Glu Ser Thr Leu Tyr Glu Ser Ser Val Val
                                                                                                                    90
ccn ggn ccn aar aar gcn ccn atg gay wsn ytn tty gay tay ggn acn Pro Gly Pro Lys Lys Ala Pro Met Asp Ser Leu Phe Asp Tyr Gly Thr
                                                                                                                                                                                                               336
tay mgn cay cay wsn wsn gay aay aar mgn tgg mgn aar aar ath ath Tyr Arg His Ser Ser Asp Asn Lys Arg Trp Arg Lys Lys Ile Ile
                                                                                                                                                                                                              384
                                                                                       120
gar aar car ccn car wsn ccn aar gcn ccn gcn ccn car ccn ccn
                                                                                                                                                                                                              432
Glu Lys Gln Pro Gln Ser Pro Lys Ala Pro Ala Pro Gln Pro Pro Pro
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135

											gay Asp					480
											tty Phe					528
aar Lys	aar Lys	mgn Arg	ytn Leu 180	acn Thr	gay Asp	gar Glu	gar Glu	tty Phe 185	mgn Arg	gar Glu	ccn Pro	wsn Ser	acn Thr 190	ggn Gly	aar Lys	576
											aay Asn					624
											acn Thr 220					672
											tay Tyr					720
											cay His					768
											gcn Ala					816
											tty Phe					864
											ath Ile 300					912
											mgn Arg					960
					His	Ala		Val	Ala	Ile	gcn Ala				Arg	1008
											ytn Leu					1056
tgy Cys	gcn Ala	mgn Arg 355	ytn Leu	tty Phe	ccn Pro	gay Asp	wsn Ser 360	aay Asn	ytn Leu	gar Glu	gcn Ala	gtn Val 365	ytn Leu	aay Asn	aay Asn	1104
gay Asp	ggn Gly 370	ytn Leu	wsn Ser	ccn Pro	ytn Leu	atg Met 375	atg Met	gcn Ala	gcn Ala	aar Lys	acn Thr 380	ggn Gly	aar Lys	ath Ile	ggn Gly	1152
ath Ile 385	tty Phe	car Gln	cay His	ath Ile	ath Ile 390	mgn Arg	mgn Arg	gar Glu	gtn Val	acn Thr 395	gay Asp	gar Glu	gay Asp	acn Thr	mgn Arg 400	1200
											ggn Gly					1248

								2	0113							
				405					410					415		
															wsn Ser	1296
gtn Val	ytn Leu	gar Glu 435	ath Ile	ytn Leu	gtn Val	tay Tyr	aay Asn 440	wsn Ser	aar Lys	ath Ile	gar Glu	aay Asn 445	mgn Arg	cay His	gar Glu	1344
							aay Asn									1392
							tay Tyr								tgy Cys 480	1440
							acn Thr									1488
acn Thr	ccn Pro	ccn Pro	tay Tyr 500	ccn Pro	tay Tyr	mgn Arg	acn Thr	acn Thr 505	gtn Val	gay Asp	tay Tyr	ytn Leu	mgn Arg 510	ytn Leu	gcn Ala	1536
							acn Thr 520									1584
							aar Lys									1632
							ytn Leu									1680
ath Ile	gtn Val	wsn Ser	gcn Ala	gcn Ala 565	ytn Leu	tay Tyr	ytn Leu	gcn Ala	ggn Gly 570	ath Ile	gar Glu	gcn Ala	tay Tyr	ytn Leu 575	gcn Ala	1728
gtn Val	atg Met	gtn Val	tty Phe 580	gcn Ala	ytn Leu	gtn Val	ytn Leu	ggn Gly 585	tgg Trp	atg Met	aay Asn	gcn Ala	ytn Leu 590	tay Tyr	tty Phe	1776
acn Thr	mgn Arg	ggn Gly 595	ytn Leu	aar Lys	ytn Leu	acn Thr	ggn Gly 600	acn Thr	tay Tyr	wsn Ser	ath Ile	atg Met 605	ath Ile	car Gln	aar Lys	1824
							mgn Arg									1872
							ytn Leu									1920
							gay Asp									1968
tay Tyr	ccn Pro	wsn Ser	tgy Cys 660	mgn Arg	gay Asp	wsn Ser	gar Glu	acn Thr 665	tty Phe	wsn Ser	acn Thr	tty Phe	ytn Leu 670	ytn Leu	gay Asp	2016
ytn Leu	tty Phe	aar Lys	ytn Leu	acn Thr	ath Ile	ggn Gly	atg Met	ggn Gly	gay Asp	ytn Leu	gar Glu	atg Met	ytn Leu	wsn Ser	wsn Ser	2064

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675		680	685	
		Ile Ile Leu	ytn gtn acn tay Leu Val Thr Tyr 700	
			ath gcn ytn atg Ile Ala Leu Met 715	
			cay ath tgg aar His Ile Trp Lys	
Trp Ala Thr			wsn tty ccn gtn Ser Phe Pro Val 750	
			acn gtn ggn aar Thr Val Gly Lys 765	
			mgn gtn gay gar Arg Val Asp Glu 780	
tgg wsn cay Trp Ser His ' 785	tgg aay car aay Trp Asn Gln Asn 790	ytn ggn ath Leu Gly Ile	ath aay gar gay Ile Asn Glu Asp 795	ccn ggn 2400 Pro Gly 800
			wsn cay acn gtn Ser His Thr Val	
Leu Arg Arg			ccn mgn gtn gtn Pro Arg Val Val 830	
			gtn ccn ytn gay Val Pro Leu Asp 845	
			ggn tay ccn mgn Gly Tyr Pro Arg 860	
	gay gcn ccn ytn Asp Ala Pro Leu 870			2613
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	ucleotide probe encoding nucleio		zes to mouse	
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WO 02/101045	60/75	PCT/EP02/06520
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	63/75	
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23

26

23

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